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UNITED STATES PATENT APPLICATION OF

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FOR

NOVEL GAMMA-LACTAMS AS BETA-SECRETASE INHIBITORS

NOVEL GAMMA-LACTAMS AS BETA-SECRETASE INHIBITORS

CROSS-REFERENCE TO RELATED APPLICATIONS

This is a non-provisional application which
claims the benefit of U.S. Provisional Application
Number 60/401,390 filed August 5, 2002.

FIELD OF THE INVENTION

This invention provides novel substituted gamma-
lactam compounds having drug and bio-affecting
properties, their pharmaceutical compositions and
method of use. In particular, the invention is
concerned a series of novel gamma-lactams which are
inhibitors of the β-amyloid peptide (β-AP) production,
thereby acting to prevent the accumulation of amyloid
protein deposits in the brain and, therefore, are
useful in the treatment of neurological disorders
related to β-amyloid production. More particularly,
the present invention relates to the treatment of
Alzheimer's Disease (AD) and similar diseases.

BACKGROUND OF THE INVENTION

Alzheimer's Disease is a progressive,
neurodegenerative disorder characterized by memory
impairment and cognitive dysfunction. AD is
characterized pathologically by the accumulation of
senile (neuritic) plaques, neurofibrillary tangles,
amyloid deposition in neural tissues and vessels,
synaptic loss, and neuronal death. It is the most
common form of dementia and it now represents the
third leading cause of death after cardiovascular
disorders and cancer. The cost of Alzheimer's Disease
is enormous (in the U.S., greater than $100 billion

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annually) and includes the suffering of the patients, the suffering of families, and the lost productivity of patients and caregivers. As the longevity of society increases, the occurrence of AD will markedly

5   increase. It is estimated that more than 10 million Americans will suffer from AD by the year 2020, if methods for prevention and treatment are not found. Currently, AD is estimated to afflict 10% of the population over age 65 and up to 50% of those over the

10   age of 85. No treatment that effectively prevents AD or reverses the clinical symptoms and underlying pathophysiology is currently available (for review see Selkoe, D.J. *Ann. Rev. Cell Biol.*, <u>1994</u>, **10**: 373-403).

There have been many theories relating to the

15   etiology and pathogenesis of AD. These theories were either based on analogies with other diseases and conditions (e.g., slow virus and aluminum theories), or based on pathologic observations (e.g., cholinergic, amyloid, or tangle theories). Genetic

20   analysis can potentially differentiate between competing theories. The identification of mutations in the β-amyloid precursor protein (β-APP) of individuals prone to early onset forms of AD and related disorders strongly supports the amyloidogenic

25   theories.

Histopathological examination of brain tissue derived upon autopsy or from neurosurgical specimens in affected individuals reveals the occurrence of amyloid plaques and neurofibrillar tangles in the

30   cerebral cortex of such patients. Similar alterations are observed in patients with Trisomy 21 (Down's syndrome). Biochemical and immunological studies reveal that the dominant proteinaceous component of

the amyloid plaque is an approximately 4.2 kilodalton (kD) protein of about 39 to 43 amino acids. This protein is designated Aβ, β-amyloid peptide, and sometimes β/A4; referred to herein as Aβ. In addition

5 to its deposition in amyloid plaques, Aβ is also found in the walls of meningeal and parenchymal arterioles, small arteries, capillaries, and sometimes, venules. Compelling evidence accumulated during the last decade reveals that Aβ is an internal polypeptide derived

10 from a type 1 integral membrane protein, termed β-amyloid precursor protein (APP) (Selkoe, D. *Physiol. Rev.* 2001, **81,** 741-766; Wolfe, M. *J. Med. Chem.* 2001, **44,** 2039-2060). βAPP is normally produced by many cells both in vivo and in cultured cells, derived from

15 various animals and humans. Several proteolytic fragments of APP are generated by proteinases referred to as secretases. A subset of these proteolytic fragments, designated β-amyloid peptide (Aβ), contains 39 to 43 amino acids and is generated

20 by the combined action of β-secretase and γ-secretase. β-secretase is a membrane-bound, aspartyl protease that forms the N-terminus of the Aβ peptide. The C-terminus of the Aβ peptide is formed by γ-secretase, an apparently oligomeric complex that includes

25 presenilin-1 and/or presenilin-2. Presenilin-1 and presenilin-2 are polytopic membrane-spanning proteins that may contain the catalytic components of γ-secretase (Seiffert, D.; Bradley, J. *et al., J. Biol. Chem.* 2000, **275,** 34086-34091).

30 Multiple lines of evidence together strongly suggest that a reduction in brain Aβ levels will

prevent the onset and progression of AD. First, Aβ is
a major constituent of the parenchemyal plaques
observed in all AD patients and the cerebral
vasculature amyloid deposits observed in 90% AD
5   patients (reviewed in Selkoe, D. *Physiol. Rev.* 2001,
81, 741-766; Wolfe, M. *J. Med. Chem.* 2001, **44,** 2039-
2060). These plaques are formed from the aggregation
of soluble Aβ whose brain levels are highly correlated
with the severity of AD neurodegeneration (McLean, C.,
10   Cherny, R. *et al., Ann. Neurol.* 1999, **46,** 860-866).
Second, mutations in three genes (APP, PS-1, or PS-2)
that increase Aβ cause familial AD (FAD), where AD
onset is accelerated by at least a decade. Included
in the mutations that increase Aβ are chromosome 21
15   Trisomy that causes Down's syndrome. Third,
transgenic mice that express one or more of the mutant
FAD genes have increased Aβ levels, form parenchymal
plaques and cerebral vascular deposits containing Aβ,
exhibit memory deficits (Chapman, P.; White, G. *et*
20   *al., Nature Neurosci.* 1999, **2,** 271-276) and enhance
neurofibrillary degeneration in mice that also
overexpress mutant tau (Lewis, J.; Dickson, D. *et al.,*
Science 2001, **293,** 1487-1491). Fourth, Aβ is toxic to
cultured cells (Dahlgren, K.; Manelli, A. *et al., J.*
25   *Biol. Chem.* 2002 **277**, 32046-32053), induces
neurofibrillary tangles in mice with mutant tau (Gotz,
J., Chen, F. *et al., Science* 2001, **293,** 1491-1495) and
interferes with long-term potentiation, a likely
component of memory (Walsh, D., Klyubin, I. *et al.,*
30   *Nature* 2002, **416,** 535-539 and references therein).
Taken together, these data lead one skilled in the art
to conclude that excess Aβ production and/or reduced

Aβ clearance cause AD. From this it follows that reducing brain Aβ levels by inhibition of γ-secretase will prevent the onset and progression of AD.

In addition to AD, excess production and/or reduced clearance of Aβ causes cerebral amyloid angiopathy (CAA) (reviewed in Thal, D., Gherbremedhin, E. *et al., J. Neuropath. Exp. Neuro.* 2002, **61,** 282-293). In these patients, vascular amyloid deposits cause degeneration of vessel walls and aneurysms that may be responsible for 10-15% hemorrhagic strokes in elderly patients. As in AD, mutations in the gene encoding Aβ lead to an early onset form of CAA, referred to as cerebral hemorrhage with amyloidosis of the Dutch type, and mice expressing this mutant protein develop CAA that is similar to patients.

A logical approach to reducing Aβ levels is to interfere with the action of the secretases that are directly involved in the cleavage of APP to Aβ. The β-secretase enzyme (BACE) is responsible for cleaving APP and forms the amino-terminus of Aβ, initiating the amyloidogenic pathway. The BACE enzyme is a transmembrane aspartyl protease and was described in the literature by several independent groups [see Hussain, I. *et al.,* (1999) *Mol. Cell. Neurosci.,* **14:** 419-427; Lin, X. *et al.,*(2000) *Proceedings of the National Academy of Sciences of the United States of America,* **97:** 1456-1460; Sinha, S., *et al.,* (1999) *Nature (London),* **402:** 537-540; Vassar, R., *et al.,* (1999) *Science (Washington, D. C.),* **286:** 735-741; Walsh, D.M. *et al.,* (2002); Wolfe, M.S. (2001); Yan, R. *et al.,* (1999) *Nature (London),* **402:** 533-537].

Removal of BACE activity in mice by gene targeting completely abolishes Aβ production [see Luo, Y., *et al.*, (2001) *Nature Neuroscience*, **4**: 231-232; Roberds, S.L. *et al.*,(2001) *Human Molecular Genetics*, **10**: 1317-1324].

BACE -/- mice also show no detectable negative phenotypes, suggesting that disruption of BACE-mediated cleavage of APP does not produce additional undesired effects. This demonstrates that a drug substance capable of inhibiting β-secretase activity should lower or halt the synthesis of Aβ and should provide a safe treatment for Alzheimer's disease.

Published article Martin, J.L. *et al.*, (1999), *Biochemistry*, **38**: 7978-7988 discloses macrocyclic inhibitors of the HIV 1 protease.

PCT Publication WO 96/16950,published June 6, 1996, discloses macrocyclic inhibitors of the HIV 1 protease.

PCT Publication WO 01/07407, published February 1, 2001, discloses lactam inhibitors of the hepatitis C virus NS3 protease.

PCT Publication WO 97/16425, published May 9, 1997, and related U.S. Patent 5,719,296 disclose pseudolactam inhibitors of peptide binding to MHC class II receptors.

U.S. Patent 5,120,718 to Goldman *et al.*, granted June 9, 1992, discloses candida acid protease inhibiting compounds.

PCT Publication WO 90/04917, published May 17, 1990, and related U.S. Patent 5,164,388 discloses heterocyclic peptide renin inhibitors.

PCT Publication WO 87/05909, published October 8, 1987, and related U.S. Patent 4,705,846 disclose renin inhibitors having a lactam pseudo dipeptide insert.

Published article Thaisrivongs *et al.*, *J.*
5   *Hypertension* (1989), Suppl. **(2)**, S21-S23 discusses related renin inhibitors.

Published article Thaisrivongs, S. *et al.*, *J. Med. Chem.* (1988), **31(7)**: 1369-76 discusses related renin inhibitors.
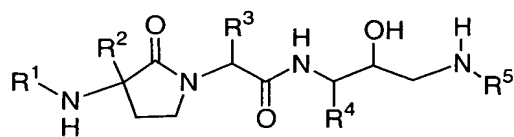
10   U.S. Patent 5,164,388 to De *et al.*, granted November 17, 1992, discloses heterocyclic renin inhibitors.

At present there remains an urgent need to develop pharmaceutical agents capable for effective
15   treatment in halting, slowing, preventing, and/or reversing the progression of Alzheimer's disease. Compounds that are effective inhibitors of beta-secretase, that inhibit beta-secretase mediated cleavage of APP, that are effective inhibitors of Aβ
20   protein production by beta-secretase, and/or are effective in reducing soluble Aβ protein, amyloid beta deposits or amyloid beta plaques, are needed for effective treatment in halting, slowing, preventing, and/or reversing neurological disorders related to Aβ
25   protein production, such as Alzheimer's disease.

SUMMARY OF THE INVENTION

A series of gamma-lactam derivatives having the Formula (I)

$$R^1 \diagdown \underset{\underset{H}{N}}{} \overset{R^2 \enspace O}{\underset{}{\diagup}} \underset{}{N} \underset{\underset{O}{\parallel}}{\overset{R^3 \enspace H}{\diagup}} \underset{}{N} \underset{\underset{R^4}{}}{} \overset{OH \enspace H}{\diagup} \underset{}{N} R^5$$

(I)

or a stereoisomer; or a pharmaceutically acceptable
salt thereof, wherein R^1, R^2, R^3, R^4, and R^5 as
defined below are effective inhibitors of the
production of β-amyloid peptide (β-AP) from β-amyloid
precursor protein (β-APP). The pharmacologic action
of these compounds makes them useful for treating
conditions responsive to the inhibition of β-AP in a
patient; e.g., Alzheimer's Disease (AD) and Down's
Syndrome. Therapy utilizing administration of these
compounds or a pharmaceutical composition containing a
therapeutically effective amount of at least one of
these compounds to patients suffering from, or
susceptible to, these conditions involves reducing β-
AP available for accumulation and deposition in brains
of these patients.

DETAILED DESCRIPTION OF THE INVENTION

The present invention comprises compounds of
Formula I, their pharmaceutical formulations, and
their use in inhibiting β-AP production in patients
suffering from or susceptible to AD or other disorders
resulting from β-AP accumulation in brain tissue. The
compounds of Formula I which include stereoisomers and
pharmaceutically acceptable salts thereof have the
following formula and meanings:

(I)
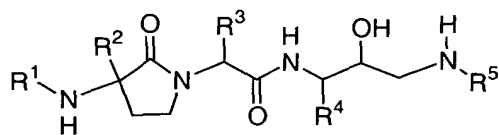
wherein

R^1 is selected from the group consisting of

-C(=O)R^{1a}, -S(=O)R^{1a}, -S(=O)$_2$R^{1a}, -C(=O)OR^{1a},

-C(=O)NHR^{1a}, and C$_1$-C$_6$ alkyl optionally

substituted with R^{1b};

R^{1a} is C$_1$-C$_6$ alkyl optionally substituted with R^{1b};

R^{1b} is independently selected from the group consisting

of halogen, -CF$_3$, -OCF$_3$, -CO$_2$R^6, -C(=O)NR^{6}R^6,

-NR^{6}C(=O)R^6, -NR^{6}R^6, -NR^{6}SO$_2$R^6, -C(=O)R^6,

-S(=O)R^6, -SO$_2$R^6, -SO$_2$NR^{6}R^6, -SR^6, -S(C$_1$-C$_4$

haloalkyl), -OR^6, -O(C$_1$-C$_4$ haloalkyl),

-(C$_3$-C$_7$)cycloalkyl, -imidazole, -thiazole,

-oxazole, -(C$_2$-C$_6$)alkenyl, and -(C$_2$-C$_6$)alkynyl;

R^2 is selected from the group consisting of

C$_1$-C$_4$ alkyl, C$_2$-C$_4$ alkenyl, C$_2$-C$_4$ alkynyl, and

C$_3$-C$_6$ cycloalkyl in which each group is optionally

substituted with halogen, -CF$_3$, -OCF$_3$, -CH$_3$,

-CH$_2$CH$_3$, -OCH$_3$, -OCH$_2$CH$_3$, or -(C$_3$-C$_7$)cycloalkyl;

R^3 is selected from the group consisting of

C_1-C_4 alkyl, C_2-C_4 alkenyl, and C_2-C_4 alkynyl

optionally substituted with R^{3a}, or phenyl

optionally substituted with R^{3b};

5   R^{3a} is selected from the group consisting of R^{3b}, C_3-C_6

cycloalkyl optionally substituted with R^{3b}, phenyl

optionally substituted with R^{3b}, and

3,4-methylenedioxyphenyl;

10   R^{3b} is independently selected at each occurrence from

the group consisting of halogen, $-NO_2$, $-CN$,

$-C_1$-C_4alkyl, $-OH$, $-OCH_3$, $-OCH_2CH_3$, $-CF_3$, $-OCF_3$,

$-SCF_3$, $-C(=O)R^6$, $-NR^6C(=O)R^6$, $-NR^6SO_2R^6$, $-NR^6R^6$,

$-OC(=O)NR^6R^6$, $-NR^6C(=O)NR^6R^6$, $-C(=O)NR^6R^6$,

15   $-C(=O)OR^6$, $-SR^6$, $-S(=O)R^6$, $-S(=O)_2R^6$, and

$-S(=O)_2NR^6R^6$;

R^4 is selected from the group consisting of C_1-C_4

alkyl, C_2-C_4 alkenyl,and C_2-C_4 alkynyl optionally

20   substituted with R^{4a};

R^{4a} is selected from R^{4b}, or phenyl optionally

substituted with R^{4b};

25   R^{4b} is selected from the group consisting of halogen,

$-NO_2$, $-CN$, $-NCS$, $-CH_3$, $-CH_2CH_3$, $-CH_2CH_2CH_3$,

$-CH(CH_3)_2$, $-CF_3$, $-OCF_3$, $-SCF_3$, $-OH$, $-OCH_3$,

$-OCH_2CH_3$, $-SH$, $-SCH_3$, $-SCH_2CH_3$, $-CO_2H$, $-CO_2CH_3$,

$-CO_2CH_2CH_3$, $-NH_2$, $-NH(CH_3)$, $-N(CH_3)_2$, $-C(=O)NH_2$,

30   $-C(=O)NH(CH_3)$, $-C(=O)N(CH_3)_2$, $-C(=O)H$, $-C(=O)CH_3$,

$-NHC(=O)CH_3$, and $-NHSO_2CH_3$;

R^5 is C_1-C_{10} alkyl optionally substituted with R^{5a};

R^{5a} is selected from the group consisting of R^{5b},
    C_3-C_8 cycloalkyl, C_2-C_6 alkenyl, C_2-C_6 alkynyl, and
    phenyl optionally substituted with R^{5b};

R^{5b} is selected from the group consisting of R^6,
    halogen, $-CN$, $-CF_3$, $-NO_2$, $-NCS$, $-OCF_3$, $-CO_2H$,
    $-C(=O)H$, $-OR^6$, $-NR^6R^6$, $-OC(=O)NR^6R^6$,
    $-NR^6C(=O)NR^6R^6$, $-C(=O)NR^6R^6$, $-C(=O)OR^6$, $-SR^6$,
    $-S(=O)R^6$, $-S(=O)_2R^6$, and $-S(=O)_2NR^6R^6$; and

R^6 is independently selected at each occurrence from
    the group consisting of hydrogen, C_1-C_6 alkyl and
    phenyl.

The present invention also provides a method for
the treatment or alleviation of disorders associated
with β-amyloid peptide, especially Alzheimer's
Disease, cerebral amyloid angiopathy and Down's
Syndrome, which comprises administering together with
a conventional adjuvant, carrier or diluent a
therapeutically effective amount of a compound of
Formula (I) or a pharmaceutically acceptable salt
thereof.

As used herein, the term "Aβ" denotes the protein
designated Aβ, β-amyloid peptide, and sometimes β/A4,
in the art. Aβ is an approximately 4.2 kilodalton
(kD) protein of about 39 to 43 amino acids found in
amyloid plaques, the walls of meningeal and

parenchymal arterioles, small arteries, capillaries, and sometimes, venules. The isolation and sequence data for the first 28 amino acids are described in U.S. Patent 4,666,829. The 43 amino acid sequence is

5   well known in the art, see Colin Dingwall, *Journal of Clinical Investigation*, Nov. 2001, **108** (9): 1243-1246; as well as PCT international patent application WO 01/92235, filed December 6, 2001, herein incorporated by reference in its entirety.

10   The term "APP", as used herein, refers to the protein known in the art as β amyloid precursor protein. This protein is the precursor for Aβ and through the activity of "secretase" enzymes, as used herein, it is processed into Aβ. Differing secretase

15   enzymes, known in the art, have been designated β secretase, generating the N-terminus of Aβ, α secretase cleaving around the 16/17 peptide bond in Aβ, and "γ secretases", as used herein, generating C-terminal Aβ fragments ending at position 38, 39, 40,

20   42, and 43 or generating C-terminal extended precursors which are subsequently truncated to the above polypeptides.

     The term "substituted," as used herein and in the claims, means that any one or more hydrogens on the

25   designated atom is replaced with a selection from the indicated group, provided that the designated atom's normal valency is not exceeded, and that the substitution results in a stable compound.

     As used herein and in the claims, "alkyl" or

30   "alkylene" is intended to include both branched and straight-chain saturated aliphatic hydrocarbon groups having the specified number of carbon atoms; for

example, "C_1-C_6 alkyl" and "C_1-C_{10} alkyl" denotes alkyl

having 1 to 6 or 1 to 10 carbon atoms. Examples of

alkyl include, but are not limited to, methyl, ethyl,

n-propyl, i-propyl, n-butyl, i-butyl, sec-butyl,

5   t-butyl, pentyl, hexyl, octyl and decyl. Preferred

"alkyl" group, unless otherwise specified, is "C_1-C_4

alkyl". Additionally, unless otherwise specified,

"propyl" denotes n-propyl or i-propyl; "butyl" denotes

n-butyl, i-butyl, sec-butyl, or t-butyl.

10   As used herein and in the claims, "alkenyl" or

"alkenylene" is intended to include hydrocarbon chains

of either a straight or branched configuration and one

or more unsaturated carbon-carbon bonds which may

occur in any stable point along the chain. Examples

15   of "C_2-C_6 alkenyl" include, but are not limited to,

ethenyl, 1-propenyl, 2-propenyl, 1-butenyl, 2-butenyl,

3-butenyl, 3-methyl-2-butenyl, 2-pentenyl, 3-pentenyl,

hexenyl, and the like.

        As used herein and in the claims, "alkynyl" or

20   "alkynylene" is intended to include hydrocarbon chains

of either a straight or branched configuration and one

or more carbon-carbon triple bonds which may occur in

any stable point along the chain, such as ethynyl, 1-

propynyl, 2-propynyl, 1-butynyl, 2-butynyl, 3-butynyl,

25   and the like.

        "Alkoxy" or "alkyloxy" represents an alkyl group

as defined above with the indicated number of carbon

atoms attached through an oxygen bridge. Examples of

alkoxy include, but are not limited to, methoxy,

30   ethoxy, n-propoxy, i-propoxy, n-butoxy, s-butoxy,

t-butoxy, n-pentoxy, and s-pentoxy. Preferred alkoxy

groups are methoxy, ethoxy, n-propoxy, i-propoxy,

n-butoxy, s-butoxy, t-butoxy. Similarly, "alkylthio"

or "thioalkoxy" is represents an alkyl group as
defined above with the indicated number of carbon
atoms attached through a sulphur bridge.

As used herein and in the claims, "halogen"
refers to fluoro, chloro, bromo, and iodo. Unless
otherwise specified, preferred halogens are fluoro and
chloro. "Counterion" is used to represent a small,
negatively charged species such as chloride, bromide,
hydroxide, acetate, sulfate, and the like.

"Cycloalkyl" is intended to include saturated
ring groups, having the specified number of carbon
atoms. For example, "C_3-C_6 cycloalkyl" and "C_3-C_8
cycloalkyl" denotes such as cyclopropyl, cyclobutyl,
cyclopentyl, cyclohexyl, or cyclooctyl.

The compounds described herein may have
asymmetric centers. An example of a preferred
stereochemical configuration is the isomer:
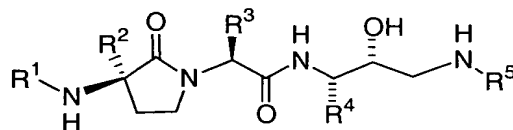
(Ia)

or pharmaceutically acceptable salt thereof, but is
not intended to be limited to this example. It is
understood, that whether a chiral center in an isomer
is "R" or "S" depends on the chemical nature of the
substituents of the chiral center. All configurations
of compounds of the invention are considered part of
the invention. Additionally, the carbon atom to which
R^1NH- and R^2 is attached may describe a chiral carbon.
Compounds of the present invention containing an
asymmetrically substituted atom may be isolated in
optically active or racemic forms. It is well known

in the art how to prepare optically active forms, such
as by resolution of racemic forms or by synthesis from
optically active starting materials. Mixtures of
isomers of the compounds of the examples or chiral
5   precursors thereof can be separated into individual
isomers according to methods which are known per se,
e.g. fractional crystallization, adsorption
chromatography or other suitable separation processes.
Resulting racemates can be separated into antipodes in
10   the usual manner after introduction of suitable salt-
forming groupings, e.g. by forming a mixture of
diastereosiomeric salts with optically active salt-
forming agents, separating the mixture into
diastereomeric salts and converting the separated
15   salts into the free compounds. The enantiomeric forms
may also be separated by fractionation through chiral
high pressure liquid chromatography columns. Many
geometric isomers of olefins and the like can also be
present in the compounds described herein, and all
20   such stable isomers are contemplated in the present
invention. Cis and trans geometric isomers of the
compounds of the present invention are described and
may be isolated as a mixture of isomers or as
separated isomeric forms. All chiral, diastereomeric,
25   racemic forms and all geometric isomeric forms of a
structure are intended, unless the specific
stereochemistry or isomeric form is specifically
indicated.

The phrase "pharmaceutically acceptable" is
30   employed herein to refer to those compounds,
materials, compositions, and/or dosage forms which
are, within the scope of sound medical judgment,
suitable for use in contact with the tissues of human
beings and animals without excessive toxicity,
35   irritation, allergic response, or other problem or

complication, commensurate with a reasonable benefit/risk ratio.

As used herein and in the claims, "pharmaceutically acceptable salts" refer to
5 derivatives of the disclosed compounds wherein the parent compound is modified by making acid or base salts thereof. Examples of pharmaceutically acceptable salts include, but are not limited to, mineral or organic acid salts of basic residues such
10 as amines; alkali or organic salts of acidic residues such as carboxylic acids; and the like. The pharmaceutically acceptable salts include the conventional non-toxic salts or the quaternary ammonium salts of the parent compound formed, for
15 example, from non-toxic inorganic or organic acids. For example, such conventional non-toxic salts include those derived from inorganic acids such as hydrochloric, hydrobromic, sulfuric, sulfamic, phosphoric, nitric and the like; and the salts
20 prepared from organic acids such as acetic, propionic, succinic, glycolic, stearic, lactic, malic, tartaric, citric, ascorbic, pamoic, maleic, hydroxymaleic, phenylacetic, glutamic, benzoic, salicylic, sulfanilic, 2-acetoxybenzoic, fumaric,
25 toluenesulfonic, methanesulfonic, ethane disulfonic, oxalic, isethionic, and the like.

The pharmaceutically acceptable salts of the present invention can be synthesized from the parent compound which contains a basic or acidic moiety by
30 conventional chemical methods. Generally, such salts can be prepared by reacting the free acid or base forms of these compounds with a stoichiometric amount of the appropriate base or acid in water or in an

organic solvent, or in a mixture of the two;
generally, nonaqueous media like ether, ethyl acetate,
ethanol, isopropanol, or acetonitrile are preferred.
Lists of suitable salts are found in *Remington's*

5   *Pharmaceutical Sciences*, 17th ed., Mack Publishing
Company, Easton, PA, 1985, p. 1418, the disclosure of
which is hereby incorporated by reference.

In the method of the present invention, the term
"therapeutically effective amount" means the total

10   amount of each active component of the method that is
sufficient to show a meaningful patient benefit, i.e.,
healing of acute conditions characterized by
inhibition of β-amyloid peptide production. When
applied to an individual active ingredient,

15   administered alone, the term refers to that ingredient
alone. When applied to a combination, the term refers
to combined amounts of the active ingredients that
result in the therapeutic effect, whether administered
in combination, serially or simultaneously. The terms

20   "treat, treating, treatment" as used herein and in the
claims means preventing or ameliorating diseases
associated with β-amyloid peptide.

In a preferred embodiment, the present invention
provides for compounds of Formula (I)

25

(I)
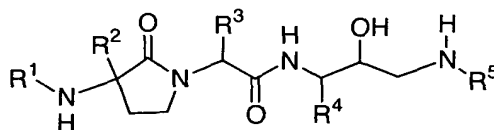
or a stereoisomer; or a pharmaceutically acceptable

30   salt thereof, wherein

R^1 is selected from the group consisting of -C(=O)R^{1a},
-S(=O)R^{1a}, -S(=O)$_2R^{1a}$, -C(=O)OR^{1a}, and -C(=O)NHR^{1a};

R^{1a} is C_1-C_6 alkyl optionally substituted with R^{1b};

5

R^{1b} is independently selected from the group consisting
of halogen, -CF$_3$, -OCF$_3$, -CO$_2R^6$, -C(=O)NR^6R^6,
-NR^6C(=O)R^6, -NR^6R^6, -OR^6, -(C3-C7)cycloalkyl,
-imidazole, -thiazole, -oxazole, -(C$_2$-C$_6$)alkenyl,

10   and -(C$_2$-C$_6$)alkynyl;

R^2 is selected from the group consisting of
C_1-C_4 alkyl, C_2-C_4 alkenyl, C_2-C_4 alkynyl, and
C_3-C_6 cycloalkyl in which each group is optionally

15   substituted with halogen, -CF$_3$, -OCF$_3$, -CH$_3$,
-CH$_2$CH$_3$, -OCH$_3$, -OCH$_2$CH$_3$, or C_3-C_7 cycloalkyl;

R^3 is C_1-C_4 alkyl optionally substituted with R^{3a};

20   R^{3a} is selected from the group consisting of R^{3b},
C_3-C_6 cycloalkyl optionally substituted with R^{3b},
phenyl optionally substituted with R^{3b}, and
3,4-methylenedioxyphenyl;

25   R^{3b} is independently selected at each occurrence from
the group consisting of halogen, -NO$_2$, -CN,
-C$_1$-C$_4$alkyl, -OH, -OCH$_3$, -OCH$_2$CH$_3$, -CF$_3$, -OCF$_3$,
-SCF$_3$, -C(=O)R^6, -NR^6C(=O)R^6, -NR^6SO$_2R^6$, -NR^6R^6,
-OC(=O)NR^6R^6, -NR^6C(=O)NR^6R^6, -C(=O)NR^6R^6,

30   -C(=O)OR^6, -SR^6, -S(=O)R^6, -S(=O)$_2R^6$, and
-S(=O)$_2$NR^6R^6;

R^4 is C_1-C_4 alkyl optionally substituted with R^{4a};

R^{4a} is R^{4b} or phenyl optionally substituted with R^{4b};

R^{4b} is selected from the group consisting of halogen, $-NO_2$, $-CN$, $-NCS$, $-CH_3$, $-CH_2CH_3$, $-CH_2CH_2CH_3$, $-CH(CH_3)_2$, $-CF_3$, $-OCF_3$, $-SCF_3$, $-OH$, $-OCH_3$, $-OCH_2CH_3$, $-SH$, $-SCH_3$, $-SCH_2CH_3$, $-CO_2H$, $-CO_2CH_3$, $-CO_2CH_2CH_3$, $-NH_2$, $-NH(CH_3)$, $-N(CH_3)_2$, $-C(=O)NH_2$, $-C(=O)NH(CH_3)$, $-C(=O)N(CH_3)_2$, $-C(=O)H$, $-C(=O)CH_3$, $-NHC(=O)CH_3$, and $-NHSO_2CH_3$;

R^5 is C_1-C_{10} alkyl optionally substituted with R^{5a};

R^{5a} is selected from the group consisting of R^{5b}, C_3-C_8 cycloalkyl, C_2-C_6 alkenyl, C_2-C_6 alkynyl optionally substituted with R^{5b}, and phenyl optionally substituted with R^{5b};

R^{5b} is selected from the group consisting of R^6, halogen, $-CN$, $-CF_3$, $-NO_2$, $-NCS$, $-OCF_3$, $-CO_2H$, $-C(=O)H$, $-OR^6$, $-NR^6R^6$, $-OC(=O)NR^6R^6$, $-NR^6C(=O)NR^6R^6$, $-C(=O)NR^6R^6$, $-C(=O)OR^6$, $-SR^6$, $-S(=O)R^6$, $-S(=O)_2R^6$, and $-S(=O)_2NR^6R^6$; and
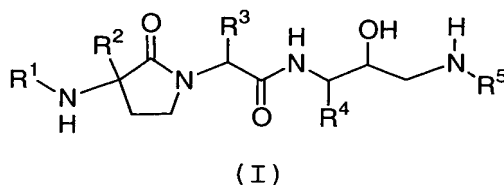
R^6 is independently selected at each occurrence from the group consisting of hydrogen, C_1-C_6 alkyl and phenyl.

In another preferred embodiment, the present invention provides compounds of Formula (I)

(I)

or a stereoisomer; or a pharmaceutically acceptable salt thereof, wherein

R^1 is selected from the group consisting of $-C(=O)R^{1a}$, $-S(=O)R^{1a}$, $-S(=O)_2R^{1a}$, $-C(=O)OR^{1a}$, and $-C(=O)NHR^{1a}$;

R^{1a} is C_1-C_6 alkyl optionally substituted with R^{1b};

R^{1b} is independently selected from the group consisting of halogen, $-CF_3$, $-OCF_3$, $-CO_2R^6$, $-C(=O)NR^6R^6$, $-NR^6C(=O)R^6$, $-NR^6R^6$, $-OR^6$, $-(C3-C7)$cycloalkyl, $-$imidazole, $-$thiazole, $-$oxazole, $-(C_2-C_6)$alkenyl, and $-C_2-C_6)$alkynyl;

R^2 is selected from the group consisting of C_1-C_4 alkyl, C_2-C_4 alkenyl, C_2-C_4 alkynyl, and C_3-C_6 cycloalkyl in which each group is optionally substituted with halogen, $-CF_3$, $-OCF_3$, $-CH_3$, $-CH_2CH_3$, $-OCH_3$, $-OCH_2CH_3$, and C_3-C_7 cycloalkyl;

R^3 is C_1-C_4 alkyl optionally substituted with R^{3a};
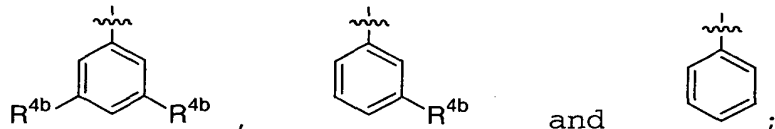
R^{3a} is selected from the group consisting of R^{3b}, C_3-C_6 cycloalkyl optionally substituted with R^{3b}, phenyl

optionally substituted with R^{3b}, and

3,4-methylenedioxyphenyl;

R^{3b} is independently selected at each occurrence from
the group consisting of halogen, $-NO_2$, $-CN$,
$-(C_1-C_4)$alkyl, $-CF_3$, $-OH$, $-OCH_3$, $-OCH_2CH_3$, OCF_3,
$-SCF_3$, $-C(=O)R^6$, $-NR^6C(=O)R^6$, $-NR^6SO_2R^6$, $-NR^6R^6$,
$-OC(=O)NR^6R^6$, $-NR^6C(=O)NR^6R^6$, $-C(=O)NR^6R^6$,
$-C(=O)OR^6$, $-SR^6$, $-S(=O)R^6$, $-S(=O)_2R^6$, and
$-S(=O)_2NR^6R^6$;

R^4 is C_1-C_4 alkyl substituted with R^{4a};

R^{4a} is selected from the group consisting of

R^{4b} is selected from the group consisting of F, Cl, Br,
$-CH_3$, $-CH_2CH_3$, $-CF_3$, $-OCF_3$, $-SCF_3$, $-OH$, $-OCH_3$,
$-SH$, $-SCH_3$, $-CO_2H$, $-CO_2CH_3$, $-NH_2$, $-NH(CH_3)$,
$-N(CH_3)_2$, $-C(=O)NH_2$, $-C(=O)CH_3$, and $-NHC(=O)CH_3$;
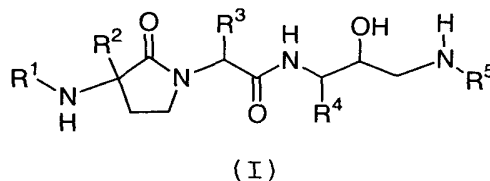
R^5 is C_1-C_{10} alkyl optionally substituted with R^{5a};

R^{5a} is selected from the group consisting of R^{5b},
C_3-C_8 cycloalkyl optionally substituted with R^{5b},
C_2-C_6 alkynyl optionally substituted with R^{5b}, and
phenyl optionally substituted with R^{5b};

R^{5b} is selected from the group consisting of R^6,
halogen, -CN, $-CF_3$, $-NO_2$, $-OCF_3$, $-CO_2H$, -C(=O)H,
$-OR^6$, $-NR^6R^6$, $-OC(=O)NR^6R^6$, $-NR^6C(=O)NR^6R^6$,
$-C(=O)NR^6R^6$, $-C(=O)OR^6$, $-SR^6$, $-S(=O)R^6$, $-S(=O)_2R^6$,
and $-S(=O)_2NR^6R^6$; and

R^6 is independently selected at each occurrence from
the group consisting of hydrogen, C_1-C_6 alkyl and
phenyl.

In yet another preferred embodiment, the present
invention provides compounds of Formula (I)

(I)

or a stereoisomer; or a pharmaceutically acceptable
salt thereof, wherein

R^1 is selected from the group consisting of $-C(=O)R^{1a}$,
$-S(=O)R^{1a}$, $-S(=O)_2R^{1a}$, $-C(=O)OR^{1a}$, and
$-C(=O)NHR^{1a}$;

R^{1a} is C_1-C_6 alkyl optionally substituted with R^{1b};

R^{1b} is independently selected from the group consisting
of halogen, $-CF_3$, $-OCF_3$, $-NR^6R^6$, $-OR^6$,
$-(C_3$-$C_7)$cycloalkyl, -imidazole, thiazole, and
oxazole;

R^2 is selected from the group consisting of C_1-C_4 alkyl optionally substituted with halogen, -CF_3, -OCH_3, -OCH_2CH_3, or C_3-C_7 cycloalkyl;

5   R^3 is C_1-C_4 alkyl optionally substituted with R^{3a};

R^{3a} is selected from the group consisting of phenyl optionally substituted with R^{3b}, and 3,4-methylenedioxyphenyl;

10

R^{3b} is independently selected at each occurrence from the group consisting of F, Cl, R^6, -CF_3, OH, -OCH_3, -OCH_2CH_3, and -NR^6R^6;

15   R^4 is C_1-C_4 alkyl substituted with R^{4a};

R^{4a} is selected from the group consisting of

R^{4b}—⟨benzene⟩—R^{4b} ,   ⟨benzene⟩—R^{4b} and   ⟨benzene⟩ ;
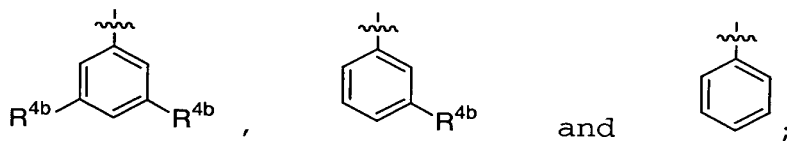
20

R^{4b} is selected from the group consisting of F, Cl, Br, -CH_3, -CF_3, -OH, -OCH_3, -NH_2, -NH(CH_3), and -N(CH_3)$_2$;

25   R^5 is C_1-C_2 alkyl optionally substituted with R^{5a};

R^{5a} is selected from the group consisting of R^{5b},

C$_3$-C$_4$ cycloalkyl optionally substituted with R^{5b}, alkynyl, and phenyl optionally substituted with R^{5b};

5   R^{5b} is selected from the group consisting of R^6, F, Cl, -CN, -OR^6, and -NR^{6}R^6; and

R^6 is independently selected at each occurrence from the group consisting of hydrogen, C$_1$-C$_6$ alkyl and
10   phenyl.

In still yet another preferred embodiment, the present invention provides stereoisomer compounds of Formula (Ia)

15

(Ia)
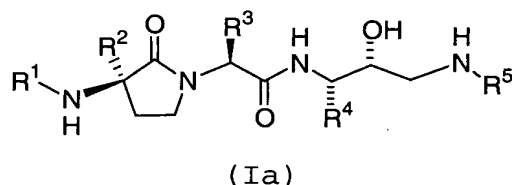
or a pharmaceutically acceptable salt thereof.
20   Preferred compounds for use in the method of the present invention include the compounds of Formula (I) listed below:
(2S)-2-(3(S)-Acetylamino-3-((S)-sec-butyl)-2-oxo-pyrrolidin-1-yl)-N-[(1S, 2R)-1-(3,5-difluoro-benzyl)-
25   2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-4-phenyl-butyramide;
(2S)-2-(3(S)-Acetylamino-3-((S)-*sec*-butyl)-2-oxo-pyrrolidin-1-yl)-*N*-[(1S, 2R)-1-benzyl-2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-4-phenyl-butyramide;

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(2S)-2-(3(S)-Acetylamino-3(-cyclopropylmethyl)-2-oxo-pyrrolidin-1-yl)-N-[(1S, 2R)-1-benzyl-2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-4-phenyl-butyramide;

5

(2S)-2-(3(S)-(2(S)-amino-5-carboxypentanoylamino)-3-((S)-sec-butyl)-2-oxo-pyrrolidin-1-yl)-N-[(1S,2R)-1-benzyl-2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-4-phenyl-butyramide;

(2S)-2-(3(S)-(2-methoxy-acetylamino)-3-((S)-sec-butyl)-2-oxo-pyrrolidin-1-yl)-N-[(1S, 2R)-1-benzyl-2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-4-phenyl-butyramide;

10

(2S)-2-(3(S)-propionylamino-3-((S)-sec-butyl)-2-oxo-pyrrolidin-1-yl)-N-[(1S, 2R)-1-benzyl-2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-4-phenyl-butyramide;

(2S)-2-(3(S)-ethoxycarbonylamino-3-((S)-sec-butyl)-2-oxo-pyrrolidin-1-yl)-N-[(1S, 2R)-1-benzyl-2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-4-phenyl-butyramide;

15

(2S)-2-(3(S)-methoxycarbonylamino-3-((S)-sec-butyl)-2-oxo-pyrrolidin-1-yl)-N-[(1S, 2R)-1-benzyl-2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-4-phenyl-butyramide;

(2S)-2-(3(S)-ethylureido-3-((S)-sec-butyl)-2-oxo-pyrrolidin-1-yl)-N-[(1S, 2R)-1-benzyl-2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-4-phenyl-butyramide;

20

(2S)-2-(3(S)-(3-hydroxypropionylamino)-3-((S)-sec-butyl)-2-oxo-pyrrolidin-1-yl)-N-[(1S, 2R)-1-benzyl-2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-4-phenyl-butyramide;

25

(2S)-2-(3(S)-(4-hydroxybutyrylamino)-3-((S)-sec-butyl)-2-oxo-pyrrolidin-1-yl)-N-[(1S, 2R)-1-benzyl-2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-4-phenyl-butyramide;

30

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(2S)-2-(3(S)-acetylamino-3-(isobutyl)-2-oxo-pyrrolidin-1-yl)-N-[(1S, 2R)-1-benzyl-2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-4-phenyl-butyramide;

(2S)-2-(3(S)-acetylamino-3-((S)-sec-butyl)-2-oxo-

5 pyrrolidin-1-yl)-N-[(1S, 2R)-1-benzyl-2-hydroxy-3-(3-chloro-benzylamino)-propyl]-4-phenyl-butyramide;

(2S)-2-(3(S)-acetylamino-3-((S)-sec-butyl)-2-hydroxy-pyrrolidin-1-yl)-N-[(1S, 2R)-1-benzyl-2-hydroxy-3-(propargylamino)-propyl]-4-phenyl-butyramide;

(2S)-2-(3(S)-acetylamino-3-((S)-sec-butyl)-2-oxo-

10 pyrrolidin-1-yl)-N-[(1S, 2R)-1-benzyl-2-hydroxy-3-(3,5-difluorobenzylamino)-propyl]-4-phenyl-butyramide;

(2S)-2-(3(S)-acetylamino-3-((S)-sec-butyl)-2-oxo-pyrrolidin-1-yl)-N-[(1S, 2R)-1-benzyl-2-hydroxy-3-((3-trifluoromethylbenzyl)amino)-propyl]-4-phenyl-

15 butyramide;

2-(3(S)-Acetylamino-3(S)-isobutyl-2-oxo-pyrrolidin-1-yl)-N-[(1S, 2R)-1-benzyl-2-hydroxy-3-benzylamino-propyl]-4-phenyl-butyramide;

(2S)-2-(3(S)-acetylamino-3-((S)-sec-butyl)-2-oxo-

20 pyrrolidin-1-yl)-N-[(1S, 2R)-1-benzyl-2-hydroxy-3-(3-fluoro,5-(trifluoromethyl)benzylamino)-propyl]-4-phenyl-butyramide;

2-(3(S)-Acetylamino-3(S)-isobutyl-2-oxo-pyrrolidin-1-yl)-N-[(1S, 2R)-1-benzyl-3-(2-cyano-ethylamino)-2-

25 hydroxy-propyl]-4-phenyl-butyramide;

(2S)-2-(3(S)-acetylamino-3-(cyclopropylmethyl)-2-oxo-pyrrolidin-1-yl)-N-[(1S, 2R)-1-(3,5-difluorobenzyl)-2-hydroxy-3-(3-methoxybenzylamino)-propyl]-4-(2-methoxyphenyl)-butyramide;

30 (2S)-2-(3(S)-acetylamino-3-(cyclopropylmethyl)-2-oxo-pyrrolidin-1-yl)-N-[(1S, 2R)-1-(3,5-difluorobenzyl)-2-

hydroxy-3-(3-methoxybenzylamino)-propyl]-4-(3,4-
methylenedioxyphenyl)-butyramide;

(2S)-2-(3(S)-acetylamino-3-(cyclopropylmethyl)-2-oxo-
pyrrolidin-1-yl)-N-[(1S, 2R)-1-(3,5-difluorobenzyl)-2-

5   hydroxy-3-(3-methoxybenzylamino)-propyl]-4-(3-
fluorophenyl)-butyramide;

(2S)-2-(3(S)-acetylamino-3-(cyclopropylmethyl)-2-oxo-
pyrrolidin-1-yl)-N-[(1S, 2R)-1-(3,5-difluorobenzyl)-2-
hydroxy-3-(3-methoxybenzylamino)-propyl]-4-(4-

10   fluorophenyl)-butyramide; and

(2S)-2-(3(S)-acetylamino-3-(cyclopropylmethyl)-2-oxo-
pyrrolidin-1-yl)-N-[(1S, 2R)-1-(3,5-difluorobenzyl)-2-
hydroxy-3-(3-methoxybenzylamino)-propyl]-4-(3-
methoxyphenyl)-butyramide;

15   or a pharmaceutically acceptable salt thereof.

In another aspect, the present invention provides
a pharmaceutical composition for the treatment of
disorders responsive to the inhibition of β-amyloid
peptide production comprising a therapeutically

20   effective amount of Formula (I) in association with a
pharmaceutically acceptable carrier or diluent.

In yet another aspect, the present invention
provides a method for the treatment of a neurological
disorder associated with β-amyloid production by β-

25   secretase comprising administering to a host in need
of such treatment a therapeutically effective amount
of a compound of Formula (I).

In a preferred embodiment the neurological
disorder associated with β-amyloid production by β-

30   secretase is Alzheimer's disease, cerebral amyloid
angiopathy and Down's Syndrome.

Thus, the present invention provides a method for inhibiting β-secretase activity comprising administering to a host in need of such inhibition a therapeutically effective amount of a compound of

5 Formula (I) that inhibits β-secretase activity.

In still another aspect, the present invention provides for the use of a compound of Formula (I) for the manufacture of a medicament for the treatment of Alzheimer's disease.

10 The compounds of the present invention can be prepared in a number of ways well known to one skilled in the art of organic synthesis. The compounds of the present invention can be synthesized using the methods described below, together with synthetic methods known

15 in the art of synthetic organic chemistry, or variations thereon as appreciated by those skilled in the art. Preferred methods include, but are not limited to, those described below. All references cited herein are hereby incorporated in their entirety

20 herein by reference.

The novel compounds of this invention may be prepared using the reactions and techniques described in this section. The reactions are performed in solvents appropriate to the reagents and materials

25 employed and are suitable for the transformations being effected. Also, in the description of the synthetic methods described below, it is to be understood that all proposed reaction conditions, including choice of solvent, reaction atmosphere,

30 reaction temperature, duration of the experiment and workup procedures, are chosen to be the conditions standard for that reaction, which should be readily recognized by one skilled in the art. It is

understood by one skilled in the art of organic
synthesis that the functionality present on various
portions of the molecule must be compatible with the
reagents and reactions proposed. Such restrictions to
5   the substituents which are compatible with the
reaction conditions will be readily apparent to one
skilled in the art and alternate methods must then be
used.

In general, compounds of the invention
10   represented by Formula I (General Reaction Scheme A)
can be prepared by coupling, under standard conditions
known to one skilled in the art, a substituted γ-lactam
2 and a substituted 2-hydroxy-1,3-diaminopropane **3**.
Methods for the synthesis of γ-lactams **2** are known in
15   the art and are disclosed in a number of references
including but not limited to those given below.
Similarly, the synthesis of substituted 2-hydroxy-1,3-
diaminopropanes **3** is known to one skilled in the art
and is disclosed in a number of references including
20   but not limited to those given below.

General Reaction Scheme
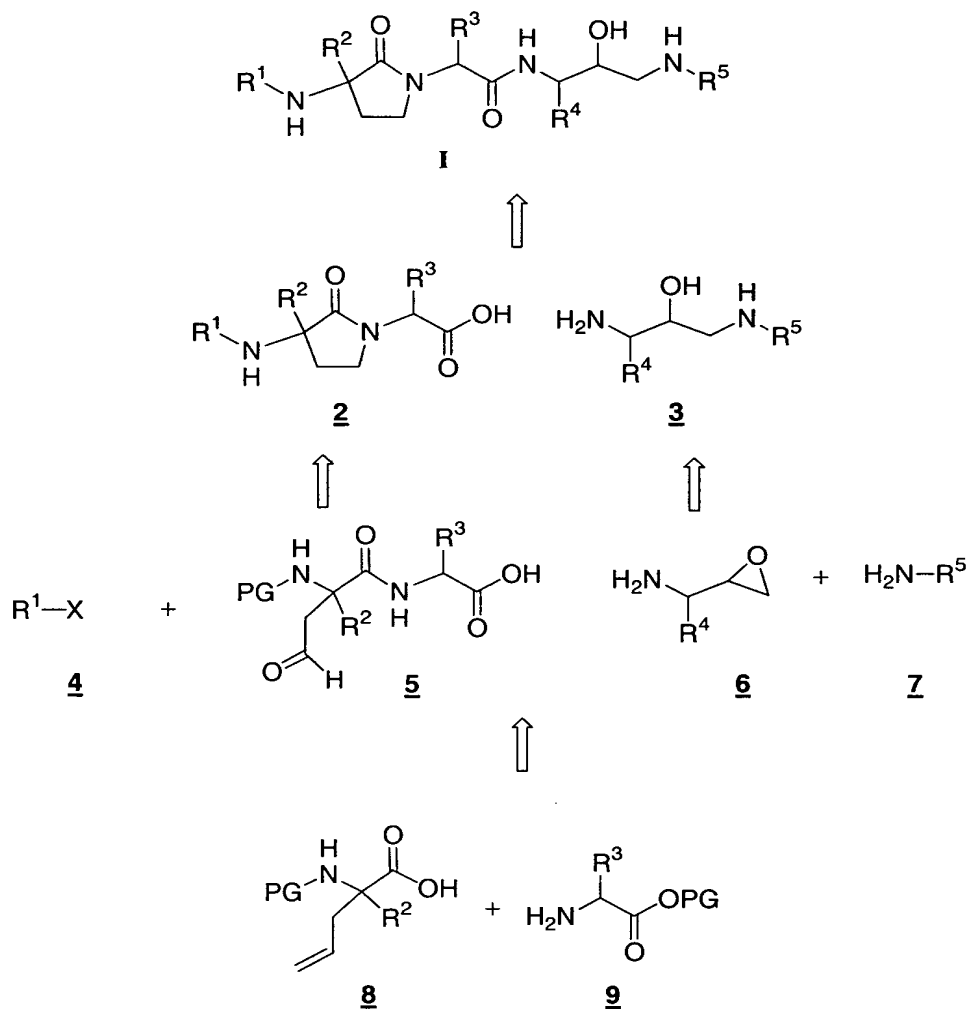
5   Substituted γ-lactams **2** can be prepared by
cyclization of an aldehyde-containing dipeptide
precursor **5** followed by deprotection of the amino
group and functionalization with a suitable reaction
partner **4**, such as a carboxylic acid or an activated
10   derivative thereof, a sulfonyl halide, isocyanate, or
chloroformate. Alternatively, the amino group can be
alkylated under standard conditons known to one
skilled in the art, for example, using an aldehyde and
a reducing agent such as sodium borohydride or

derivatives thereof. The dipeptide precursor **5** is prepared by coupling a natural or unnatural amino acid ester **9** to a quaternary α-allyl amino acid **8**, followed by oxidation of the allyl group to the requisite

5   aldehyde and cyclization. Substituted 2-hydroxy-1,3-diaminopropanes **3** are prepared by reacting an amine with an epoxide **6** which is derived from an amino acid. Further details of the preparation of compounds of the invention are provided below.

10   Synthesis of a substituted quaternary α-allyl amino acid **8** is carried out according to one of several literature methods. Scheme 1 shows the method of Seebach, et. al., (Seebach, D.; Hoffmann, M. *European Journal of Organic Chemistry* **1998**, 1337-1351,

15   Hoffmann, M.; Blank, S.; Seebach, D.; Kusters, E.; Schmid, E. *Chirality* **1998**, *10*, 217-222, Hoffmann, M.; Seebach, D. *Chimia* **1997**, *51*, 90-92, Blank, S.; Seebach, D. *Angew. Chem.* **1993**, *105*, 1780-1781 (See also Angew. Chem., Int. Ed. Engl., 1993, 1732(1712),

20   1765-1786), where (R)- or (S)-tert-butyl 2-tert-butyl-4-methoxy-2,5-dihydro-1,3-imidazole-1-carboxylate (**10**) is alkylated sequentially with allyl iodide and a R^2-group eletrophile (which can be suitably protected by one skilled in the art if necessary) to provide a

25   protected amino acid equivalent with high diastereoselectivity.

Scheme 1
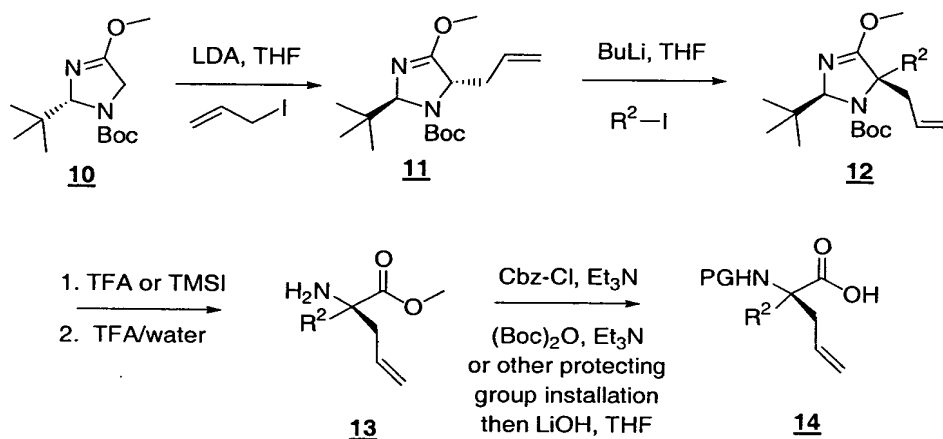
5   The scalemic amino acid is then regenerated by
deprotection of the Boc group and acidic deprotection
of the trimethylacetyl acetal., The resulting amino
acid methyl ester (**13**) can then be protected under
standard conditions with protecting groups well known
10   to those skilled in the art, such as
t-butyloxycarbonyl (Boc) or benzyloxycarbonyl (Cbz),
and saponification provides the free carboxylic acid
14.

      Alternatively, quaternary amino acids can be
15   synthesized from the corresponding amino acid (Scheme
2). Using isoleucine as an example, formation of the
benzylidene imine followed by cyclization with
benzyloxycarbonyl chloride provides a protected amino
acid precursor **17** (Seebach, D.; Fadel, A. *Helv. Chim.*
20   *Acta.* **1985**, *68*, 1243 and Altmann, E.; Nebel, K.;
Mutter, M. *Helv. Chim, Acta* **1991**, *74*, 800; De, B.;
Dellaria, J. F.; Baker, W. R.; Zydowsky, T. M.;
Rosenberg, S. H. *et al.*, EP 365992, **1990**). Alkylation
with allyl bromide or iodide provides the alkylated
25   lactone **18** which can be deprotected under basic

conditions to provide the protected amino acid
derivative **19** which can be directly coupled as is
shown in Scheme 2.

5

Scheme 2
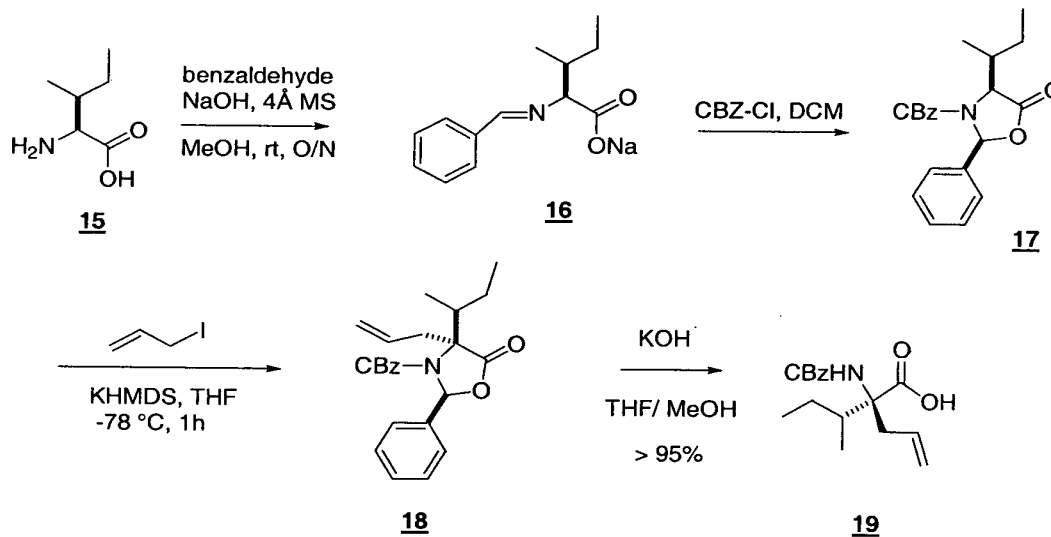
An additional method for the preparation of
10   quaternary amino acids is shown in Scheme 3. Treatment
of an amino acid **20** with allyl bromide in the present
of Cs_2CO_3 provides the amino acid allylic ester **21**.
Ester enolate Caisen rearrangement of **21** results in **22**
(Kazmaier, U. and Maier, S. *Tetrahedron* **1996**, *52*,
15   941).

Scheme 3

20
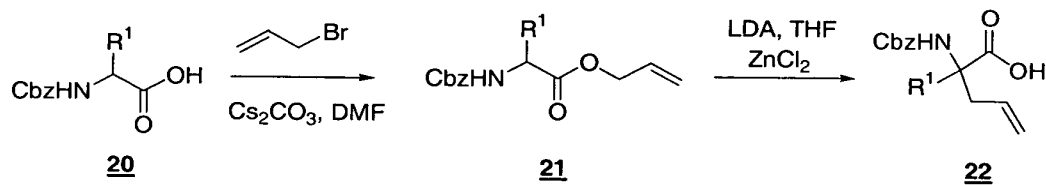
Amino acids used as the starting materials in the chemistry reported herein can be natural or unnatural., Many are available as items of commerce in suitably protected form, or unprotected where

5 protecting groups can be installed under standard conditions to one skilled in the art. Additional methods for the preparation of unnatural amino include the Strecker synthesis or amidomalonate synthesis. In addition, the Myers pseudoephedrine glycinamide

10 alkylation method (Myers, A. G.; Gleason, J. L.; Yoon, T.; Kung, D. W. *J. Am. Chem. Soc.* **1997**, *119*, 656-673), Schollkopf stereoselective alkylation (Schollkoft, U.; Hartwig, W.; Groth, U. *Angew. Chem. Int. Ed. Engl.* **1979**, *18*, 863), and Evans electrophilic azidation

15 (Evans, D. A.; Britton, T. C.; Ellman, J. A.; Dorow, R. L. *J. Am. Chem. Soc.* **1990**, *112*, 4011) may be used to prepare natural or unnatural amino acids in enantionmerically pure form.

The quaternary amino acid **23** may then be coupled

20 under standard conditions to a natural or unnatural amino acid ester using standard coupling reagents like HATU (*O*-(7-azabenzotriazol-1-yl)-1,1,3,3,-tetramethyluronium hexafluorophosphate) or PyBOP (benzotriazole-1-yl-oxy-tris-pyrrolidino-phosphonium

25 hexafluorophosphate) in the presence of a tertiary amine base such as triethylamine, *N,N*-diisopropylethylamine, or *N*-methylmorpholine (Scheme 4). Oxidation of the allyl group using oxonolysis or osmium tetroxide/sodium periodate gives the aldehyde

30 which is cyclized to the γ-lactam **27** using trietthylsilane and trifluoroacetic acid (Holladay, M. W.; Nadzan, A. M. *J. Org. Chem.* **1991**, *56*, 3900-3905;

Duan, J. PCT International Publication WO 0059285,
2000.

Scheme 4
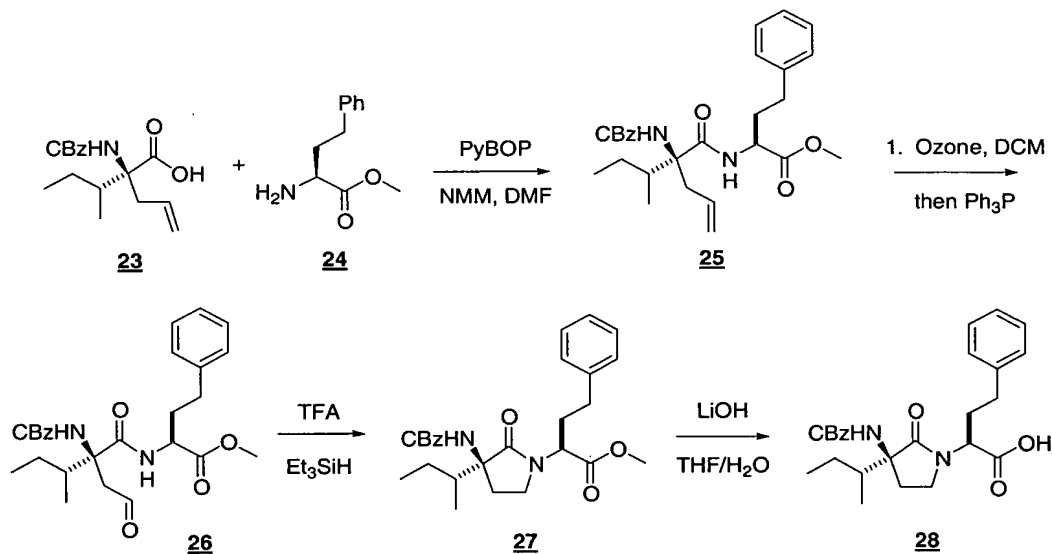
Cleavage of the amino acid ester using
saponification conditions such as lithium or sodium
hydroxide in aqueous solution provides the protected
lactam **28** for coupling to the diaminopropane fragment.

Lactams may also be synthesized in the manner
demonstrated in Scheme 5, where the quaternary amino
acid is directly oxidized to the aldehyde, and a
second amino acid ester is introduced by reductive
alkylation using a reducing agent such as sodium
borohydrode, sodium triacetoxyborohydride, or sodium
cyanoborohydride to produce an amine **32**. The product
can then be cyclized directly to form the desired γ-
lactam (see, for instance, Scheidt, K. A.; Roush, W.
R.; McKerrow, J. H.; Selzer, P. M.; Hansell, E.;
Rosenthal, P. J. *Bioorganic & Medicinal Chemistry*
1998, *6*, 2477-2494.

Scheme 5
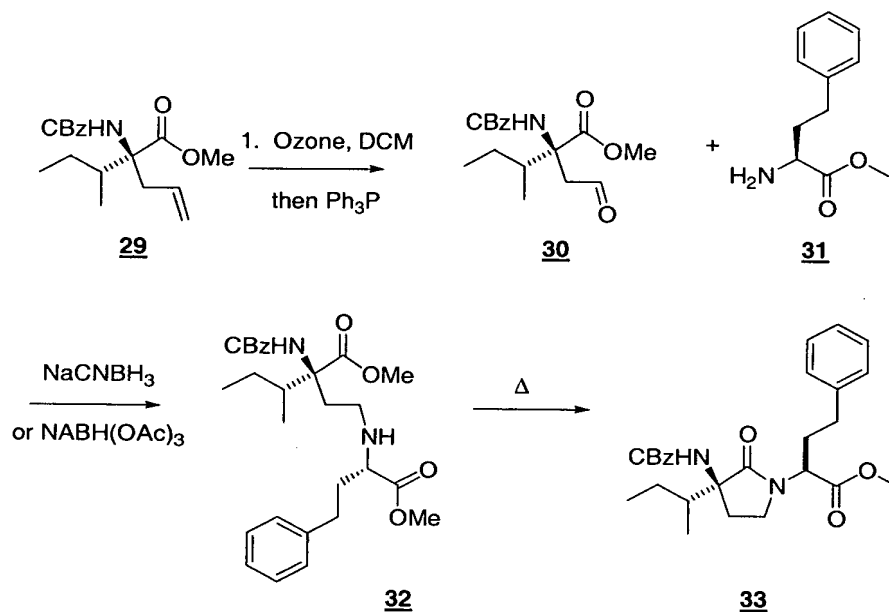
5   The lactam amine protecting group may now be removed by catalytic hydrogenation or other suitable methods (Scheme 6), and the primary amine center may be further functionalized by reacting with agents such as carboxylic acids or their activated variants such

10   as acid chlorides or acid anhydrides to make amides such as **36**. A number of other derivatives **36** can be prepared, including but not limited to the reaction with sulfonic acids or sulfonyl halides to prepare sulfonamides, chloroformates to provide carbamates, or

15   carbamoyl chlorides or isocyanates to provide ureas. Saponification of the methyl ester of these derivatives provides the carboxylic acid **37** ready to couple to the diaminopropane fragment in protected or unprotected form.

20
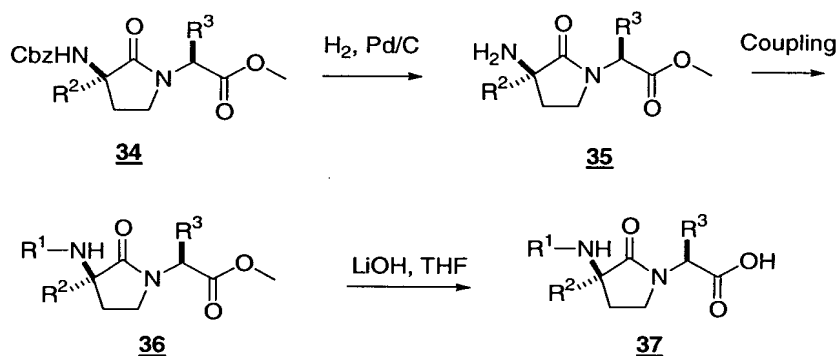
Scheme 6

5   Scheme 7 discloses methods for preparing 2-
hydroxy-1,3-diaminopropanes of type **43** that are used
as a coupling partner to lactam acids **37**. The starting
materials for the process of preparing amino alcohols
43 in accordance with the present invention are
10   activated esters represented by **38** wherein R^4 and R^5
are as defined above and X is Cl or a phenyl ester
substituted in the ortho or para position on the
phenyl ring by hydrogen, halogen or a nitro group. The
compounds represented by formula **38** are commercially
15   available or can be prepared by techniques well known
to those of ordinary skill in the art. The protecting
group on the amino function is preferably Boc or Cbz,
but can also be other art recognized amino function
protecting groups.

20

Scheme 7
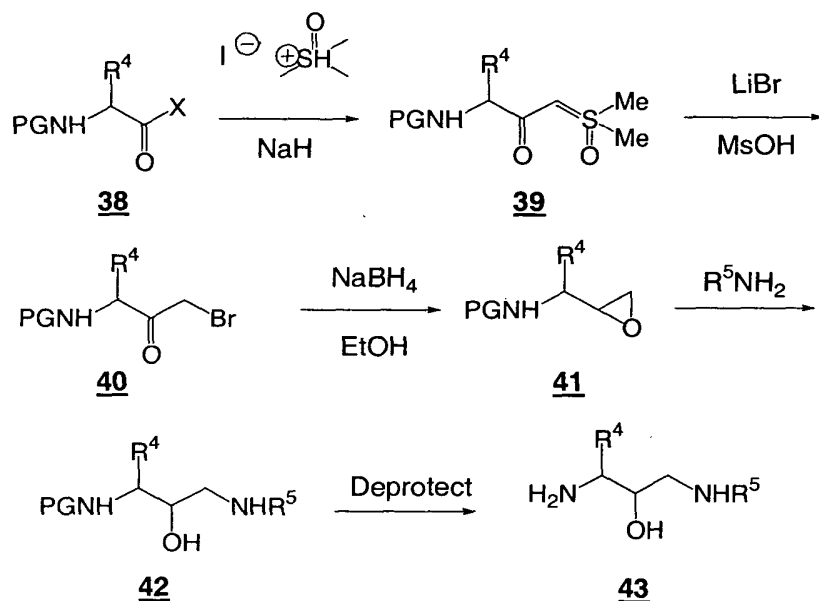
5   In accordance with the process of the present
invention, the starting material represented by
formula **38** above is treated with a sulfur ylide to
produce an intermediate keto ylide compound
represented by **39** [Kronenthal, D. *et al.*, WO 02/14256

10   Al]. The sulfur ylide reagent is conveniently prepared
from a sulfoxonium salt by reaction with a suitable
base in an organic solvent. Suitable sulfoxonium
compounds include trialkyl sulfoxonium halides, such
as trimethylsuloxonium iodide. Preferable bases

15   include, for example, sodium hydride, potassium tert-
butoxide and potassium tert-amylate, with the latter
being particularly preferred. The reaction is carried
out in an organic solvent such as dimethylformamide,
tetrahydrofuran or, preferably, toluene with mild

20   heating, i.e. at a temperature of from about 60 °C to
about 80 °C, preferably about 70 °C. Once the sulfur
ylide reagent is formed, it is reacted with the

starting material **38**, optionally in the presence of a co-solvent. As an example of the use of a mixed solvent reaction medium, the reaction of the trialkylsulfoxonium compound and base is carried out

5   in toluene as described, the resulting solution is cooled to about 0 °C, and then added to a solution of the starting material in tetrahydrofuran to form the keto ylide intermediate compound represented by **39** above.

10   The keto-ylide compound **39** is then converted to the bromoketone **40** by reaction with a source of bromide, preferably a basic source of bromide, most preferably lithium bromide, and an organic acid, for example, methanesulfonic acid. The treatment with the

15   bromide source is carried out in an organic solvent, such as tetrahydrofuran, toluene or, preferably, acetonitrile. The reaction is initiated at low temperature, from about 0 °C to about 5 °C. As the reaction proceeds however, the temperature is raised

20   to about 65 °C.

The bromoketone compound **40** is then converted to the epoxide **41** by reaction with a suitable hydride source such as borohydride or aluminum hydride, most preferably sodium borohydride. The reaction is carried

25   out in a protic solvent such as alcohol or water, most preferably in ethanol. The reaction is initiated at low temperature such as 0 °C to about 5 °C and as the reaction proceeds the temperature is elevated to about 25 °C.

30   The epoxide **41** is then converted to amino alcohol **42** by reaction with an amine as defined above in a suitable polar solvent such as tetrahydrofuran, acetonitrile or alcohol. The reaction can be carried

out with a Lewis acid additive such as lithium-based salts, titanium-based salts or aluminum-based salts. The reaction is carried out at a temperature range of 20-80 °C.

5   The amine protecting group of compound **42** is then removed to give amine **43**. The conditions for protecting group removal are dictated by the nature of the protecting group and are widely known to those skilled in the art. Optionally, the free amine

10   intermediate **42** may be reacted with a suitable, orthogonal protecting group to provide a bis-protected intermediate (Scheme 8). Unmasking of the primary amino group then provides a protected suitable coupling partner **44**. Preferred protecting groups PG_1

15   include Cbz, preferred protecting groups X for NHR^5 include Boc.

Scheme 8

PG_1NH R^4 NHR^5 $\xrightarrow[\text{Deprotect } PG_1]{\text{Protect } NHR^5}$ H_2N R^4 NHR^5
       OH   OH X

20   **42**   **44** X = H, PG
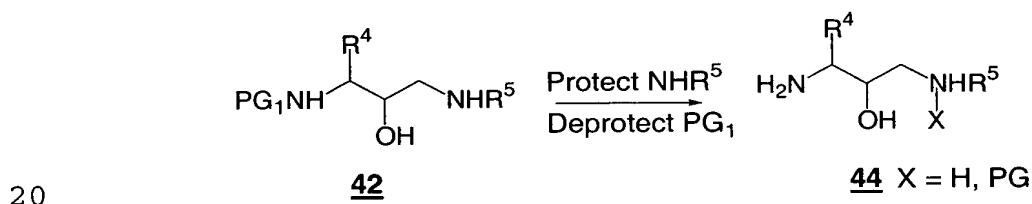
Additional methods for the preparation of 2-hydroxy-1,3-diaminopropanes exist, including those described in Maillaird, M.; Hom, C.; Gailunas, A.;

25   Jagodzinska, B.; Fang, L. Y.; John, V.; Freskos, J. N.; Pulley, S. R.; Beck, J. P.; Tenbrink, R. E. WO 0202512, **2002.** Additionally, a modification of the method of Ellman and coworkers (Kick, E. K.; Ellman, J. A. *J. Med. Chem.* **1995**, *38*, 1427-30.) provides

Scheme 9
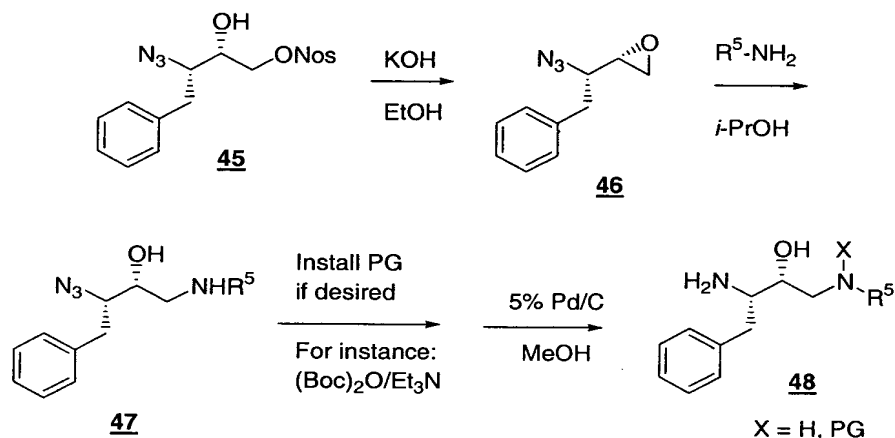
for the preparation of 3-azido-2-hydroxy-1-
aminopropanes (**47**, Scheme 9) which are useful
precursors to 2-hydroxy-1,3-diaminopropanes. These
intermediates may be prepared from the reported
intermediates of type **45**. Treatment of the p-
nitrophenylsulfonate (Nos) intermediate with base
provides the azido epoxide **46**. This versatile
intermediate can be opened with amines to provide the
azido alcohols. Optional protection of the secondary
amine can then be performed, and the azide is then
reduced under mild conditions to provide the primary
amine **48** ready for coupling to lactam acids.

Coupling of a lactam acid **49** with a protected or
unprotected amino alcohol **50** using methods previously
described for making amide bonds, such as HATU and
DIEA in DMF, provides a protected or unprotected
product, which can be deprotected if necessary to
provide the compounds **Ia** of the present invention
(Scheme 10). Preferably, if a protecting group X is
used, it is a Boc group, which is removed by treatment
with trifluoroacetic acid in dichloromethane.

Scheme 10
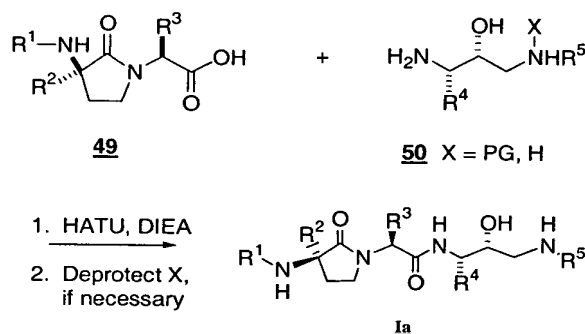
5   Additional examples of intermediate homophenylalanine derivatives related to compound **31** can be prepared using the chemistry shown in scheme 11. Commercial Boc-aspartic acid benzyl ester can be reduced through the intermediate succinimide ester to

10   produce the alcohols **54**. Iodination followed by formation of the alkyl zinc iodide and Negishi-type coupling under palladium catalysis produces substituted, protected homophenylalanines **56** which can be deprotected in the standard manner using

15   trifluoroacetic acid or HCl to produce indermediates **57**, useful in the formation of substituted lactams of type **33**.

Scheme 11
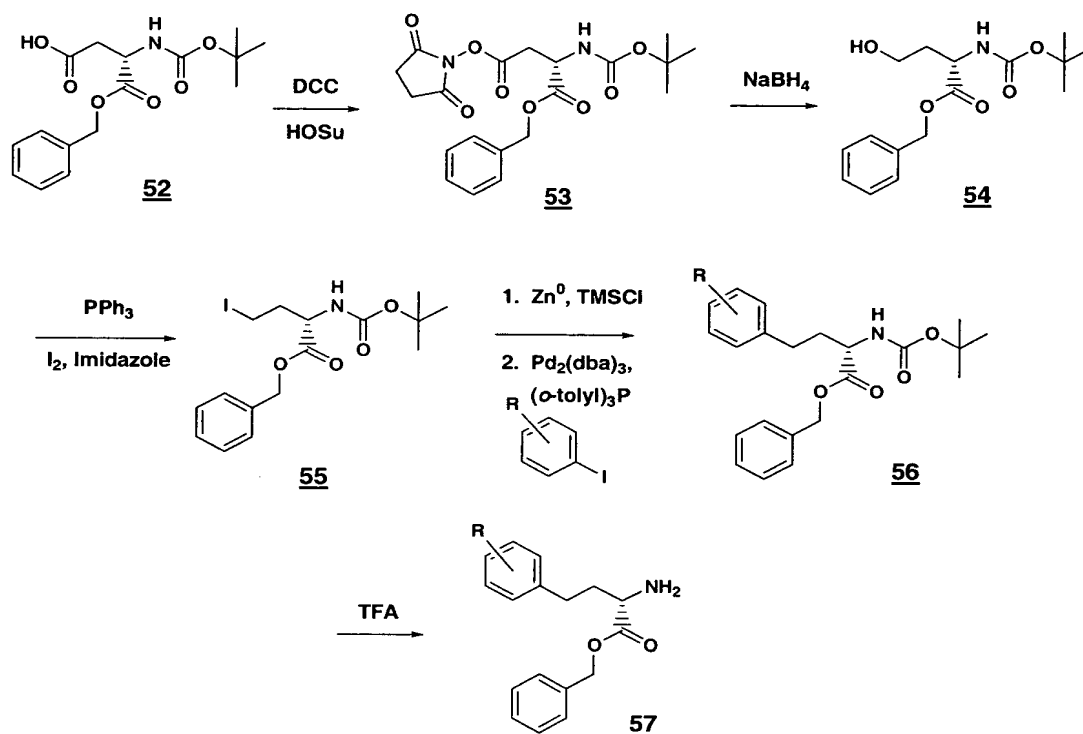
5

DESCRIPTION OF THE SPECIFIC EMBODIMENTS

The compounds of this invention and their preparation can be understood further by the following

10   working examples. These examples are meant to be illustrative of the present invention, and are not to be taken as limiting thereof.

Chemical abbreviations used in the specification and Examples are defined as follows:

15   "Ac" for acetate,

"APCI" for atmospheric pressure chemical ionization,

"Boc" or "BOC" for t-butyloxycarbonyl,

"BOP" for benzotriazol-1-yloxytris-(dimethylamino)-
       phosphonium hexafluorophosphate,

20   "Cbz" for benzyloxycarbonyl,

"CDI" for 1,1'-carbonyldiimidazole,

"CD₃OD" for deuteromethanol,

"CDCl₃" for deuterochloroform,

"DCC" for 1,3-dicyclohexylcarbodiimide,

5 "DCM" for dichloromethane

"DEAD" for diethyl azodicarboxylate,

"DIEA" for *N,N*-diisopropylethylamine,

"DIPEA" for *N,N*-diisopropylethylamine,

"DMF" for N,N-dimethylformamide,

10 "DMAP" for 4-dimethylaminopyridine,

"DMPU" for 1,3-dimethyl-3,4,5,6-tetrahydro-2(1H)-
    pyrimidone,

"DMSO" for dimethylsulfoxide,

"EDC" or "EDCI" for 1-(3-dimethylaminopropyl)-3-

15   ethylcarbodiimide hydrochloride,

"Et" for ethyl,

"EtOAC" for ethyl acetate,

"HOAc" for acetic acid,

"HOBt" for 1-hydroxybenzotriazole hydrate,

20 "HATU" for O-(7-azabenzotriazol-1-yl)-N, N, N', N'-
    tetramethyluronium hexafluorophosphate,

"HMPA" for hexamethylphosphoramide,

"LDA" for lithium diisopropylamide,

"LiHMDS" for lithium bis(trimethylsilyl)amide,

25 "NaHMDS" for sodium bis(trimethylsilyl)amide,

"NMM" for 4-methylmorpholine,

"PyBOP" for benzotriazole-1-yl-oxy-tris-pyrrolidino-
    phosphonium hexafluorophosphate,

"TMSCH₂N₂" for (trimethylsilyl)diazomethane,

30 "TMSN₃" for Azidotrimethylsilane,

"TBTU" for O-(1H-benzotriazol-1-yl)-N,N,N',N'-
    tetramethyluronium tetrafluoroborate,

"TEA" for triethylamine,

"TFA" for trifluoroacetic acid, and

"THF" for tetrahydrofuran.

Abbreviations used in the Examples are defined as
follows: "°C" for degrees Celsius, "MS" for mass
5   spectrometry, "ESI" for electrospray ionization mass
spectroscopy, "HR" for high resolution, "LC-MS" for
liquid chromatography mass spectrometry, "eq" for
equivalent or equivalents, "g" for gram or grams, "h"
for hour or hours, "mg" for milligram or milligrams,
10   "mL" for milliliter or milliliters, "mmol" for
millimolar, "M" for molar, "min" for minute or
minutes, "rt" for room temperature, "NMR" for nuclear
magnetic resonance spectroscopy, "tlc" for thin layer
chromatography, "atm" for atmosphere, and "α", "β",
15   "R", "S", "E", and "Z" are stereochemical designations
familiar to one skilled in the art.

"HPLC" is an abbreviation used herein for high
pressure liquid chromatography. Reverse-phase HPLC can
be carried out using a Vydac C-18 column with gradient
20   elution from 10% to 100 % buffer B in buffer A (buffer
A: water containing 0.1% trifluoroacetic acid, buffer
B: 10% water, 90% acetonitrile containing 0.1%
trifluoroacetic acid). If necessary, organic layers
can be dried over sodium sulfate unless otherwise
25   indicated. However, unless otherwise indicated, the
following conditions are generally applicable. "LC-
MS" refers to high pressure liquid chromatography
carried out according to the definition for HPLC with
a mass spectrometry detector.

30   Melting points were determined on a Mel-Temp II
apparatus and are uncorrected. IR spectra were
obtained on a single-beam Nicolet Nexus FT-IR
spectrometer using 16 accumulations at a resolution of

4.00 cm-1 on samples prepared in a pressed disc of KBr
or as a film on KBr plates. Proton NMR spectra (300
MHz, referenced to tetramethylsilane) were obtained on
a Varian INOUA 300, Bruker Avance 300, Avance 400, or
5   Avance 500 spectrometer. Data were referred to the
lock solvent. Electrospray Ionization (ESI)
experiments were performed on a Micromass II Platform
single-quadrupole mass spectrometer, or on a Finnigan
SSQ7000 mass spectrometer. HPLC analyses were obtained
10   using a Rainin Dynamax C18 column with UV detection at
223 nm using a standard solvent gradient program as
follows:

HPLC solvent conditions: When described as
performed under "standard conditions", Samples were
15   dissolved in methanol (1 mg/mL) and run using the
following gradient program with a solvent flow rate of
1.0 mL/min.

Time (min)	Acetonitrile (0.05% TFA)	H_2O (0.05% TFA)
Initial	10	90
20.0	90	10
20-30	90	10

Preparatory HPLC: : When described as performed
20   under "standard conditions", Samples (approx. 20 mg)
were dissolved in methanol (10 mg/mL) and purified on
a 25mm X 50 mm Vydac C18 colum with a 5 minute
gradient elution from 10% to 100 % buffer B in buffer
A (buffer A: water containing 0.1% trifluoroacetic
25   acid, buffer B: 10% water, 90% acetonitrile
containing 0.1% trifluoroacetic acid) at 10 mL/minute.

Analytical HPLC: When described as "Method A" , a
sample dissolved in a suitable carrier solvent

(methanol, acetonitrile, or the like) was analyzed on an Xterra 3.0 X 50 mm s7 column with a run time of 3 min and a gradient of 0-100% B over 2 min at a flowrate of 5 mL/min. Absorbance was monitored at 220

5   μM. Solvent A = 0% MeOH/90% water/0.1% TFA and Solvent B = 10% water /90% MeOH /0.1% TFA.

Analytical HPLC: When described as "Method B" , a sample dissolved in a suitable carrier solvent (methanol, acetonitrile, or the like) was analyzed on

10   an Xterra 3.0 X 50 mm s7 column with a run time of 4 min and a gradient of 0-100% B over 3 min at a flowrate of 5 mL/min. Absorbance was monitored at 220 μM. Solvent A = 0% MeOH/90% water/0.1% TFA and Solvent B = 10% water /90% MeOH /0.1% TFA.

15   The examples provided are intended to assist in a further understanding of the invention. Particular materials employed, species and conditions are intended to be further illustrate of the invention and not limit the reasonable scope thereof.

20
Synthesis of Intermediates

Preparation A

(3S,2R) 3-Amino-4-(3,4-difluoro-phenyl)-1-(3-methoxy-

25   benzylamino)-butan-2-ol.
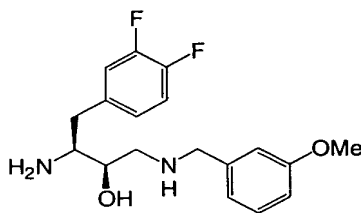
Step A(1).   3-(S)-2-oxo-3-(tertbutyloxycarbonylamino)-
4-(3,4-difluororphenyl)butylide dimethylsulfoxonium.

Trimethylsulfoxonium iodide (1.1 g, 4.7 mmol)
5   suspended in THF (3.5 mL) was treated with potassium
tert-butoxide (3.5 mL, 3.5 mmol, 1 M in THF) via
syringe over one minute at RT. The reaction was
stirred at 70 °C for two hours to afford the
corresponding ylide that was reacted in solution
10   without isolation. The reaction mixture was cooled to
0 °C and a solution of N-(2-t-butoxycarbonyl)-L-3,4-
difluorophenylalanine-4-nitrophenyl ester (0.5 g, 1.2
mmol) in THF (3.0 mL) was added via cannula over 1
minute. The reaction was stirred at this temperature
15   for five minutes and then was allowed to warm to
ambient temperature over 30 minutes. The reaction
mixture was stirred at ambient temperature for a
further 30 minutes. The reaction was quenched with
saturated sodium bicarbonate solution, diluted with
20   ethyl acetate (25 mL), washed with saturated sodium
bicarbonate solution (2 X 25 mL), the organic layer
separated, dried (MgSO$_4$) and the solvent removed at
reduced pressure. The yellowish residue was
recrytallized from hexane to give the desired product
25   as a yellow solid (0.41 g, 93%). MS: 376 (M + H,
100%).

Step A(2).   (1S)-[3-Bromo-1-(3,4-difluoro-benzyl)-2-
oxo-propyl]-carbamic acid tert-butyl ester

30
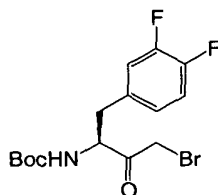
A solution of 3-(S)-2-oxo-3-
(tertbutyloxycarbonyl-amino)-4-(3,4-
difluororphenyl)butylide dimethylsulfoxonium (0.34 g,
0.91 mmol) in THF (3 mL) was cooled to 0 °C and
treated with lithium bromide (0.087 g, 0.91 mmol) in
one portion. After all the lithium bromide dissolved,
methanesulfonic acid (0.056 mL, 0.91 mmol) was added
dropwise over 30 seconds. A slurry begins to form
after 5 minutes and the cooling bath is replaced with
an oil bath. The reaction was heated to 65 °C for two
hours. The reaction was allowed to cool to ambient
temperature, quenched with saturated sodium
bicarbonate solution, diluted with ethyl acetate (25
mL), washed with saturated sodium bicarbonate solution
(2 X 25 mL), the organic layer separated, dried
(MgSO$_4$) and the solvent removed at reduced pressure.
The desired product was obtained as an orange oil and
used without further purification (0.3 g, 90%). MS:
380 (M + H, 100%).

Step A(3). (1S,2S) [2-(3,4-Difluoro-phenyl)-1-
oxiranyl-ethyl]-carbamic acid tert-butyl ester
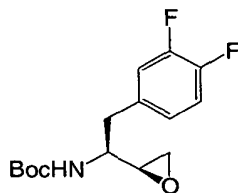
A solution of (1S)-[3-Bromo-1-(3,4-difluoro-benzyl)-2-oxo-propyl]-carbamic acid tert-butyl ester (0.14 g, 0.36 mmol) in ethanol:THF (2:1) (3 mL) was cooled to 0 °C and treated with sodium borohydride

5   (0.014 g, 0.36 mmol) in one portion. After 30 minutes the cooling bath was removed and the reaction mixture was stirred at ambient temperature for 12 hours. The solvent was removed and the residue partitioned between saturated sodium bicarbonate solution (20 mL)

10   and ethyl acetate (20 mL). The organic phase was washed with saturated sodium bicarbonate solution (25 mL), separated, dried (MgSO₄) and the solvent removed at reduced pressure. The product was recrystallized from hexane to give the desired product as the major

15   isomer (>9:1) by NMR (0.095g, 90%). MS: 300 (M + H, 100%).

Step A(4).   (1S,2R)- [1-(3,4-Difluoro-benzyl)-2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-carbamic

20   acid tert-butyl ester.
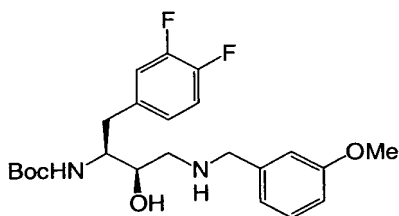
A solution of (1R,2S)-[2-(3,4-difluoro-phenyl)-1-

25   oxiranyl-ethyl]-carbamic acid tert-butyl ester (0.03 g, 0.1 mmol) in acetonitrile (1 mL) was treated with lithium triflate (0.032 g, 0.2 mmol) and stirred at ambient temperature for 20 minutes. Then 3-methoxybenzylamine (0.016 mL, 0.12 mmol) was added to

the reaction neat in one portion. The reaction was
stirred at ambient temperature for 14 hours. The
reaction was poured into saturated ammonium chloride
solution (5 mL), extracted with ethyl acetate, the
organic phase separated, dried (MgSO$_4$) and the solvent
removed at reduced pressure. The product was obtained
as an oil and used without further purification (0.041
g, 94%). MS: 437 (M + H, 100%).

Step A(5): Preparation A. (3S,2R) 3-Amino-4-(3,4-
difluoro-phenyl)-1-(3-methoxy-benzylamino)-butan-2-ol.
        A solution of (1R,2S)-[1-(3,4-Difluorobenzyl)-2-
hydroxy-3-(3-methoxy-benzylamino)-propyl]-carbamic
acid tert-butyl ester (0.04g, 0.09 mmol) in methylene
chloride (1 mL) was treated with 4 N HCl in dioxane (1
mL) at ambient temperature in one portion. The
reaction was stirred for 12 hours and the solvent
removed at reduced pressure to give the product as a
solid (0.033 g, 98%). MS: 337 (M+ H, 100%).

Preparation B

(3S,2R)-3-Amino-4-(3,5-difluoro-phenyl)-1-(3-methoxy-
benzylamino)-butan-2-ol.
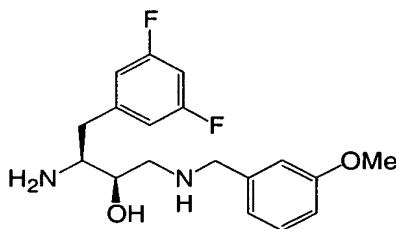
        Following the general procedure for Preparation
(A) using N-(2-t-butoxycarbonyl)-L-3,5-
difluorophenylalanine-4-nitrophenyl ester as the

starting material the title compound was obtained. MS: 337 (M+ H, 100%).

Preparation C

5   (3S,2R) 3-Amino-4-(4-chloro-phenyl)-1-(3-methoxy-benzylamino)-butan-2-ol.
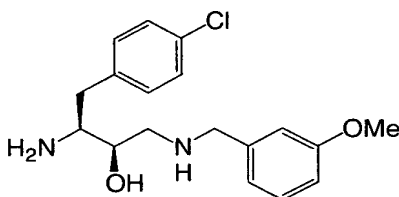
10   Following the general procedure for Preparation (A) using N- (2-t-butoxycarbonyl)-L-4-chlorophenylalanine-4-nitrophenyl ester as the starting material the title compound was obtained. MS: 335 (M+ H, 100%).

15

Preparation D

(3S,2R) (3-Amino-2-hydroxy-4-phenyl-butyl)-(3-methoxy-benzyl)-carbamic acid tert-butyl ester.
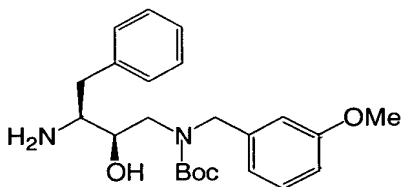
20

Step D(1). (1S,2S) 2-(1-Azido-2-phenyl-ethyl)-oxirane
     A solution of 1.0 g (2.5 mmol)of 4-Nitro-benzenesulfonic acid 3-azido-2-hydroxy-4-phenyl-butyl

25   ester (Kick, E. K.; Ellman, J. A. *J. Med. Chem.* **1995**, *38*, 1427-30) in 12 mL of Ethanol and 8 mL of ethyl

acetate is treated with KOH (157 mg, 2.8 mmol). After 2 h at rt 10 mL of water is added and 20 mL of CH_2Cl_2 is added. The organic layer is separated, washed with brine and Na_2SO_4, and concentrated. Chromatography

5 eluting with hexanes/ethyl acetate 5:1 provides 450 mg (95%) of the desired product. ^1H NMR (δ) 7.21-7.36 (m, 5H), 3.58 (dt, 2H, J = 9, 5 Hz), 3.05 (m, 1H) 2.97 (dd, 1H, J = 13.9, 4.8), 2.77-2.83 (m, 3H).

10 Step D(2). (3S,2R) 3-Azido-1-(3-methoxy-benzylamino)-
4-phenyl-butan-2-ol
A solution of the compound of intermediate D(1) (225 mg, 1.18 mmol) in 9 mL of isopropyl alcohol is treated with 3-methoxybenzylamine (1.74 mmol, 230 µL).

15 The solution was heated to 85 °C and stirred for 3 h. The reaction mixture was then directly concentrated and the product obtained by chromatography eluting with 2% methanol in CH_2Cl_2 to obtain 200 mg (52%) of the desired product as a clear oil. MS $(M+H)^+ = 327.3$

20

Step D(3). (3S,2R) (3-Azido-2-hydroxy-4-phenyl-butyl)-
(3-methoxy-benzyl)-carbamic acid tert-butyl ester
A solution of the compound of intermediate D(2) (100 mg, 0.308 mmol) in 2 mL of CH_2Cl_2 is treated with

25 di(tert-butyl)dicarbonate (0.33 mmol, 73 mg) and triethylamine (0.46 mmol, 63 µL). After stirring at rt for 2 h water was added (5 mL) and the organic layer was separated. The aqueous layer was extracted with 2 additional 10 mL portions of ethyl acetate and

30 the combined organic layer was dried over Na_2SO_4 and concentrated. Chromatography eluting with a gradient of hexanes/ethyl acetate 4:1 to 1:1 provided 125 mg

(95%) of the desired product. MS $(M+H-Boc)^+= 327.3$,

NMR (δ) 1.47 (s, 9H, Boc group)

Step D(4): Preparation D. (3S,2R) (3-Amino-2-hydroxy-
4-phenyl-butyl)-(3-methoxy-benzyl)-carbamic acid tert-
butyl ester.

   A solution of the compound of intermediate D(3)
(125 mg, 0.29 mmol) was dissolved in 3 mL of methanol
and 25 mg of 5% palladium on carbon was added. The
suspension was placed under 50 psi of hydrogen on a
parr apparatus and shaken overnight. The catalyst was
removed by filtration and the product was obtained by
concentration with no purification necessary (100 mg,
86%). MS $(M+H)^+=401.3$

Preparation E

(3S,2R) (3-Amino-2-hydroxy-5-methyl-hexyl)-(3-methoxy-
benzyl)-carbamic acid tert-butyl ester
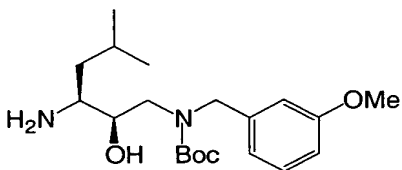
Step E(1) {1(S)-[1(R)-Hydroxy-2-(3-methoxy-
benzylamino)-ethyl]-3-methyl-butyl}-carbamic acid
benzyl ester

   Following the general procedure for intermediate
A(4) using N-(benzyloxycarbonyl)-L-leucine-4-
nitrophenyl ester as the starting material the title
compound is obtained. MS $(M+H)^+=401.4$.

Step E(2). (3(S)-Benzyloxycarbonylamino-2(R)-hydroxy-5-methyl-hexyl)-(3-methoxy-benzyl)-carbamic acid tert-butyl ester

5   Following the general procedure for the synthesis of intermediate D(4) 1.0 g of the compound of Step E(1) is converted to 800 mg (80%) of the title compound. MS (M+H)$^+$=501.4, (M+Na)$^=$ = 523.4.

Step E(3): Preparation E. (3(S)-Amino-2(R)-hydroxy-5-methyl-hexyl)-(3-methoxy-benzyl)-carbamic acid tert-butyl ester

       In a Parr flask 60 mg of 5% palladium on carbon was suspended in 5 ml of methanol. The compound of intermediate E(2) (300 mg, 0.6 mmol) was added and the resulting slurry was placed under 40 psi of hydrogen for 16 h in a Parr apparatus. The catalyst was then removed by filtration and the title compound of Preparation (E) (250 mg, 85%) was isolated by concentrating the resulting sloution. MS (M+H)$^+$=367.4

Preparation F

(3S,2R) (3-Amino-2-hydroxy-4-phenyl-butyl)-(3-methoxy-benzyl)-carbamic acid benzyl ester.
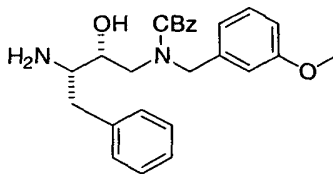
Step F(1). (1S, 2R) [1-Benzyl-2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-carbamic acid tert-butyl ester.

Following the general procedure for the preparation of Preparation (A), step A(4), but using N-Boc-phenylalanine as the starting material the compound of step F(1) was obtained. MS ESI (M+H)$^{+}$ =
5   401.3.

Step F(2). (1S, 2R) {1-Benzyl-3-[benzyloxycarbonyl-(3-methoxy-benzyl)-amino]-2-hydroxy-propyl}-carbamic acid tert-butyl ester.

10   The compound from step F(1) (1.5 g, 3.7 mmol) was dissolved in 30 mL of CH_2Cl_2 and benzyl chloroformate (0.6 mL, 3.9 mmol) and triethylamine (1 mL) were added. After stirring at rt for 2 h, the reaction solution was diluted with water and extracted with 2

15   50 mL portions of CH_2Cl_2. The combined organic layers were dried and concentrated to provide a crude product which was purified by chromatography eluting with 25-50% ethyl acetate in hexanes to provide 1.7 g (86%) of the desired product.

20

Step F(3). Preparation F. (3S, 2R) (3-Amino-2-hydroxy-4-phenyl-butyl)-(3-methoxy-benzyl)-carbamic acid benzyl ester.

      The compound from step F(2) (29 mg, 0.054 mmol)
25   was dissolved in 3 mL of 4.0 M HCl in dioxane. After 1 h at rt, the solvent was removed by evaporation to provide the amine HCl salt which was used without further purification. ESI MS (M+H)$^{=}$ = 435.3.

30   **Preparation G**

(3S,2R)-3-Amino-4-(2-fluoro-phenyl)-1-(3-methoxy-benzylamino)-butan-2-ol.
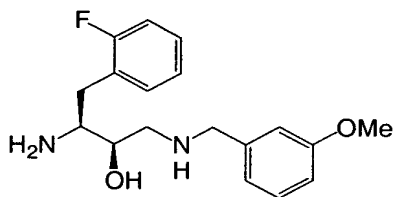
Following the general procedure for Preparation (A) using N-(2-t-butoxycarbonyl)-L-2-fluorophenylalanine-4-nitrophenyl ester as the starting material the title compound was obtained. MS: 319.3 (M+ H, 100%).

Preparation H

(3S,2R)-3-Amino-4-(2-fluoro-phenyl)-1-(3-chloro-benzylamino)-butan-2-ol.
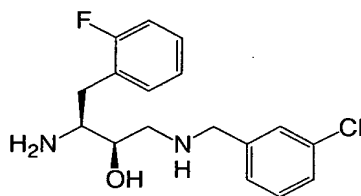
Step H(1): Following the general procedure for the synthesis of the compound of Preparation A(3) but using N-(2-t-butoxycarbonyl)-L-2-fluorophenylalanine-4-nitrophenyl ester as the starting material the epoxide was obtained. APCI MS: (M+ H)$^+$ = 282.

Step H(2): Following the general procedure for the synthesis of the compound of Preparation A(4) but using the epoxide from step H(1) and 3-chlorobenzylamine the amine of step H(2) was prepared. APCI MS: (M+ H)$^+$ = 423.

Step H(3): Following the general procedure for the synthesis of the compound of Preparation A(5) but using the amine from step H(2) the title compound of Preparation (H) was prepared. APCI MS: (M+ H)$^+$ = 323.

Preparation I

(3S)-(3-Amino-2-hydroxy-4-phenyl-butyl)-(3-methoxy-benzyl)-carbamic acid tert-butyl ester.

Step I(1): Following the general procedure for the synthesis of the compound of Preparation (A) but using N-(2-t-butoxycarbonyl)-L-phenylalanine-4-nitrophenyl ester as the starting material and omitting the crstallization step in the preparation of the intermediate I(3), the compound of Preparation (I) was prepared as an approximately 1:2 mixture of the erythro:threo diastereomers at C2. ESI MS (M+H)$^+$ = 401.43.

Preparation J

(3S, 2R)-3-Amino-1-(3-chloro-benzylamino)-4-phenyl-butan-2-ol
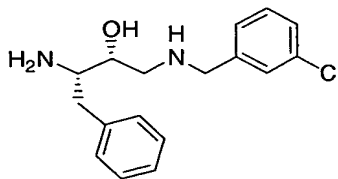
Step J(1): Following the general procedure for
the synthesis of the compound of Preparation A(3) but
using N-(2-t-butoxycarbonyl)-L-phenylalanine-4-
nitrophenyl ester as the starting material the epoxide
5   was obtained. APCI MS: (M+ H)$^+$ = 264.

Step J(2): Following the general procedure for
the synthesis of the compound of Preparation A(4) but
using the epoxide from step J(1) and 3-
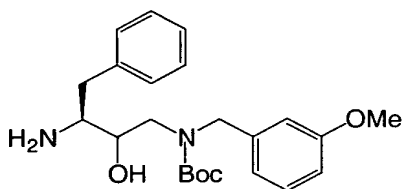10   chlorobenzylamine the amine of step J(2) was prepared.
APCI MS: (M+ H)$^+$ = 405.2.

Step J(3): Following the general procedure for
the synthesis of the compound of Preparation A(5) but
15   using the amine from step J(2) the title compound of
Preparation (H) was prepared as the solid HCl salt.
APCI MS: (M+ H)$^+$ = 305.2.

Preparation K

20   (3S, 2R)-3-Amino-4-phenyl-1-prop-2-ynylamino-butan-2-
ol
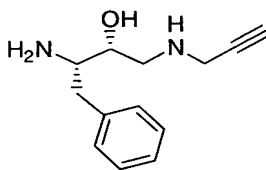
25   Step K(1): Following the general procedure for
the synthesis of the compound of Preparation A(4) but
using the epoxide from step J(1) and propargylamine
the amine of step K(1) was prepared. APCI MS: (M+ H)$^+$ =
319.

30

Step K(2): Following the general procedure for the synthesis of the compound of Preparation A(5) but using the amine from step K(1) the title compound of Preparation (K) was prepared as the solid HCl salt.

5   APCI MS: (M+ H)$^+$ = 219.

Preparation L

(3S, 2R) (3-Amino-2-hydroxy-4-phenyl-butyl)-cyclopropylmethyl-carbamic acid tert-butyl ester.
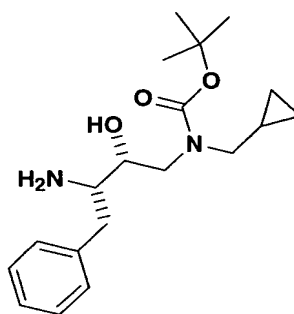
10

Following the general procedure for Preparation (A) (steps A4-A5) using (1S, 2S) (1-Oxiranyl-2-phenyl-

15   ethyl)-carbamic acid benzyl ester as the starting material, the title compound was obtained as a colorless oil (136 mg) in a 44 % yield. LC-MS (column = XTERRA C18 S7, 3 x 50 mm, start %B = 0, final %B = 100, gradient time = 2 min, flow rate = 5 ml/min) m/e

20   335.28 (M + H)$^+$, t_R 1.30 min.

Preparation M

(3S, 2R)-(3-Amino-2-hydroxy-4-phenyl-butyl)-(3,5-difluoro-benzyl)-carbamic acid tert-butyl ester

25

Following the general procedure for Preparation (A) (steps A4-A5) using (1S, 2S) (1-Oxiranyl-2-phenyl-ethyl)-carbamic acid benzyl ester as the starting material the title compound was obtained. LC-MS (column = XTERRA C18 S7, 3 x 50 mm, start %B = 0, final %B = 100, gradient time = 2 min, flow rate = 5 ml/min) m/e 407.27 (M + H)$^+$, t_R 1.44 min.

Preparation N

(3S, 2R)-(3-Amino-2-hydroxy-4-phenyl-butyl)-(3-trifluoromethyl-benzyl)-carbamic acid tert-butyl ester

Following the general procedure for Preparation (A) (steps A4-A5) using (1S, 2S) (1-Oxiranyl-2-phenyl-ethyl)-carbamic acid benzyl ester as the starting material the title compound was obtained as an amber oil (728 mg) in a quantitative yield. LC-MS (column = XTERRA C18 S7, 3 x 50 mm, start %B = 0,

final %B = 100, gradient time = 2 min, flow rate = 5 ml/min) m/e 439.24 (M + H)$^+$, t_R 1.60 min.

Preparation O

(3S, 2R)-(3-Amino-2-hydroxy-4-phenyl-butyl)-benzyl-carbamic acid tert-butyl ester

Following the general procedure for Preparation (A) (steps A4-A5) using (1S, 2S) (1-Oxiranyl-2-phenyl-ethyl)-carbamic acid benzyl ester as the starting material the title compound was obtained as an colorless oil (190 mg) in a 52 % yield. LC-MS (column = XTERRA C18 S7, 3 x 50 mm, start %B = 0, final %B = 100, gradient time = 2 min, flow rate = 5 ml/min) m/e 371.30 (M + H)$^+$, t_R 1.47 min.

Preparation P

(3S, 2R)- (3-Amino-2-hydroxy-4-phenyl-butyl)-(3-fluoro-5-trifluoromethyl-benzyl)-carbamic acid tert-butyl ester
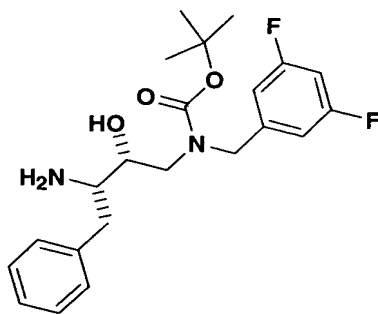
Following the general procedure for Preparation
(A) (steps A4-A5) using (1S, 2S) (1-Oxiranyl-2-
phenyl-ethyl)-carbamic acid benzyl ester as the
starting material the title compound was obtained as
an amber oil (580 mg) in a 64 % yield. LC-MS (column =
XTERRA C18 S7, 3 x 50 mm, start %B = 0, final %B =
100, gradient time = 2 min, flow rate = 5 ml/min) m/e
457.23 (M + H)$^+$, t_R 1.65 min.

Preparation Q

(3S, 2R)-(3-Amino-2-hydroxy-4-phenyl-butyl)-(2-cyano-
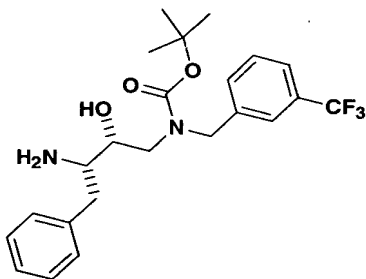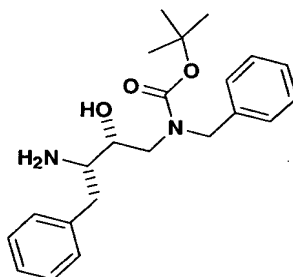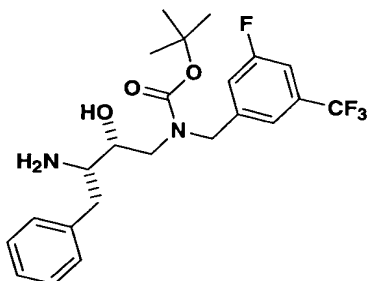ethyl)-carbamic acid tert-butyl ester
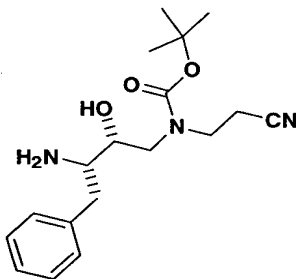
Following the general procedure for Preparation
(A) (steps A4-A5) using (1S, 2S) (1-Oxiranyl-2-
phenyl-ethyl)-carbamic acid benzyl ester as the
starting material the title compound was obtained as
an amber oil (250 mg) in a 32 % yield. LC-MS (column =
XTERRA C18 S7, 3 x 50 mm, start %B = 0, final %B =

100, gradient time = 2 min, flow rate = 5 ml/min) m/e
234.33 $(M + H)^+$, t_R 1.08 min.

<div align="center">

EXAMPLE 1
</div>

5

(2S)-2-(3-Acetylamino-3-isopropyl-2-oxo-pyrrolidin-1-
yl)-N-[(1S,2R)-1-(3,5-difluoro-benzyl)-2-hydroxy-3-(3-
methoxy-benzylamino)-propyl]-4-phenyl-butyramide.
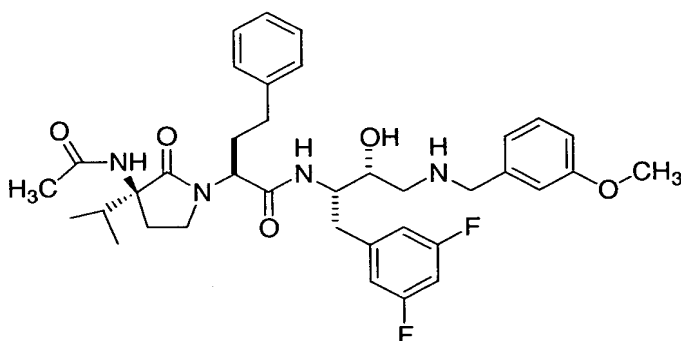
10

Step (1a): Cs_2CO_3 was added to a stirred solution
of N-Cbz-Valine (7.03 g, 27.9 mmol) in dry DMF (45
mL). The reaction mixture was stirred for 15 min

15 after which allyl bromide (5.0 ML, 56 mmol) was added
dropwise. The reaction mixture was stirred overnight
and then filtered through celite. The filtrate was
diluted with ethyl acetate, washed with water (3 ×)
and brine (1 ×). The organic phase was dried over

20 $MgSO_4$, filtered and concentrated *in vacuo*. The
residue was purified by silica gel column
chromatography to yield 8.1 g (99%) allyl ester. ESI
$(M+Na)^+$ = 314.3. ^{1}H-NMR($CDCl_3$) δ 7.33 (m, 5H), 5.88 (m,
1H), 5.49 (d, j = 8.8 Hz, 1H), 5.35-5.16 (m, 2H), 5.10

25 (s, 2H), 4.62 (m, 2H), 4.36-4.31 (m, 1H), 2.20-2.15
(m, 1H), 0.96 (d, J = 7 Hz, 3H), 0.89 (d, J = 7 Hz,
3H).

Step (1b): A freshly prepared LDA solution
(14.31 mmol) was added to a stirred mixture of allylic
ester from (1a) (1.39 g, 4.77 mmol) and $ZnCl_2$ (10.5
mL, 5.25 mmol) in 20 mL THF at -20 °C. The mixture was
5   allowed to warm up to room temperature overnight. The
reaction solution was diluted with ether and
hydrolyzed with 1 N hydrochloric acid. The aqueous
phase was extracted with ether. The combined ether
extracts were washed with brine, dried over $MgSO_4$,
10   filtered and concentrated. The residue was purified
by silica gel chromatography to provide acid 0.39 g
(28%) of viscous oil. ESI $(M-H)^- = 290.2$.

Step (1c): Acid from (1b) (200 mg, 0.686 mmol),
15   DIEA (0.385 mL, 2.74 mmol) PyBOP (713 mg, 3.19 mmol)
were mixed in 5 mL of CH_2Cl_2 and stirred for 5 min.
homo-Phe methyl ester (265 mg, 1.372 mmol) was then
added and the mixture was stirred for over night. The
reaction solution was concentrated *in vacuo*. The
20   residue was purified by silica gel chromatography to
provide amide 0.60 g (75%). ESI $(M+H)^+= 467.4$;
$(M+Na)^+= 489.3$. ^{1}H-NMR($CDCl_3$) $\delta 7.36-7.15$ (m, 10H),
5.72-5.70 (m, 1H), 5.14-5.09 (m, 3H), 4.65-4.58 (m,
1H), 3.71 (s, 3H), 2.95-1.95 (m, 9H), 1.01-0.94 (dd, J
25   = 5.5 Hz, 6H).

Step (1d): Ozone was bubbled through a solution
of amide from (1c) in 10 mL of CH_2Cl_2 (0.83 g, 1.78
mmol) at -78 °C until a blue color persisted.
30   Residual ozone was removed with a stream of oxygen.
Triphenyl phosphine (0.70 g, 2.67 mmol) was added, and
the reaction mixture was allowed to warm to rt. After

1h, the solution was concentrated under reduced pressure. The residue was purified by chromatography on silica gel to provide aldehyde 0.59 g (70%). ESI $(M+H)^+$ = 469.5. ^{1}H-NMR(CDCl$_3$) δ7.32-7.18 (m, 10H),

5   5.60-4.88 (m, 3H), 5.30 (s, 2H), 3.73-3.67 (dd, J = 7.4 Hz, 2H), 2.95-2.07 (m, 4H), 1.0-0.95 (m, 6H).

Step (1e): 6 mL of TFA/Et$_3$SiH (1:1) was added to the solution of aldehyde from (1d) (0.59 g, 1.26 mmol)

10   in 10 mL of CH$_2$Cl$_2$ at 0 °C. The mixture was stirred at 0 °C for 3h. The reaction solution was concentrated under reduced pressure. The residue was purified by chromatography on silica gel to provide lactam 0.41 g (72%). ESI $(M+H)^+$ = 453.4. ^{1}H-NMR(CDCl$_3$) δ7.36-7.17 (m,

15   10H), 5.29-4.60 (m, 3H), 3.71-3.67 (ss, 3H), 3.60-3.40 (m, 1H), 3.40-3.25 (m, 1H), 2.85-2.05 (m, 6H), 1.01-0.92 (m, 6H).

Step (1f): A solution of lactam from (1e) (350

20   mg, 0.77 mmol) in 15 mL of Methanol/Ethyl Acetate (1:1) was hydrogenated over 10% palladium on carbon (100 mg) for overnight. The solution was filtered through celite and concentrated under reduced pressure to afford the desired amine 0.23 g (95%). ESI $(M+H)^+$ =

25   319.4. ^{1}H-NMR(CDCl$_3$) δ7.32-7.16 (m, 5H), 4.79-4.70 (m, 1H), 3.70-3.68 (s, 3H), 3.50-3.40 (m, 1H), 3.30-3.20 (m, 1H), 2.65-1.20 (m, 6H), 1.35-0.86 (m, 6H).

Step (1g): A mixture of amine (0.23 g, 0.73

30   mmol) from (1f), acetic anhydride (0.5 mL, 0.53 mmol), DIEA (1.0 mL, 7.1 mmol) and DMAP (50 mg, 0.41 mmol) in 5 mL of CH$_2$Cl$_2$ were stirred at room temperature for

overnight. The reaction solution was concentrated and the residue was purified by chromatography on silica gel to provide amide 0.20 g(76%). ESI (M+H)$^+$ = 361.4; (M+Na)$^+$ = 383.4.

Step (1h): The amide (0.20 g, 0.56 mmol) from (1g) was dissolved in 10 mL of THF/H$_2$O (1:1). LiOH (100 mg, 2.44 mmol) was added and the mixture was stirred for overnight. It was then diluted with ethyl acetate and acidified with 1N HCl. The aqueous layer was extracted with 3× ethyl acetate. The combined organic layer was dried over MgSO$_4$, filtered and evaporated under reduced pressure to give acid 0.19 g (98%). ESI (M-H)$^-$ = 345.3. ^{1}H-NMR(CDCl$_3$) δ7.28-7.16 (m, 5H), 5.30 (s, 3H), 4.81-4.77 (m, 1H), 4.13-3.42 (m, 3H), 2.80-1.80 (m, 6H), 2.07 (s, 3H), 1.30-1.23 (m, 1H), 0.99-0.81 (dd, J = 6.9 Hz, 6H).

Step (1i): A mixture of acid from (1h) (15 mg, 0.043 mmol), PyBOP (24 mg, 0.018 mmol), and DIEA (20 μL, 0.14 mmol) in 4 mL of CH$_2$Cl$_2$ was stirred at room temperature for 5 min. Amine-HCl salt Preparation (B) (15 mg, 0.040 mmol) in 1 mL of CH$_2$Cl$_2$ was added and the solution was continued to stir for overnight. The reaction solution was concentrated under reduced pressure and the residue was purified on preparative LC-MS (reverse phase HPLC) to afford the desired product. ESI (M+H)$^+$ = 665.34. ^{1}H-NMR(CDCl$_3$) δ8.21-6.40 (m, 10H), 4.50-3.20 (m, 10H), 3.78 (s, 3H), 2.50-1.40 (m, 10 H), 2.02 (s, 3H), 1.07-0.90 (dd, J = 6.6 Hz, 6H).

EXAMPLE 2

(2S)-2-(3(S)-Acetylamino-3-((S)-sec-butyl)-2-oxo-
pyrrolidin-1-yl)-N-[(1S, 2R)-1-(3,5-difluoro-benzyl)-
2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-4-phenyl-
butyramide
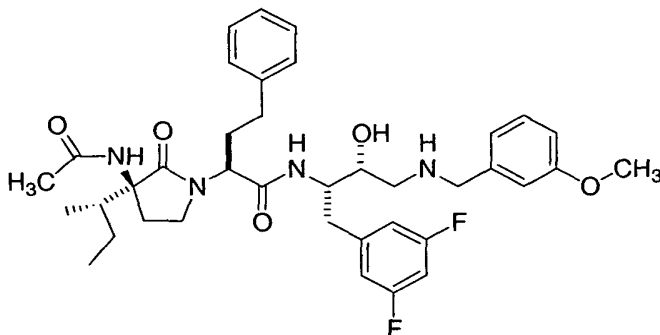
Step (2a):   L-Isoleucine (10.0 g, 76.24 mmol),
benzaldehyde (8.57 g, 76.24 mmol) and 4 Å molecular
sieve (20 g) were added to a solution of NaOH (3.05 g,
76.24 mmol) in anhydrous MeOH (100 mL). The mixture
was stirred at room temperature for overnight. After
removal of molecular sieve by filtration with celite,
the filtrate was evaporated under reduced pressure to
give a solid, which was further dried under vacuum for
8 h to give Schiff base 18.0 g (98%) as an off-white
solid. ^{1}H-NMR (DMSO-d6) δ 8.12 (s, 1H), 7.65 (m, 2H),
7.36 (m, 3H), 2.45 (m, 1H), 1.38 (m, 1H), 0.91 (m,
1H), 0.76 (m, 6H).

Step (2b):   250 mL of CH_2Cl_2 was added to the
Schiff base from (2a) (12.0 g, 49.74 mmol). The
solution was cooled to -20 °C. After which 10.7 mL
(74.61 mmol, 1.5 eq) benzyl chloroformate was added.
Stirred at -20 °C for 96 h, warmed to room
temperature, and diluted with CH_2Cl_2. The reaction

mixture was washed 2 × each with water, aq. NaHCO$_3$,

aq. sodium bisulfite and water again. The organic

layer was dried over MgSO$_4$, filtered, and the filtrate

was concentrated and the residue was purified by

chromatography on silica gel to give oxazolidinone 11

g (63%) as oil. APCI (M+H)$^+$ = 354.3. ^{1}H-NMR(CDCl$_3$) δ

7.54-7.26 (m, 10H), 6.76 (s, 1H), 5.23 (s, 2H), 4.36-

4.34 (dd, J = 5.8 Hz, 1H), 1.80 (m, 1H), 1.60-1.20 (m,

2H), 0.86-0.80 (m, 6H).

Step (2c): 570 mg (1.613 mmol) of oxazolidinone

from (2b) in 10 mL anhydrous of THF was cooled to −78

°C. Then added 0.22 mL (2.42 mmol, 1.5 eq) of allyl

iodide followed by 4.8 mL of 0.5N (2.4 mmol, 1.5 eq)

potassium bis(trimethylsilyl)amide. TLC at 60 min

showed the reaction was complete, so it was quenched

with aqueous NH$_4$Cl and warmed to room temperature.

Then the solution was diluted with water and extracted

with ethyl acetate. The combined ethyl acetate

extracts were washed with dilute aqueous NH$_4$Cl, dried

over MgSO$_4$, filtered, and the filtrate was

concentrated. The residue was purified by silica gel

chromatography to give disubstituted oxazolidinone 567

mg (89%). ESI (M+H)$^+$ = 394.4. ^{1}H-NMR(CDCl$_3$) δ 7.42-7.26

(m, 10H), 6.34 (s, 1H), 5.68-5.57 (m, 1H), 5.16-5.12

(dd, J = 9 Hz, 2H), 5.06 (s, 2H), 2.72-2.66 (m, 2H),

1.70-1.30 (m, 2H), 1.12-0.88 (m, 6H).

Step (2d): 567 mg (1.44 mmol) of disubstituted

oxazolidinone from (2c) was dissolved in 40 mL of THF-

MeOH (3 : 1). 10 mL 2N NaOH was added and the mixture

was refluxed for 2h. The THF and MeOH was evaporated,

diluted ethyl acetate and acidified with HCl. Extracted 2 × with ethyl acetate, dried the organic layer with $MgSO_4$, filtered and the filtrate was evaporated. The residue was pumped on high vacuum to

5   give crude acid 695 mg. ESI (M-H)$^-$ = 304.3.

    Step (2e): 695 mg of acid (2.27 mmol) from (2d), 15 mL of CH_2Cl_2, 488 mg HOBt (3.19 mmol, 1.4 eq) and 655 mg EDC (3.42 mmol, 1.5 eq) were mixed and stirred

10   for 5 min. 660 mg (3.42 mmol, 1.5 eq) of homo-Phe methyl ester and 0.80 mL of DIEA (5.68 mmol, 2.5 eq) were then added and the mixture was stirred for 4 h. The reaction solution was diluted with ethyl acetate and washed with 5% citric acid and 5% $NaHCO_3$, dried

15   over $MgSO_4$, filtered, and the filtrate was evaporated. The residue was purified by silica gel chromatography to provide Amide 0.53 g (76.7% for steps 2d and 2e). ESI (M+H)$^+$= 481.5. ^{1}H-NMR($CDCl_3$) δ 7.36-7.14 (m, 10H), 5.80-5.65 (m, 2H), 5.20-5.00 (m, 2H), 5.08 (s, 2H),

20   4.65-4.55 (m, 1H), 3.70 (s, 3H), 2.90-1.90 (m, 7H), 1.63-1.00 (m, 2H), 1.00-0.91 (m, 6H).

    Step (2f): Ozone was bubbled through a solution of alkene from (2e) in 10 mL of CH_2Cl_2 (0.78 g, 1.62

25   mmol) at -78 °C until a blue color persisted. Residual ozone was removed with a stream of oxygen. Triphenyl phosphine (0.60 g, 2.29 mmol) was added, and the reaction mixture was allowed to warm to rt. After 1h, the solution was concentrated under reduced

30   pressure. The residue was purified by chromatography on silica gel to provide aldehyde 0.47 g (61%). ESI (M+H)$^+$ = 483.4, (M+Na)$^+$ = 505.4.

Step (2g): 5 mL of TFA/Et₃SiH (1:1) was added to
the solution of aldehyde from (2f) (0.47 g, 0.97 mmol)
in 10 mL of CH_2Cl_2 at 0 °C. The mixture was stirred at
0 °C for 3h. The reaction solution was concentrated
under reduced pressure. The residue was purified by
chromatography on silica gel to provide lactam 0.23 g
(50%). ESI (M+H)⁺ = 467.38. ^{1}H-NMR(CDCl₃) δ7.34-7.16
(m, 10H), 5.45 (br, 1H), 5.05 (s, 2H), 4.87-4.82 (dd,
J = 4 Hz, 1H)), 3.65-3.35 (m, 2H), 3.67 (s, 3H), 2.90-
1.45 (m, 8H), 1.20-1.00 (m, 1H), 0.98-0.90 (m, 6H).

Step (2h): A solution of lactam from (2g) (225
mg, 0.48 mmol) in Methanol (15 mL) was hydrogenated
over 10% palladium on carbon (40 mg) for overnight.
The solution was filtered through celite and
concentrated under reduced pressure to afford the
desired amine. ESI (M+H)⁺ = 333.4. ^{1}H-NMR(CDCl₃) δ7.29-
7.16 (m, 5H), 4.78-4.60 (m, 3H), 3.68 (s, 3H), 3.44-
3.37 (m, 2H), 2.68-1.85 (m, 8H), 1.20-1.00 (m, 1H),
0.96-0.91 (m, 6H).

Step (2i): A mixture of acetic acid (54 μL, 0.91
mmol), HATU (348 mg, 0.92 mmol), and DIEA (257 μL,
0.91 mmol) in 5 mL of DMF was stirred at room
temperature for 5 min. Amine (152 mg, 0.46 mmol)
from (2h) in 1 mL of DMF was added and the solution
was continued to stir for overnight. The reaction
solution was diluted with ethyl acetate and washed 3 ×
with water, 1 × brine, dried the organic layer with
MgSO₄, filtered and the filtrate was evaporated. The
residue was purified by chromatography on silica gel
to provide lactam 160 mg (94%). ESI (M+H)⁺ = 375.2. ^{1}H-

NMR(CDCl$_3$) δ 7.33-7.17 (m, 5H), 6.07 (br, 1H), 4.66-4.61 (m, 1H), 3.73 (s, 3H), 3.60-3.20 (m, 2H), 2.70-1.60 (m, 8H), 1.20-1.00 (m, 1H), 0.98-0.86 (m, 6H).

5   Step (2j): Compound (0.20 g, 0.53 mmol) from (2h) was dissolved in 5 mL of THF/H$_2$O (4:1). LiOH (120 mg, 2.9 mmol) was added and the mixture was stirred for overnight. It was then diluted with ethyl acetate, acidified with 1N HCl. The aqueous layer was

10   extracted with 3× ethyl acetate. The combined organic layer was dried over MgSO$_4$, filtered and evaporated under reduced pressure to give acid 98 mg (85%). ESI (M-H)$^-$ = 359.2.

15   Step (2k): A mixture of acid from (2j) (6 mg, 0.017 mmol), HATU (7 mg, 0.018 mmol), and DIEA (7 μL, 0.05 mmol) in 2 mL of DMF was stirred at room temperature for 5 min. Amine Preparation (B) (5.6 mg, 0.017 mmol) in 1 mL of DMF was added and the solution

20   was continued to stir for overnight. The reaction solution was diluted with ethyl acetate and washed 3× with water, 1× brine, dried the organic layer with MgSO$_4$, filtered and the filtrate was evaporated. The residue was purified on preparative LC-MS (reverse

25   phase HPLC) to afford the desired product. ESI (M+H)$^+$ = 679.36. . ^{1}H-NMR(CDCl$_3$) δ 8.00 (d, J = 5Hz, 1H), 7.28-6.50 (m, 12H), 4.20-1.40 (m, 21H), 3.79 (s, 3H), 1.91 (s, 3H), 1.20-1.05 (m, 1H), 1.05-0.92 (m, 6H).

EXAMPLE 3

(2S)-2-(3(S)-Acetylamino-3-((S)-sec-butyl)-2-oxo-pyrrolidin-1-yl)-N-[(1S, 2R)-1-benzyl-2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-4-phenyl-butyramide

5
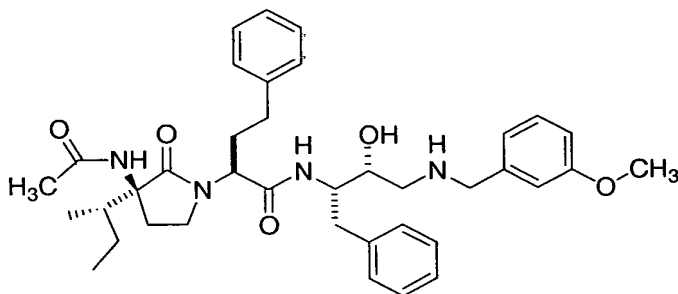
Step (3a): 20.0 mg of acid (0.056 mmol) from
(2j), 3 mL of DMF, 9.4 mg HOBt (0.061 mmol) and 11.7
10   mg EDC (0.061 mmol) were mixed and stirred for 5 min.
22 mg (0.055 mmol) of amine Preparation (D) and DIEA
(16 μL, 0.114 mmol) were then added and the mixture
was stirred for overnight. The reaction solution was
diluted with ethyl acetate and washed with 5% citric
15   acid and 5% NaHCO$_3$, dried over MgSO$_4$, filtered, and the
filtrate was evaporated. The residue was treated with
2 mL of TFA/CH$_2$Cl$_2$ (1:1) at rt for 30 min and
evaporated under reduced pressure. The residue was
purified by preparative LC-MS to give the product 9.1
20   mg (27%). ESI (M+H)$^+$ = 643.38. ^{1}H-NMR(CDCl$_3$) δ 7.91-
7.88 (d, J = 9.1 Hz, 1H), 7.28-6.88 (m, 14H), 6.24
(br, 1H), 4.20-4.00 (m, 2H), 3.79 (s, 3H), 3.41-2.00
(m, 20H), 1.91 (s, 3H), 1.80-1.05 (m, 3H), 1.02-0.87
(m, 6H).

25

EXAMPLE 4

2(S)-(3(S)-((S)-sec-butyl)-2-oxo-3-propionylamino-
pyrrolidin-1-yl)-N-(S)-{1(R)-[1-hydroxy-2-(3-methoxy-
benzylamino)-ethyl]-3-methyl-butyl}-4-phenyl-
5 butyramide.
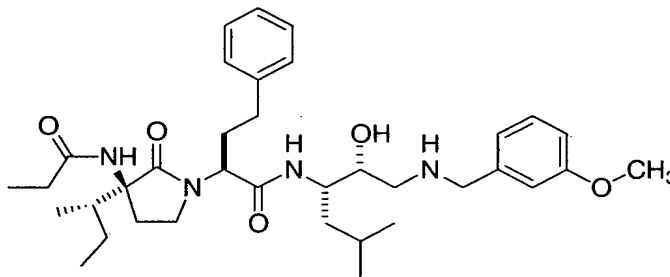
Step (4a): A mixture of propionic acid (0.12g,
10 1.61 mmol), HATU (0.57 g, 1.5 mmol), and DIEA (450 µL,
2.58 mmol) in 3 mL of DMF was stirred at room
temperature for 10 min. Amine (0.30 g, 0.90 mmol)
from (2h) in 1 mL of DMF was added and the solution
was continued to stir for overnight. The reaction
15 solution was diluted with ethyl acetate and washed 3×
with water, 1× brine, dried the organic layer with
MgSO$_4$, filtered and the filtrate was evaporated. The
residue was purified by chromatography on silica gel
to provide amide 0.26 g (74 %). ESI (M+H)$^+$ = 389.56.
20 ^{1}H-NMR(CDCl$_3$) δ 7.31-7.16 (m, 5H), 6.25 (br, 1H),
4.85-4.80 (m, 1H), 3.73-1.50 (m, 11H), 3.69 (s, 3H),
1.20-0.80 (m, 11H).

Step (4b): Compound (0.26 g, 0.67 mmol) from
25 (4a) was dissolved in 5 mL of THF. 5 mL 1N LiOH was
added and the mixture was stirred for 2h. It was then
diluted with ethyl acetate, acidified with 1N HCl.
The aqueous layer was extracted with 3× ethyl acetate.

The combined organic layer was dried over MgSO$_4$, filtered and evaporated under reduced pressure to give acid 0.21 g (84%). ESI (M+H)$^+$ = 375.59.

5   Step (4c): A mixture of acid from (4b) (37.4 mg, 0.1 mmol), HATU (76 mg, 0.2 mmol), and DIEA (70 μL, 0.4 mmol) in 2 mL of DMF was stirred at room temperature for 10 min. Amine Preparation (E) (45 mg, 0.15 mmol) in 1 mL of DMF was added and the solution

10   was continued to stir for overnight. The reaction solution was diluted with ethyl acetate and washed 3 × with water, 1 × brine, dried the organic layer with MgSO$_4$, filtered and the filtrate was evaporated. The residue was treated with 2 mL of TFA/CH$_2$Cl$_2$ (1:3) at rt

15   for 10 min and evaporated under reduced pressure. The residue was purified on preparative LC-MS (reverse phase HPLC) to afford the desired product 36.8 mg (59%). ESI (M+H)$^+$ = 623.64. . ^{1}H-NMR(CD$_3$OD) δ 8.00-6.80 (m, 9H), 4.55-1.40 (m, 22H), 3.75 (s, 3H), 1.30-0.70

20   (m, 15H).

EXAMPLE 5

(2S)-2-(3-((S)-sec-butyl)-3-(2-methoxy-acetylamino)-2-oxo-pyrrolidin-1-yl)-N-(S)-{1(R)-[1-hydroxy-2-(3-

25   methoxy-benzylamino)-ethyl]-3-methyl-butyl}-4-phenyl-butyramide
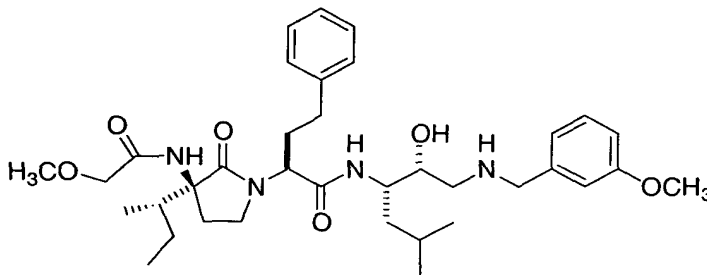
Step (5a): A mixture of methoxyacetic acid (0.14 g, 1.56 mmol), HATU (0.57 g, 1.5 mmol), and DIEA (450 μL, 2.58 mmol) in 3 mL of DMF was stirred at room temperature for 10 min. Amine (0.30 g, 0.90 mmol)

5 from (2h) in 1 mL of DMF was added and the solution was continued to stir for overnight. The reaction solution was diluted with ethyl acetate and washed 3× with water, 1× brine, dried the organic layer with MgSO$_4$, filtered and the filtrate was evaporated. The

10 residue was purified by chromatography on silica gel to provide amide 0.27 g (75 %). ESI (M+H)$^+$ = 405.53. ^{1}H-NMR(CDCl$_3$) δ 7.30-7.18 (m, 5H), 7.04 (br, 1H), 4.89- 4.84 (m, 1H), 3.98-1.80 (m, 9H), 3.92 (s, 2H), 3.69 (s, 3H), 3.43 (s, 3H), 1.65-1.10 (m, 2H), 1.05-0.90

15 (m, 6H).

Step (5b): Compound (0.27 g, 0.69 mmol) from (5a) was dissolved in 5 mL of THF. 5 mL 1N LiOH was added and the mixture was stirred for 2h. It was then

20 diluted with ethyl acetate, acidified with 1N HCl. The aqueous layer was extracted with 3× ethyl acetate. The combined organic layer was dried over MgSO$_4$, filtered and evaporated under reduced pressure to give acid 0.22 g (82%). ESI (M+H)$^+$ = 391.59.

25

Step (5c): A mixture of acid from (5b) (39 mg, 0.1 mmol), HATU (76 mg, 0.2 mmol), and DIEA (70 μL, 0.4 mmol) in 2 mL of DMF was stirred at room temperature for 10 min. Amine Exmaple E (45 mg, 0.15

30 mmol) in 1 mL of DMF was added and the solution was continued to stir for overnight. The reaction solution was diluted with ethyl acetate and washed 3 ×

with water, 1 × brine, dried the organic layer with MgSO$_4$, filtered and the filtrate was evaporated. The residue was treated with 2 mL of TFA/CH$_2$Cl$_2$ (1:3) at rt for 10 min and evaporated under reduced pressure. The residue was purified on preparative LC-MS (reverse phase HPLC) to afford the desired product 24.2 mg (34%). ESI (M+H)$^+$ = 640.63. ^{1}H-NMR(CD$_3$OD) δ 8.00-6.80 (m, 9H), 4.20-1.00 (m, 22H), 4.10 (s, 2H), 3.75 (s, 3H), 3.35 (s, 3H), 1.00-0.70 (m, 12H).

EXAMPLE 6

(2S)-2-(3(R)-Acetylamino-3(-cyclopropylmethyl)-2-oxo-pyrrolidin-1-yl)-N-[(1S, 2R)-1-benzyl-2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-4-phenyl-butyramide
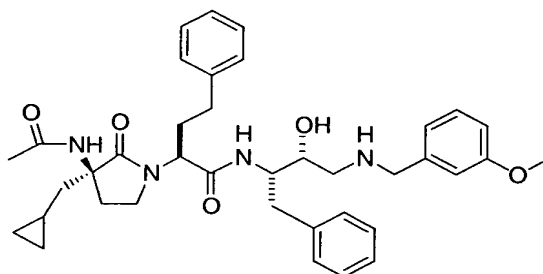
Step (6a): To 3.2 mL of diisopropylamine (22.75 mmol) in 30 mL of dry THF at -70 °C was added 9.1 mL of a 2.5 M solution of n-BuLi in hexanes (22.75 mmol), and the LDA thus formed was stirred for 15 min at that temperature. Separately, a 4.86 g portion (19 mmol) of tert-butyl 2-tert-butyl-4-methoxy-2,5-dihydro-1,3-imidazole-1-carboxylate (Boc-BDI, Seebach, D. and Hoffmann, M. (1998), *European Journal of Organic Chemistry* (7), 1337-1351.) was dissolved in 30 mL of dry THF and was also chilled to -70 °C. The LDA was

added via cannula to the Boc-BDI solution and the
reaction solution was stirred for 40 min. Allyl
iodide (1.82 mL, 19 mmol, freshly purified over
alumina) was then added and the reaction solution was

5 stirred at -70 °C for an additional hour. After this
time, an additional 9.10 mL (22.75 mmol) of n-BuLi was
added and reaction solution was stirred for 30 min.
Cyclopropylmethyl bromide (1.83 g, 19 mmol, freshly
purified over alumina) was then added and the reaction

10 solution was brought to 0 °C and stirred at that
temperature for 1 h. A saturated aqueous ammonium
chloride solution (20 mL) was then added, and ether
(200 mL) was added. The organic layer was separated
and the aqueous layer was extracted with one

15 additional equal portion of ether. The commined
organic layers were dried and concentrated to an oil,
which was purified by chromatography eluting with 5%
ethyl acetate in hexanes to afford a 73% yield (4.85
g)of the desired product. ESI MS, $(M+H)^+ = 351.6$

20

Step (6b): The imidate from step (6a) (3.5 g, 10
mmol) was dissolved in 50 mL of a 9:1 solution of
CH_2Cl_2 and TFA. The reaction solution was stirred at
rt for 7 h. The solution was then neutralized with a

25 satd. $NaHCO_3$ solution and 300 mL of CH_2Cl_2 was added.
The organic layer was removed and the aqueous layer
was extracted with an additional portion of CH_2Cl_2.
The combined organic layers were dried and
concentrated to an oil which was carried on to step

30 (6c) without further purification.

Step (6c): The oil from step (6b) was dissolved in 300 mL of THF and 30 mL of a 15% solution of TFA in water was added. The reaction solution was stirred at rt for 4 days. The mixture was then extracted with

5   ether (30 mL) and the ether was discarded. The aqueous layer was neutralized to pH 10 with a 10% aqueous solution of ammonium hydroxide, and the resulting solution was extracted 3X with 100 mL of ether. The combined organic layers were dried and

10   concentrated to an oil (1 g, 54% from step (6b)) which was carried on to step (6d) without further purification. ESI MS, $(M+H)^+ = 184.5$

Step (6d): To 1 g of the amine from step (6c)

15   (5.46 mmol) dissolved in 10 mL of CH_2Cl_2 was added 4 mL of a 2N solution of NaOH, followed by benzyl chloroformate (1.2 mL, 8.2 mmol). The reaction solution was stirred at rt for 18 h and then was diluted with 300 mL of CH_2Cl_2 and a satd. $NaHCo_3$

20   solution. The organic layer was separated, dried, and concentrated to an oil. Purification by chromatography eluting with 10-20% ethyl acetate in hexanes provided the protected quaternary amino acid ester(1.6 g, 92%). ESI MS, $(M+H)^+ = 318.5$

25

Step (6e): The ester from step (6d) 1.4 g (4.4 mmol) was dissolved in 25 mL of methanol and 1.85 g of LiOH (44 mmol) dissolved in 14 mL of water was added. The solution was transferred in to 11 3.5 mL microwave

30   tubes and heated with a microwave to 120 °C for 360 seconds. The samples were then combined, and the methanol was removed by rotary evaporation. The

reaction solution was diluted with 50 mL of water and 20 mL of ethyl acetate. The organic layer was separated and discarded. The aqueous layer was brought to pH 3 with 1N HCl and extracted with 2 X 100 mL of ethyl acetate. The combined organic layers were separated, dried, and concentrated to an oil (1.17 g, 86%) which was taken on without further purification. ESI MS, $(M+H)^+ = 304.4$.

Step (6f): The acid from step (6e) (1.17g, 3.86 mmol) was dissolved in 30 mL of CH_2Cl_2 and 15 mL of DMF. Homophenylalanine methyl ester (0.89 g, 4.6 mmol) was added, followed by DIEA (2.7 mL, 15.4 mmol) and PyBOP (2.41 g, 4.6 mmol). The reaction solution was stirred at rt for 18 h, then diluted with water (20 mL) and extracted with two 200 mL portions of CH_2Cl_2. The combined organic layers were separated, dried, and concentrated to an oil. Purification by chromatography eluting with 20-50% ethyl acetate in hexanes provided 1.45 g (78%) of the desired product. ^1H NMR (δ, representative) (7.1-7.4, m, 10 H, 2 phenyl groups), 5.6-5.75 (m, 2H, alkene), 3.67 (s, 3H, methyl ester).

Step (6g): 0.5 g (1.04 mmol) of the compound from step (6f) was dissolved in 10 mL of CH_2Cl_2 and chilled to -70 °C. Ozone was introduced by bubbling through the solution until a blue color persisted. Dioxygen was then bubbled through the solution until the blue color dissipated. The reaction solution was then treated with triphenylphosphine (0.33 g, 1.25 mmol) and the solution was brought to rt and stirred

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for 2 h. The reaction solution was then cooled to 0 °C and 10 mL of a 1:1 solution of TFA and triethylsilane was added. The reaction solution was srirred for 3 h, and then the solvent was removed. The resulting

5   product was purified by chromatography eluting with 10-20 ethyl acetate in hexanes to provide 0.150 g of the desired lactam (31%). ESI MS, (M+H)$^+$ = 465.5.

Step (6h): The lactam from step (6g) was

10   dissolved in 20 mL of methanol and 30 mg of 10% palladium on carbon was added. The reaction solution was placed in a Parr apparatus under 50 psi of dihydrogen and shaken for 2 h. The catalyst was then removed by filtration and the reaction solution was

15   concentrated to an oil (0.10 g, 100%) and taken on without further purification

Step (6i): The amine from step (6h) (0.10 g,

20   0.30 mmol) was dissolved in 5 mL of DMF. Acetic acid (22 μL, 0.36 mmol), HATU (0.14 g, 0.36 mmol) and DIEA (0.2 mL, 1.2 mmol) were added and the reaction solution was stirred at rt for 18 h. The reation solution was then diluted with 5 mL of water and

25   extracted with 2 X 25 mL of ethyl acetate. The combined organic layers were washed with a 1N HCl solution (5 mL) and a satd. NaHCO3 solution (5 mL), dried, and concentrated to an oil. The product was purified by chromatography eluting with 20-50% ethyl acetate in hexanes to provide 2 separate diastereomers

30   at the lactam stereocenter (diastereomer 1, 20 mg, diastereomer 2, 30 mg, 50%). ESI MS, (M+H)$^+$ = 373.5

Step (6j): The lower-eluting diastereomer from step (6i) (30 mg, 0.080 mmol) was dissolved in 3 mL of THF and a solution of 33 mg (0.80 mmol) of lithium hydroxide dissolved in 0.5 mL of water was added. After stirring at rt for 3 h, 5 mL each of water and ethyl acetate were added and the organic layer was separated and discarded. The aqueous layer was brought to pH 3 with 1N HCl and the reaction solution was extracted with ethyl acetate (2 X 30 mL). The combined organic layers were separated, dried, and concentrated to an oil (18 mg, 94%) which was taken on without further purification. ESI MS, $(M+H)^+ = 359.5$

Step (6k): To 18 mg (0.05 mmol) of the acid from step (6j) in 2 mL of DMF was added 24 mg (0.055 mmol) of intermediate 6 dissolved in 1 mL of DMF. HATU (22 mg 0.055 mmol) and DIEA (17 µL, 0.1 mmol) were added, and the reaction solution was stirred at rt for 18 h. The solution was then diluted with water and extracted with 2 X 25 mL of ethyl acetate. The combined organic layers were washed with a 1N HCl solution (5 mL) and a satd. NaHCO$_3$ solution (5 mL), dried, and concentrated to an oil. The product was purified by chromatography eluting with 80-100% ethyl acetate in hexanes to provide 20 mg (50%) of the desired product. ESI MS, $(M+H)^+ = 775.8$

Step (6l): The compound from step (6k) (20 mg, 0.025 mmol) was dissolved in 8 mL of methanol and 13 mg of 5% palladium on carbon was added. The reaction solution was place in a Parr apparatus and hydrogenated at 50 psi for 3 h. The catalyst was removed by filtration and the resulting solution was

concentrated to an oil. The product was purified by
chromatography eluting with 10% methanol in ethyl
acetate to provide 10 mg (62%) of the desired product.
ESI MS, $(M+H)^+$ = 641.6. ^1H NMR (δ) (7.05-7.26, m,
14H), (6.88, s, 1H), (6.22, s, 1H), (4.15-4.2, m, 1H),
(4.09, q, 1H, J = 7.0), (3.77, s, 3H), (3.6-3.7, m,
1H), (3.12-3.18, m, 1H), (3.08, q, 1H, J = 8.7),
(2.94, dd, 1H, J = 4.1, 0.8), (2.71, d, 2H, J = 5.1),
(2.1-2.4, m, 11H), (1.98, s, 2H), (1.63, dd, 1H, J =
5.5, 1.0), (1.48, dd, 1H, J = 7.7, 1.0), (1.23, t, 1H,
J = 6.9), (0.62-0.67, m, 1H), (0.56, d, 2H, J = 7.4),
(0.15, m, 2H).

EXAMPLE 7

(2S)-2-(3(S)-Acetylamino-3(-cyclopropylmethyl)-2-oxo-
pyrrolidin-1-yl)-N-[(1S, 2R)-1-benzyl-2-hydroxy-3-(3-
methoxy-benzylamino)-propyl]-4-phenyl-butyramide
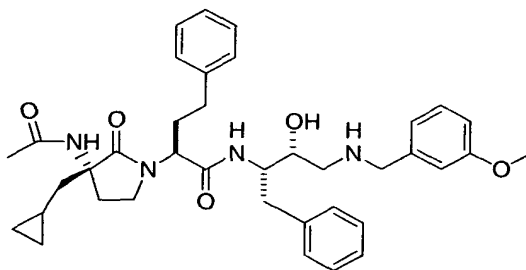
Following the general procedure for steps (6j-
l)but using the higher eluting diastereomer from step
(6i), the title compound was prepared. ESI MS, $(M+H)^+$
= 641.6.

EXAMPLE 8

(2S)-2-(3(S)-(2(S)-amino-5-carboxypentanoylamino)-3-
((S)-sec-butyl)-2-oxo-pyrrolidin-1-yl)-N-[(1S,2R)-1-
benzyl-2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-4-
phenyl-butyramide
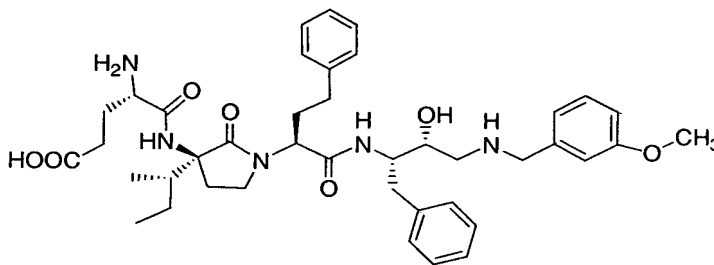
Step (8a): The lactam from step (2g) (2.0 g,
4.28 mmol) was dissolved in 100 mL of a 1:1 solution
of THF and a 1 M lithium hydroxide solution. After 2 h
at rt the reaction solution was concentrated to remove
THF, and the resulting aqueous solution was acidified
with HCL and extracted with ethyl acteate. The
organic layers were combined, dried, and concentrated.
The crude acid was purified by HPLC using the
described standard conditions to provide 0.75 g (40%)
of the purified material., ESI MS, $(M+H)^+ = 453.49$.

Step (8b): The acid from step (8a) (1.06 g, 2.35
mmol) was dissolved in 20 mL of DMF and treated with
1.64 mL (9.4 mmol) of DIEA and 2.31 g (6.1 mmol) of
HATU. After the solution was stirred at rt for 10
min, the compound of Preparation (D) (1.0 g, 2.5 mmol)
was added and the reaction solution was stirred at rt
for 2 h. The reaction solution was diluted with water
and extracted with 3 parts of a 2:1 solution of
hexanes/DCM. The combined organic layers were dried
and concentrated and the desired amide was isolated by

chromatography eluting with a gradient of 40-60% ethyl acetate in hexanes to provide 1.99 g (97%). ESI MS, $(M+H)^+$ = 835.31.

5   Step (8c): A solution of the compound of Example (8b) (2.0 g, 2.34 mmol in Methanol (50 mL) was placed in a Parr apparatus and hydrogenated over 10% palladium on carbon (400 mg) at 60 psi for 24 h. The slurry was filtered through celite and the resulting
10   solution was concentrated under reduced pressure to afford 1.5 g (91%) of the desired amine. ESI MS, $(M+H)^+$ = 701.46.

     Step (8d): A solution of Boc-Glu(O-t-Bu)-OH
15   (Bachem, 42 mg, 0.136 mmol) dissolved in DMF (0.6 mL) was treated with DIEA (63 μL, 0.36 mmol) and HATU (52 mg, 0.136 mmol). After 10 min, the amine from step (8c) (38 mg, 0.045 mmol) was added and the reaction solution was stirred at rt overnight. Water was then
20   added, and the solution was extracted with 3 portions of ethyl acteate. The organic layers were combined and concentrated to a crude product, which was then dissolved in a 1:1 solution of TFA and DCM. After 2 h at rt, the solvents were removed and the residue was
25   purified by preparative HPLC under the described conditions to provide 4.0 mg (12%) of the title compound of Example (8). ESI MS (M+H)+ = 730.52. ^{1}H NMR (300 MHz, CD$_3$OD) δ ppm 0.55 (d, J=6.59 Hz, 3 H), 0.95 (m, 5 H), 1.59 (m, 2 H), 1.91 (m, 2 H), 2.05 (m,
30   4 H), 2.51 (m, 6 H), 2.61 (s, 2 H), 2.82 (dd, J=12.81, 8.42 Hz, 2 H), 2.98 (m, 2 H), 3.15 (dd, J=14.28, 3.30 Hz, 2 H), 3.44 (d, J=3.66 Hz, 1 H), 3.74 (s, 2 H), 3.91 (m, 3 H), 4.13 (s, 3 H), 4.32 (dd, J=10.43, 4.94

Hz, 1 H), 6.85 (dd, J=8.06, 2.20 Hz, 1 H), 6.95 (d, J=7.69 Hz, 1 H), 6.99 (d, J=1.83 Hz, 1 H), 7.17 (m, 11 H), 8.01 (d, J=8.79 Hz, 1 H).

5 **EXAMPLE 9**

(2S)-2-(3(S)-(2-methoxy-acetylamino)-3-((S)-*sec*-butyl)-2-oxo-pyrrolidin-1-yl)-N-[(1S, 2R)-1-benzyl-2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-4-phenyl-butyramide
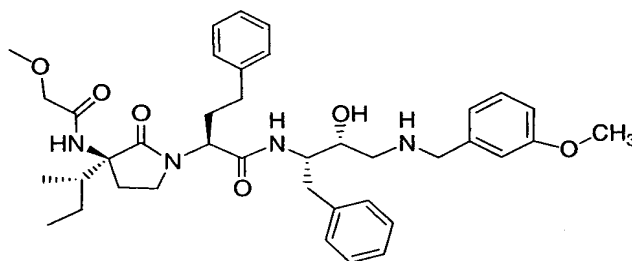

10

In a similar manner to the synthesis of the compound of Example (8), the amine from the compound of Example (8c) (55 mg, 0.1 mmol) was reacted with

15 methoxyacetic acid and the Boc group was removed with 1:1 TFA/DCM to provide the compound of Example (9) (27 mg, 53%). ESI MS $(M+H)^+$ = 673.42

^{1}H NMR (300 MHz, CD$_3$OD) δ ppm 0.76 (d, J=6.59 Hz, 3 H),

20 0.95 (t, J=7.14 Hz, 3 H), 1.68 (s, 2 H), 1.90 (d, J=4.03 Hz, 1 H), 2.10 (m, 3 H), 2.50 (m, 3 H), 2.84 (m, 2 H), 3.10 (m, 2 H), 3.17 (dd, J=14.28, 3.66 Hz, 1 H), 3.34 (s, 3 H), 3.69 (d, J=2.20 Hz, 2 H), 3.75 (s, 3 H), 3.85 (m, 4 H), 4.11 (m, 4 H), 6.90 (dd, J=8.42,

25 2.56 Hz, 1 H), 6.96 (m, 2 H), 7.16 (m, 11 H), 7.68 (s, 1 H), 8.01 (d, J=8.79 Hz, 1 H).

EXAMPLE 10

(2S)-2-(3(S)-propionylamino-3-((S)-_sec_-butyl)-2-oxo-pyrrolidin-1-yl)-_N_-[(1S, 2R)-1-benzyl-2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-4-phenyl-butyramide
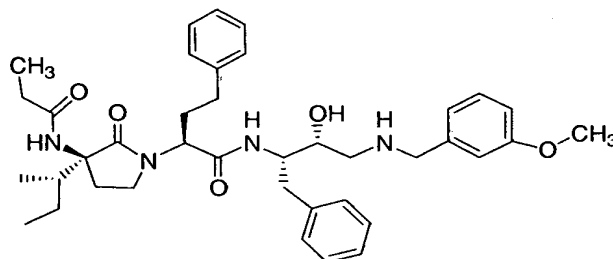
5

In a similar manner to the synthesis of the compound of Example (8), the amine from the compound
10 of Example (8c) (55 mg, 0.1 mmol) was reacted with propionic acid and the Boc group was removed with 1:1 TFA/DCM to provide the compound of Example (10) (29 mg, 58%). ESI MS (M+H)$^+$ = 657.44

15 ^{1}H NMR (300 MHz, Solvent) δ ppm 0.74 (d, _J_=6.96 Hz, 3 H), 0.95 (t, _J_=6.96 Hz, 3 H), 1.05 (t, _J_=7.51 Hz, 3 H), 1.63 (m, 2 H), 1.87 (m, 1 H), 2.07 (m, 2 H), 2.22 (m, 3 H), 2.45 (m, 1 H), 2.61 (m, 2 H), 2.85 (dd, _J_=12.63, 8.60 Hz, 2 H), 3.04 (m, 1 H), 3.11 (dd,
20 _J_=14.28, 3.30 Hz, 1 H), 3.22 (dd, _J_=9.15, 3.30 Hz, 1 H), 3.74 (s, 1 H), 3.74 (s, 3 H), 3.82 (dd, _J_=9.89, 4.76 Hz, 2 H), 3.90 (m, 1 H), 4.08 (m, 2 H), 4.14 (s, 2 H), 6.90 (dd, _J_=8.42, 2.56 Hz, 1 H), 6.96 (d, _J_=7.69 Hz, 1 H), 6.99 (d, _J_=2.20 Hz, 1 H), 7.17 (m, 11 H),
25 7.93 (s, 1 H), 8.09 (d, _J_=8.42 Hz, 1 H).

EXAMPLE 11

(2S)-2-(3(S)-acetylamino-3-((S)-*sec*-butyl)-2-oxo-pyrrolidin-1-yl)-*N*-[(1S, 2R)-1-(2-fluorobenzyl)-2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-4-phenyl-butyramide

The acid of Example (2j) (5.0 mg, .014 mmol) was dissolved in 1 mL of DCM and coupled with the amine of Preparation (G) (20 mg, 0.06 mmol) using EDCI (0.06 mmol, 10 mg). After 2 h at rt, the solvent was removed and the crude compound was purified by preparative HPLC under the described standard conditions to provide 0.5 mg (7%) of the desired title compound of Example (11). ESI MS $(M+H)^+$ = 661.4

EXAMPLE 12

(2S)-2-(3(S)-acetylamino-3-((S)-*sec*-butyl)-2-oxo-pyrrolidin-1-yl)-*N*-[(1S, 2R)-1-(2-fluorobenzyl)-2-hydroxy-3-(3-chloro-benzylamino)-propyl]-4-phenyl-butyramide

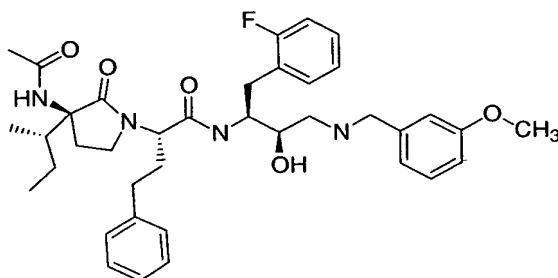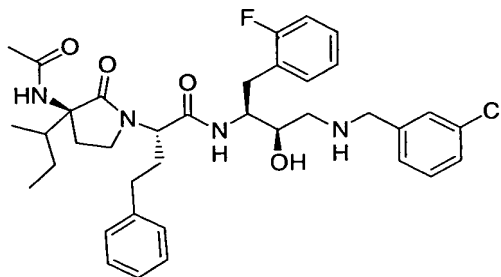The acid of Example (2j) (5.0 mg, .014 mmol) was dissolved in 1 mL of DCM and coupled with the amine of Preparation (H2) (20 mg, 0.06 mmol) using EDCI (0.06 mmol, 10 mg). After 2 h at rt, the solvent was

5 removed and the crude compound was purified by preparative HPLC under the described standard conditions to provide 0.5 mg (7%) of the desired title compound of Example (12). ESI MS $(M+H)^+$ = 665.4

10   **EXAMPLE 13**

(2S)-2-(3(S)-ethoxycarbonylamino-3-((S)-sec-butyl)-2-oxo-pyrrolidin-1-yl)-N-[(1S, 2S)-1-benzyl-2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-4-phenyl-butyramide
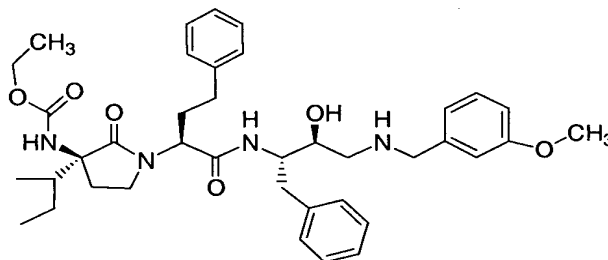
15

Step (13a): The lactam of Example (8a) (1.0 g, 2.34 mmol) was dissolved in 20 mL of DMF and treated with 1.6 mL (9.4 mmol) of DIEA and 2.3 g (6 mmol) of

20 HATU. After the solution was stirred at rt for 10 min, the compound of Preparation (I) (1 g, 2.5 mmol) was added and the reaction solution was stirred at rt for 2 h. The reaction solution was diluted with water and extracted with 3 parts of a 2:1 solution of

25 hexanes/DCM. The combined organic layers were dried and concentrated and the desired amide was isolated by chromatography eluting with a gradient of 40-60% ethyl acetate in hexanes to provide 1.98 g (96%) of the

desired amide as a 2:1 mixture of diastereoisomers. ESI MS, $(M+H)^+$ = 835.31.

Step (13b) A solution of the compound of Example (8b) (1.96 g, 2.25 mmol in Methanol (50 mL) was placed in a Parr apparatus and hydrogenated over 10% palladium on carbon (400 mg) at 60 psi for 24 h. The slurry was filtered through celite and the resulting solution was concentrated under reduced pressure to afford 1.5 g (91%) of the desired amine as a 2:1 mixture of diastereoisomers. ESI MS, $(M+H)^+$ = 701.46.

Step (13c) The amine from step (13b) (60 mg, 0.085 mmol) was dissolved in 4 mL of CH_2Cl_2 and ethyl chloroformate (50 μL, 0.5 mmol) and DIEA (0.2 mL, 1.14 mmol) were added. After stirring at rt for 2 h, the reaction solution was treated with approximately 200 mg of polymer-bound trisamine resin (Argonaut Technologies) to remove excess chloroformate. The reaction solution was removed by filtration and the resin was washed with 3 additional 2 mL portions of DCM. The combined organic layers were dried and concentrated to provide a crude product which was treated directly with 2 mL of 1:1 TFA/DCM solution at rt for 1 h. Removal of the solvents and purification by prep HPLC under the standard reported conditions provided the compound of Example (13) (5.4 mg, 8%) as the earliest eluting diastereomer. ESI MS $(M+H)^+$ = 673.32 ^{1}H NMR (500 MHz, Chloroform-d) δ ppm 0.93 (d, J=6.71 Hz, 3 H), 0.98 (d, J=7.02 Hz, 3 H), 1.16 (t, J=7.02 Hz, 3 H), 1.38 (d, J=6.71 Hz, 1 H), 1.43 (d, J=6.71 Hz, 1 H), 1.63 (d, J=13.73 Hz, 2 H), 1.85 (m, 5 H), 2.18 (m, 3 H), 2.43 (m, 2 H), 2.82 (dd, J=12.36,

4.12 Hz, 1 H), 2.95 (m, 3 H), 3.12 (m, 1 H), 3.32 (td, J=9.46, 3.97 Hz, 1 H), 3.76 (s, 3 H), 3.91 (m, 1 H), 4.07 (m, 3 H), 5.19 (s, 1 H), 6.87 (dd, J=8.39, 2.29 Hz, 1 H), 6.92 (d, J=7.32 Hz, 1 H), 7.02 (s, 1 H), 7.08 (d, J=7.02 Hz, 1 H), 7.22 (m, 10 H), 7.62 (d, J=7.93 Hz, 1 H).

EXAMPLE 14

(2S)-2-(3(S)-ethoxycarbonylamino-3-((S)-sec-butyl)-2-oxo-pyrrolidin-1-yl)-N-[(1S, 2R)-1-benzyl-2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-4-phenyl-butyramide
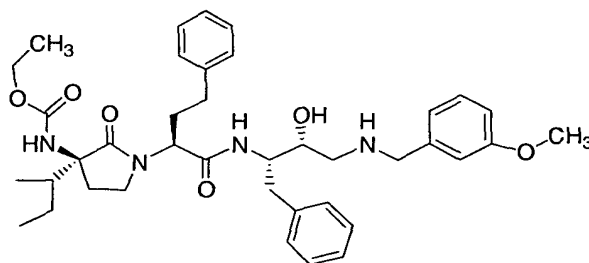

Purification of the compound of step (13c) by prep HPLC under the standard reported conditions provided the compound of Example (14) as the major, slower-eluting diastereomer (16.6 mg, 28%). ESI MS (M+H)$^+$ = 673.4. ^{1}H NMR (500 MHz, Chloroform-d) δ ppm 0.89 (d, J=6.71 Hz, 3 H), 0.97 (t, J=7.32 Hz, 3 H), 1.11 (m, 1 H), 1.18 (t, J=7.02 Hz, 3 H), 1.60 (m, 1 H), 1.69 (m, 1 H), 2.10 (m, 4 H), 2.21 (m, J=5.80 Hz, 1 H), 2.35 (m, 1 H), 2.42 (m, 1 H), 2.68 (dd, J=13.73, 10.07 Hz, 1 H), 2.82 (s, 1 H), 3.02 (m, 2 H), 3.15 (m, 2 H), 3.26 (d, J=14.34 Hz, 1 H), 3.51 (s, 1 H), 3.74 (d, J=4.58 Hz, 1 H), 3.76 (s, 3 H), 3.90 (m, 3 H), 4.06 (m, 2 H), 5.11 (s, 1 H), 6.87 (dd, J=8.24, 2.14

Hz, 1 H), 6.93 (d, *J*=7.32 Hz, 1 H), 7.04 (d, *J*=7.32

Hz, 2 H), 7.09 (m, 1 H), 7.22 (m, 9 H), 7.60 (s, 1 H).

EXAMPLE 15

5 (2S)-2-(3(S)-methoxycarbonylamino-3-((S)-sec-butyl)-2-
oxo-pyrrolidin-1-yl)-N-[(1S, 2S)-1-benzyl-2-hydroxy-3-
(3-methoxy-benzylamino)-propyl]-4-phenyl-butyramide
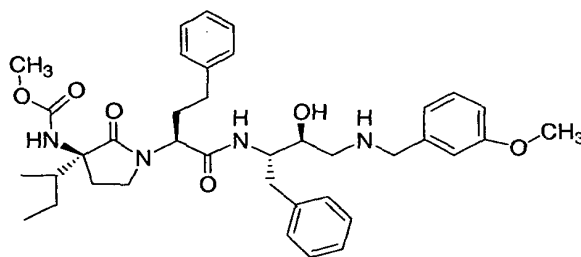
10

In a manner analogous to the synthesis of the
compound of Example (13), but using methyl
chloroformate, the title compound of Example (15) was
prepared as the faster eluting diastereomer (3.3 mg,

15 4%). ESI MS (M+H)$^{+}$ = 659.29. ^{1}H NMR (300 MHz, MeoH-d)
δ ppm 0.99 (m, 10 H), 1.29 (s, 2 H), 1.74 (m, *J*=5.86
Hz, 4 H), 2.37 (d, *J*=8.42 Hz, 1 H), 2.79 (m, 2 H),
2.92 (m, 1 H), 3.03 (m, 2 H), 3.58 (s, 3 H), 3.59 (m,
2 H), 3.78 (s, 3 H), 3.80 (m, 2 H), 3.97 (m, 1 H),

20 4.15 (d, *J*=4.39 Hz, 1 H), 4.23 (m, 1 H), 6.99 (d,
J=8.42 Hz, 2 H), 6.98 (m, 2 H), 7.28 (m, 10 H), 8.01
(s, 1 H).

EXAMPLE 16

25 (2S)-2-(3(S)-methoxycarbonylamino-3-((S)-sec-butyl)-2-
oxo-pyrrolidin-1-yl)-N-[(1S, 2R)-1-benzyl-2-hydroxy-3-
(3-methoxy-benzylamino)-propyl]-4-phenyl-butyramide
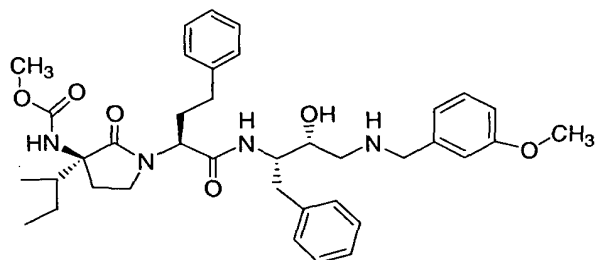
Purification of the compound of Example (15) by
prep HPLC under the standard reported conditions

5   provided the compound of Example (16) as the major,
slower-eluting diastereomer (17.6 mg, 29%). ESI MS
(M+H)$^+$ = 659.4 ^{1}H NMR (300 MHz, MeOH-d) δ ppm 0.72 (d,
J = 7.34 Hz, 3 H), 0.95 (t, J=6.96 Hz, 3 H), 1.04 (m,
1 H), 1.65 (m, 2 H), 1.86 (m, 1 H), 2.12 (t, J=7.32

10   Hz, 2 H), 2.17 (m, 1 H), 2.47 (m, 1 H), 2.66 (m, 3 H),
2.88 (m, 2 H), 3.06 (m, 1 H), 3.15 (d, J=2.93 Hz, 1
H), 3.20 (m, 1 H), 3.25 (d, J=8.06 Hz, 1 H), 3.58 (s,
3 H), 3.78 (s, 3 H), 3.88 (dd, J=8.24, 2.75 Hz, 2 H),
3.97 (m, 2 H), 4.17 (s, 2 H), 6.92 (dd, J=8.24, 2.38

15   Hz, 1 H), 7.01 (m, 2 H), 7.19 (m, 11 H), 8.01 (d,
J=8.06 Hz, 1 H).

EXAMPLE 17

(2S)-2-(3(S)-propylureido-3-((S)-sec-butyl)-2-oxo-

20   pyrrolidin-1-yl)-N-[(1S, 2R)-1-benzyl-2-hydroxy-3-(3-
methoxy-benzylamino)-propyl]-4-phenyl-butyramide
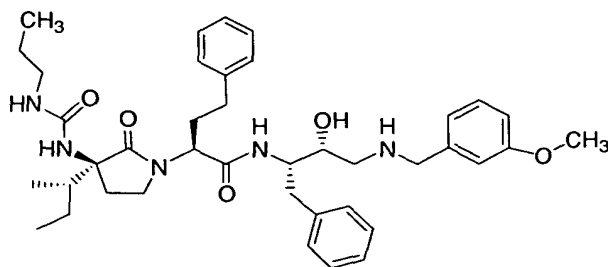
The amine from step (13b) (60 mg, 0.085 mmol) was dissolved in 1.5 mL of THF and propyl isocyanate (1000 µL, 10 mmol) was added. After stirring at rt for 48 h, the reaction solution was treated with 2 mL of DCM

5 and approximately 200 mg of polymer-bound trisamine resin (Argonaut Technologies) to remove excess isocyanate. The reaction solution was removed by filtration and the resin was washed with 3 additional 2 mL portions of DCM. The combined organic layers

10 were dried and concentrated to provide a crude product which was purified by Prep HPLC under the standard conditions described to provide 19 mg (30%) of the desired title compound of Example (17). ESI MS (M+H)$^+$ = 686.37. ^{1}H NMR (300 MHz, Methanol-D) δ ppm 0.79 (d,

15 J=6.59 Hz, 3 H), 0.85 (t, J=7.32 Hz, 3 H), 0.95 (t, J=7.14 Hz, 3 H), 1.06 (d, J=2.20 Hz, 1 H), 1.28 (m, 1 H), 1.33 (dd, J=6.59, 2.56 Hz, 1 H), 1.42 (m, 2 H), 1.60 (m, 3 H), 2.09 (m, 3 H), 2.30 (m, 1 H), 2.46 (m, 1 H), 2.69 (m, 1 H), 2.84 (m, 2 H), 3.01 (t, J=6.77

20 Hz, 2 H), 3.07 (m, 1 H), 3.15 (m, 2 H), 3.69 (m, 2 H), 3.75 (s, 3 H), 3.90 (s, 3 H), 4.14 (s, 2 H), 6.91 (dd, J=8.06, 2.20 Hz, 1 H), 6.97 (m, 2 H), 7.17 (m, 11 H), 8.24 (d, J=8.05 Hz, 1 H).

25 **EXAMPLE 18**

(2S)-2-(3(S)-ethylureido-3-((S)-*sec*-butyl)-2-oxo-pyrrolidin-1-yl)-*N*-[(1S, 2R)-1-benzyl-2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-4-phenyl-butyramide

In a manner analogous to the synthesis of the compound of Example (17), but using ethyl isocyanate, the title compound of Example (18) was prepared as the major eluting diastereomer (3.3 mg, 4%). ESI MS (M+H)$^+$ = 672.39.

1H NMR (300 MHz, Methanol-D) δ ppm 0.80 (d, J=6.59 Hz, 3 H) 0.96 (t, J=7.14 Hz, 3 H) 1.03 (t, J=7.32 Hz, 3 H) 1.17 (t, J=7.14 Hz, 1 H) 1.61 (m, 2 H) 1.90 (s, 1 H) 2.10 (m, 3 H) 2.30 (m, 1 H) 2.45 (m, 1 H) 2.63 (m, 3 H) 2.87 (m, J=7.69 Hz, 1 H) 3.03 (d, J=6.96 Hz, 1 H) 3.08 (m, J=7.32 Hz, 3 H) 3.16 (m, 3 H) 3.68 (dd, J=9.89, 4.76 Hz, 1 H) 3.75 (s, 3 H) 3.88 (m, 3 H) 4.14 (s, 2 H) 6.91 (dd, J=7.87, 2.01 Hz, 1 H) 6.97 (m, 3 H) 7.18 (m, 10 H) 8.25 (d, J=7.32 Hz, 1 H)

EXAMPLE 19

(2S)-2-(3(S)-(trifluoroacetyl)amino-3-((S)-sec-butyl)-2-oxo-pyrrolidin-1-yl)-N-[(1S, 2R)-1-benzyl-2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-4-phenyl-butyramide
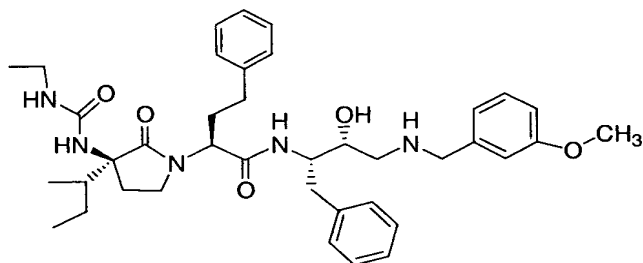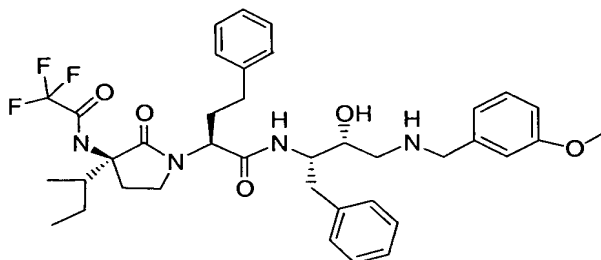
In a manner analogous to the synthesis of the
compound of Example (17), but using trifluoroacetic
anhydride, the title compound of Example (19) was
prepared as the major eluting diastereomer (21.7 mg,
30%). ESI MS (M+H)$^+$ = 697.3

^{1}H NMR (300 MHz, Methanol-D) δ ppm 8.06 (s, 1 H), 7.57
(d, J=8.42 Hz, 1 H), 7.21 (m, 10 H), 6.98 (m, 4 H),
4.45 (dd, J=11.17, 3.48 Hz, 1 H), 4.32 (dd, J=11.17,
3.11 Hz, 1 H), 4.10 (m, 2 H), 3.96 (m, 1 H), 3.86
(ddd, J=13.36, 6.96, 6.77 Hz, 1 H), 3.77 (s, 3 H),)
3.74 (dd, J=3.84, 2.01 Hz, 1 H), 2.98 (m, 3 H), 2.80
(dd, J=14.65, 12.08 Hz, 1 H), 2.40 (m, 3 H),) 2.07
(m, 1 H), 1.88 (m, 1 H), 1.49 (m, 1 H), 1.27 (dd,
J=9.15, 6.59 Hz, 6 H), 1.04 (d, J=6.59 Hz, 2 H), 0.92
(t, J=6.96 Hz, 3 H).

EXAMPLE 20

(2S)-2-(3(S)- (3-3H-imidazol-4-yl-propionylamino)-3-
((S)-sec-butyl)-2-oxo-pyrrolidin-1-yl)-N-[(1S, 2R)-1-
benzyl-2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-4-
phenyl-butyramide

A solution of deaminohistidine (100 mg, 0.71
mmol) dissolved in 2 mL of DMF was treated with DIEA
(200 µL, 1.15 mmol) and HATU (200 mg, 0.52 mmol).
After 10 min at rt, a solution of 60 mg (0.085 mmol)

of the amine of step (8b) was added and the reaction
solution was stirred at rt for 3 h. A 5 mL portion of
water was then added, and the mixture was extracted
with three 5 mL portions of ethyl acetate. The

5   organic layers were separated, dried, and
concentrated, and the crude product was treated
directly with 2 mL of 1:1 TFA/DCM solution at rt for 1
h. Removal of the solvents and purification by prep
HPLC under the standard reported conditions provided

10   the compound of Example (20) (13 mg, 20%) as the
earliest eluting diastereomer. ESI MS $(M+H)^+ = 723.4$.
^{1}H NMR (300 MHz, DEUTERIUM OXIDE) δ ppm 0.68 (d, J=6.59
Hz, 3 H), 0.95 (t, J=6.59 Hz, 3 H), 1.04 (d, J=18.68
Hz, 1 H), 1.63 (m, 3 H), 2.09 (m, 4 H), 2.45 (m, 1 H),

15   2.60 (m, 6 H), 2.86 (m, 1 H), 2.94 (m, 2 H), 3.03 (m,
1 H), 3.14 (dd, J=14.10, 3.11 Hz, 1 H), 3.22 (d,
J=7.32 Hz, 1 H), 3.33 (q, J=4.27 Hz, 1 H), 3.75 (s, 3
H), 3.84 (m, 1 H), 3.94 (m, 2 H), 4.09 (d, J=4.76 Hz,
1 H), 4.12 (d, J=5.13 Hz, 1 H), 4.16 (s, 1 H), 6.88

20   (dd, J=8.24, 1.65 Hz, 1 H), 6.99 (m, 2 H), 7.18 (m, 11
H), 8.05 (m, 2 H), 8.62 (d, J=1.46 Hz, 1 H).

EXAMPLE 21

(2S)-2-(3(S)- (3-3H-imidazol-4-yl-propionylamino)-3-
25   ((S)-*sec*-butyl)-2-oxo-pyrrolidin-1-yl)-*N*-[(1S, 2S)-1-
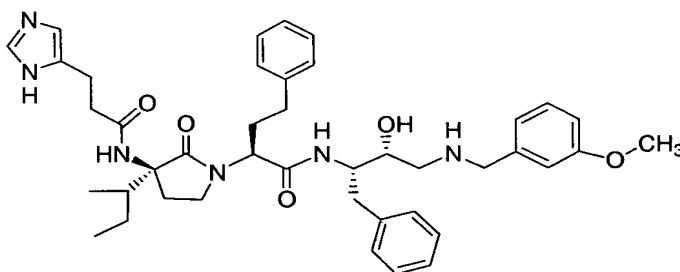benzyl-2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-4-
phenyl-butyramide
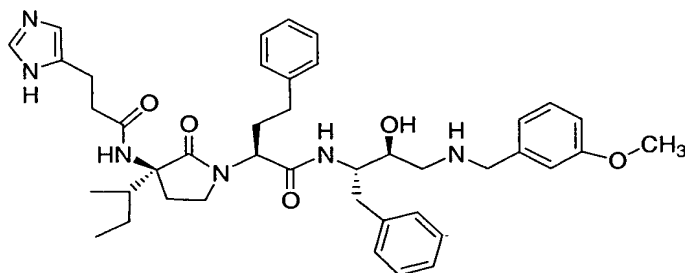
Purification of the compound of Example (20) by prep HPLC under the standard reported conditions provided the compound of Example (21) as the minor, slower-eluting diastereomer (9.0 mg, 14%). ESI MS $(M+H)^+$ = 723.4. ^{1}H NMR (500 MHz, CHLOROFORM-D) δ ppm 0.89 (d, J=6.41 Hz, 3 H), 0.94 (t, J=7.32 Hz, 3 H), 1.08 (s, 2 H), 1.60 (m, 2 H), 1.73 (m, 2 H), 2.10 (m, 3 H), 2.24 (d, J=8.24 Hz, 1 H), 2.35 (m, 1 H), 2.46 (m, 2 H), 2.64 (m, 2 H), 2.89 (m, J=5.80 Hz, 1 H), 2.96 (m, 2 H), 3.07 (d, J=15.26 Hz, 2 H), 3.21 (d, J=4.58 Hz, 2 H), 3.46 (s, 1 H), 3.72 (s, 3 H), 3.98 (m, 4 H), 6.84 (dd, J=8.24, 2.14 Hz, 1 H), 6.90 (d, J=7.32 Hz, 1 H), 7.02 (m, 4 H), 7.19 (m, 8 H), 7.52 (dd, J=8.42, 4.39 Hz, 1 H), 8.47 (dd, J=8.42, 1.47 Hz, 1 H), 8.75 (dd, J=4.58, 1.28 Hz, 1 H), 9.35 (s, 1 H).

EXAMPLE 22

(2S)-2-(3(S)- (2-aminoacetylamino)-3-((S)-sec-butyl)-2-oxo-pyrrolidin-1-yl)-N-[(1S, 2R)-1-benzyl-2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-4-phenyl-butyramide
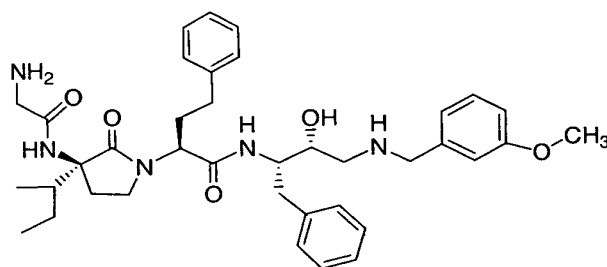
In a manner analogous to the synthesis of the compound of Example (20), but using *N*-Boc glycine, the title compound of Example (22) was prepared as the major, faster-eluting diastereomer (19.9 mg, 31%). ESI MS (M+H)$^+$ = 658.4 ^{1}H NMR (300 MHz, CD$_3$OD) δ ppm 0.59 (d, *J*=6.59 Hz, 3 H), 0.99 (m, 5 H), 1.60 (m, 3 H), 1.77 (m, 1 H), 1.93 (m, 1 H), 2.15 (m, 2 H), 2.44 (m, 2 H), 2.66 (m, 2 H), 2.84 (m, 2 H), 3.03 (m, 1 H), 3.20 (dd, *J*=14.28, 3.30 Hz, 1 H), 3.45 (m, *J*=9.89, 9.89, 4.03 Hz, 1 H), 3.66 (s, 2 H), 3.76 (s, 3 H), 3.82 (m, 1 H), 4.00 (m, 2 H), 4.17 (s, 2 H), 4.39 (dd, *J*=10.07, 5.31 Hz, 1 H), 6.85 (dd, *J*=8.06, 1.83 Hz, 1 H), 6.98 (d, *J*=7.69 Hz, 2 H), 7.03 (d, *J*=1.83 Hz, 1 H), 7.23 (m, 11 H), 8.08 (d, *J*=8.79 Hz, 1 H).

EXAMPLE 23
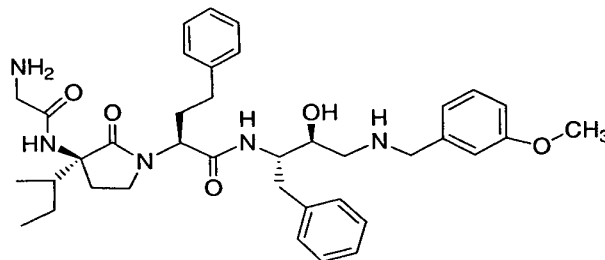
(2S)-2-(3(S)- (2-aminoacetylamino)-3-((S)-*sec*-butyl)-2-oxo-pyrrolidin-1-yl)-*N*-[(1S, 2S)-1-benzyl-2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-4-phenyl-butyramide

Purification of the compound of Example (22) by prep HPLC under the standard reported conditions provided an enriched sample of the compound of Example (23) as the minor, slower-eluting diastereomer (2.1

5   mg, 3%) contaminated with the compound of Example (22) (~2:1 favoring the compound of Example (23). ESI MS (M+H)$^+$ = 658.4 ^{1}H NMR (300 MHz, CD$_3$OD) δ ppm 0.96 (t, J=7.14 Hz, 3 H), 1.08 (d, J=6.59 Hz, 3 H), 1.52 (m, 2 H), 1.75 (dd, J=8.24, 6.77 Hz, 2 H), 2.30 (m, 6 H),

10   2.54 (m, 2 H), 2.97 (m, 3 H), 3.12 (m, 2 H), 3.76 (m, 1 H), 3.80 (s, 3 H), 3.83 (s, 1 H), 3.96 (m, 1 H), 4.08 (m, 1 H), 4.18 (m, 2 H), 4.36 (dd, J=11.35, 2.56 Hz, 1 H), 4.46 (dd, J=11.35, 3.30 Hz, 1 H), 6.99 (m, 1 H), 7.06 (dd, J=6.23, 2.20 Hz, 2 H), 7.12 (dd, J=7.14,

15   1.65 Hz, 2 H), 7.26 (m, 11 H), 7.77 (d, J=8.79 Hz, 1 H).

EXAMPLE 24

(2S)-2-(3(S)-(3-hydroxypropionylamino)-3-((S)-*sec-*

20   butyl)-2-oxo-pyrrolidin-1-yl)-*N*-[(1S, 2R)-1-benzyl-2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-4-phenyl-butyramide

25

In a manner analogous to the synthesis of the compound of Example (20), but using 3-hydroxypropionic acid, the title compound of Example (24) was prepared

(10.6 mg, 16%). ESI MS (M+H)$^+$ = 673.4. ^{1}H NMR (300 MHz, CD$_3$OD) δ ppm 0.80 (d, *J*=6.59 Hz, 3 H), 0.99 (t, *J*=6.96 Hz, 3 H), 1.66 (d, *J*=6.59 Hz, 3 H), 2.13 (m, 2 H), 2.48 (m, 2 H), 2.63 (m, 2 H), 2.88 (m, 2 H), 3.11 (m, 2 H), 3.21 (m, 2 H), 3.67 (m, 2 H), 3.75 (d, *J*=2.56 Hz, 1 H), 3.77 (s, 2 H), 3.78 (s, 3 H), 3.88 (m, 3 H), 3.97 (s, 2 H), 4.15 (dd, *J*=8.79, 5.86 Hz, 2 H), 6.93 (dd, *J*=8.24, 2.38 Hz, 1 H), 7.01 (m, 2 H), 7.21 (m, 11 H), 8.10 (m, 2 H).
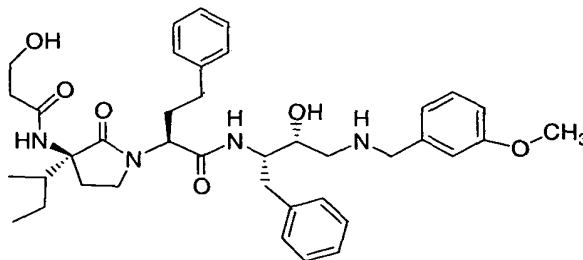
EXAMPLE 25

(2S)-2-(3(S)-(4-hydroxybutyrylamino)-3-((S)-*sec*-butyl)-2-oxo-pyrrolidin-1-yl)-*N*-[(1S, 2R)-1-benzyl-2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-4-phenyl-butyramide
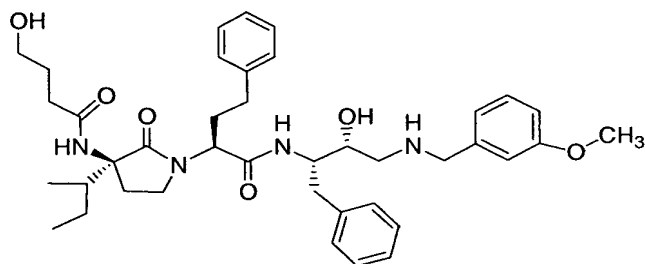
In a manner analogous to the synthesis of the compound of Example (20), but using 4-hydroxybutyric acid, the title compound of Example (25) was prepared (11.8 mg, 18%). ESI MS (M+H)$^+$ = 687.4. ^{1}H NMR (300 MHz, CD$_3$OD) δ ppm 0.75 (m, 1 H), 0.76 (d, *J*=6.59 Hz, 3 H), 0.97 (t, *J*=6.77 Hz, 3 H), 1.07 (m, 1 H), 1.67 (m, 3 H), 1.77 (m, 3 H), 1.98 (m, 1 H), 2.07 (m, 3 H), 2.21 (dd, *J*=9.70, 4.94 Hz, 1 H), 2.31 (m, 3 H), 2.46 (m, 2 H), 2.63 (m, 2 H), 2.84 (m, 3 H), 3.04 (d, *J*=1.83 Hz, 1 H), 3.10 (dd, *J*=7.69, 2.93 Hz, 1 H), 3.17 (m, 1 H),

3.24 (dd, *J*=9.34, 5.68 Hz, 2 H), 3.32 (s, 1 H), 3.54
(td, *J*=6.41, 2.20 Hz, 2 H), 3.77 (s, 3 H), 3.84 (dd,
J=10.07, 4.58 Hz, 1 H), 3.90 (m, 2 H), 3.96 (s, 1 H),
4.10 (m, 2 H), 4.16 (s, 2 H), 4.36 (t, *J*=6.41 Hz, 1

5 H), 6.91 (dd, *J*=8.42, 2.56 Hz, 1 H), 6.99 (m, 2 H),
8.02 (d, *J*=8.79 Hz, 1 H), 8.09 (m, 1 H).

EXAMPLE 26

(2S)-2-(3(S)-(ethylsulfonamido)-3-((S)-*sec*-butyl)-2-
10 oxo-pyrrolidin-1-yl)-*N*-[(1S)-1-benzyl-2-hydroxy-3-(3-
methoxy-benzylamino)-propyl]-4-phenyl-butyramide
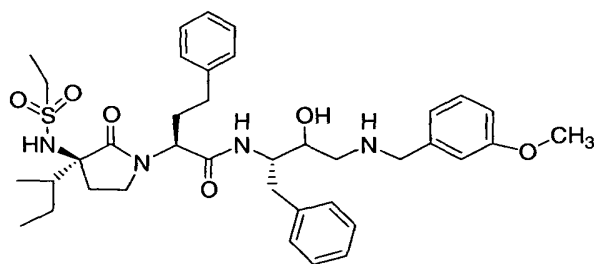
15   The amine from step (13b) (60 mg, 0.085 mmol) was
dissolved in 4 mL of DCM and ethyl sulfonyl chloride
(15 µL, 0.158 mmol), DIEA (100 µL, 0.63 mmol), and a
catalytic amount of DMAP was added. The reaction
solution was heated to reflux temperature for 3 h.

20 The solution was cooled, and the solvents were
removed, and the residue was directly deprotected with
4 mL of a 1:1 solution of TFA in DCM for 1 h at rt,
followed by removal of the solvents. Purification of
the residue by prep LC/MS under the standard described

25 conditions provided the compound of Example (26) as a
1.3:1 mixture of diastereomers at the alcohol center
(16.7 mg, 25%). ESI MS (M+H)$^+$ = 693.4. ^{1}H NMR (300
MHz, CD$_3$OD) δ ppm 0.67 (d, *J*=6.96 Hz, 1 H), 0.97 (m, 6

H), 1.25 (t, *J*=7.32 Hz, 3 H), 1.93 (m, 3 H), 2.19 (m, 1 H), 2.36 (m, 1 H), 2.54 (m, 2 H), 2.67 (m, 1 H), 2.76 (q, *J*=7.44 Hz, 2 H), 2.94 (m, 2 H), 3.43 (m, 3 H), 3.61 (m, 1 H), 3.72 (d, *J*=6.96 Hz, 2 H), 3.75 (s, 2 H), 3.94 (s, 3 H), 4.10 (d, *J*=3.66 Hz, 1 H), 4.17 (d, *J*=3.66 Hz, 1 H), 4.56 (m, 1 H), 4.68 (dd, *J*=9.52, 4.76 Hz, 1 H), 5.46 (d, *J*=10.25 Hz, 1 H), 6.80 (dd, *J*=8.42, 2.56 Hz, 1 H), 6.87 (d, *J*=5.86 Hz, 1 H), 6.97 (m, 4 H), 7.22 (m, 8 H).

EXAMPLE 27

(2S)-2-(3(S)-(propylsulfonamido)-3-((S)-*sec*-butyl)-2-oxo-pyrrolidin-1-yl)-*N*-[(1S)-1-benzyl-2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-4-phenyl-butyramide
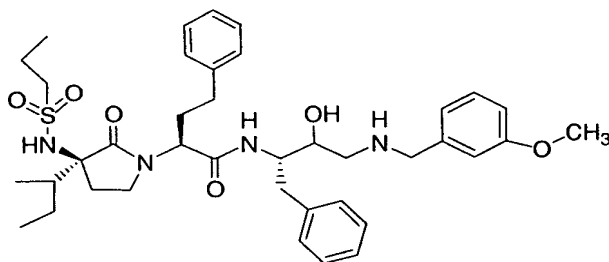
In a manner similar to the synthesis of the compound of Example (26), but using propylsulfonyl chloride the title compound of Example (27) was prepared (24.3 mg, 37%) as a 1:1 mixture of diastereomers at the alcohol center. ESI MS (M+H)$^+$ = 707.4. ^{1}H NMR (300 MHz, CD$_3$OD) δ ppm 0.58 (m, 1 H), 0.67 (d, *J*=6.59 Hz, 1 H), 0.97 (m, 8 H), 1.76 (m, 2 H), 1.94 (m, 3 H), 2.35 (m, 1 H), 2.54 (m, 2 H), 2.72 (m, 2 H), 2.82 (m, 1 H), 2.96 (m, 1 H), 3.11 (dd, *J*=13.36, 6.04 Hz, 1 H), 3.40 (m, 3 H), 3.74 (m, 4 H), 3.93 (m, 3 H), 4.16 (m, 2 H), 4.38 (m, 1 H), 4.55 (m,

1 H), 4.68 (dd, J=9.52, 4.76 Hz, 1 H), 5.10 (m, 1 H), 5.46 (d, J=9.52 Hz, 1 H), 6.80 (dd, J=8.06, 2.20 Hz, 1 H), 6.95 (m, 4 H), 7.22 (m, 9 H).

EXAMPLE 28

<u>(2S)-2-(3(S)-acetylamino-3-(isobutyl)-2-oxo-pyrrolidin-1-yl)-N-[(1S, 2R)-1-benzyl-2-hydroxy-3-(3-methoxy-benzylamino)-propyl]-4-phenyl-butyramide</u>
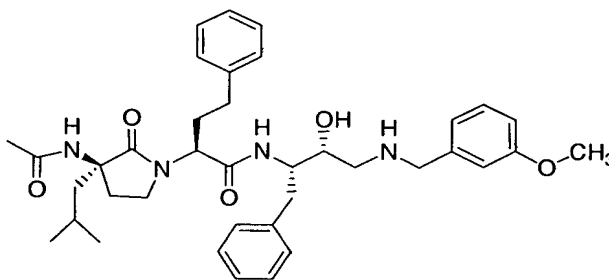
In a manner similar to the synthesis of the compound of Example (3), but using the γ-lactam prepared from L-Leucine the title compound of Example (28) was prepared. ESI MS (M+H)$^+$ = 243.4 ^{1}H NMR (300 MHz, CD$_3$OD) δ ppm 1.02 (d, J=6.59 Hz, 6 H), 1.50 (dd, J=14.46, 7.14 Hz, 1 H), 1.68 (dd, J=14.28, 5.49 Hz, 1 H), 1.83 (dd, J=12.27, 6.41 Hz, 1 H), 1.92 (s, 3 H), 2.05 (m, 2 H), 2.24 (dd, J=10.25, 6.59 Hz, 2 H), 2.34 (s, 5 H), 2.64 (dd, J=13.91, 10.62 Hz, 1 H), 2.89 (s, 1 H), 3.10 (s, 1 H), 3.19 (m, 2 H), 3.28 (dd, J=14.46, 4.21 Hz, 1 H), 3.41 (dd, J=8.24, 6.77 Hz, 1 H), 3.78 (s, 3 H), 4.10 (d, J=4.39 Hz, 3 H), 5.98 (s, 1 H), 6.91 (m, 2 H), 7.03 (d, J=8.42 Hz, 2 H), 7.10 (m, J=8.06 Hz, 1 H), 7.23 (m, 9 H), 7.75 (d, J=9.15 Hz, 1 H).

EXAMPLE 29

(2S)-2-(3(S)-acetylamino-3-((S)-*sec*-butyl)-2-oxo-
pyrrolidin-1-yl)-*N*-[(1S, 2R)-1-benzyl-2-hydroxy-3-(3-
chloro-benzylamino)-propyl]-4-phenyl-butyramide

Following the general procedure for the synthesis
of the compound of Example (3) but using the
intermediate of Preparation (J) the title compound was
prepared. Purification by prep HPLC under the
standard described conditions provided 6.5 mg of the
title compound of Example (30) as the TFA salt.   ESI
MS (M+H)$^+$ = 647.7

EXAMPLE 30

(2S)-2-(3(S)-acetylamino-3-((S)-*sec*-butyl)-2-oxo-
pyrrolidin-1-yl)-*N*-[(1S, 2R)-1-benzyl-2-hydroxy-3-
(propargylamino)-propyl]-4-phenyl-butyramide

Following the general procedure for the synthesis
of the compound of Example (3) but using the
intermediate of Preparation (K) the title compound was
prepared. Purification by prep HPLC under the
5   standard described conditions provided 2.0 mg of the
title compound of Example (30) as the TFA salt.   ESI
MS (M+H)$^+$ = 561

EXAMPLE 31

10   (2S)-2-(3(S)-acetylamino-3-((S)-sec-butyl)-2-oxo-
pyrrolidin-1-yl)-N-[(1S, 2R)-1-benzyl-2-hydroxy-3-
(cyclopropylmethyl)amino-propyl]-4-phenyl-butyramide

15

Following the general procedure for Example (3),
but using the intermediate of Preparation (L) the
title compound was prepared and isolated as a white
solid TFA salt (11.6 mg) in a 35 % yield. LC-MS
20   (column = XTERRA C18 S7, 3 x 50 mm, start %B = 0,
final %B = 100, gradient time = 2 min, flow rate = 5
ml/min) m/e 578.51 (M + H)$^+$, t_R 1.46 min.

EXAMPLE 32

25   (2S)-2-(3(S)-acetylamino-3-(isobutyl)-2-oxo-
pyrrolidin-1-yl)-N-[(1S, 2R)-1-benzyl-2-hydroxy-3-
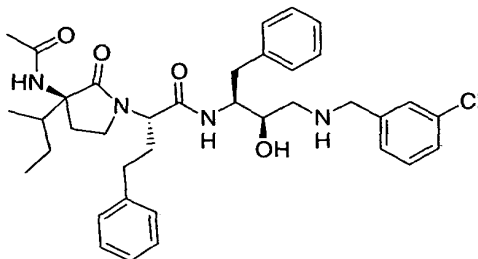(cyclopropylmethyl)amino-propyl]-4-phenyl-butyramide
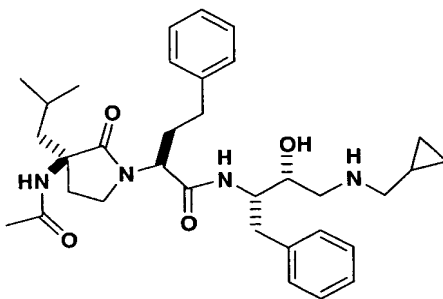
Following the general procedure for Example (3), but using the lactam from Example (28) and the amine of intermediate (L) the title compound was prepared and isolated as a white solid TFA salt (6.3 mg) in a 29 % yield. LC-MS (column = XTERRA C18 S7, 3 x 50 mm, start %B = 0, final %B = 100, gradient time = 2 min, flow rate = 5 ml/min) m/e 577.49 (M + H)$^+$, t_R 1.52 min. ^{1}H NMR (400 MHz, CHLOROFORM-D) δ ppm 0.32 (m, 3 H) 0.64 (d, J=7.58 Hz, 3 H) 1.02 (m, 2 H) 1.52 (m, 1 H) 1.68 (m, 1 H) 1.84 (m, 1 H) 2.23 (m, 13 H) 3.33 (m, 8 H) 3.88 (m, 1 H) 4.13 (m, 1 H) 6.06 (m, 1 H) 7.15 (m, 10 H) 7.80 (m, 1 H) 8.85 (s, 1 H) 9.23 (s, 1 H).

EXAMPLE 33

(2S)-2-(3(S)-acetylamino-3-((S)-*sec*-butyl)-2-oxo-pyrrolidin-1-yl)-*N*-[(1S, 2R)-1-benzyl-2-hydroxy-3-(3,5-difluorobenzylamino)-propyl]-4-phenyl-butyramide
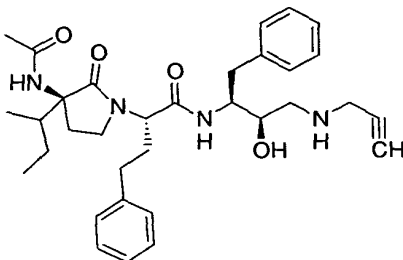
Following the general procedure for Example (3),
but using the intermediate of Preparation (M) the
title compound as a TFA salt was obtained as a pale-
yellow solid (10.3mg) in a 6 % yield. LC-MS (column =

5   XTERRA C18 S7, 3 x 50 mm, start %B = 0, final %B =
100, gradient time = 2 min, flow rate = 5 ml/min) m/e
649.52 (M + H)$^+$, t_R 1.59 min.. ^{1}H NMR (400 MHz,
CHLOROFORM-D) δ ppm 0.93 (m, 6 H) 1.14 (m, 1 H) 1.57
(m, 1 H) 1.73 (m, 1 H) 1.94 (s, 3 H) 2.09 (m, J=6.60

10 Hz, 2 H) 2.30 (m, 2 H) 3.15 (m, 13 H) 4.13 (m, 2 H)
6.10 (m, 1 H) 6.82 (m, 1 H) 7.16 (m, 12 H) 7.93 (s, 1
H).

EXAMPLE 34

15 (2S)-2-(3(S)-acetylamino-3-((S)-sec-butyl)-2-oxo-
pyrrolidin-1-yl)-N-[(1S, 2R)-1-benzyl-2-hydroxy-3-((3-
trifluoromethylbenzyl)amino)-propyl]-4-phenyl-
butyramide
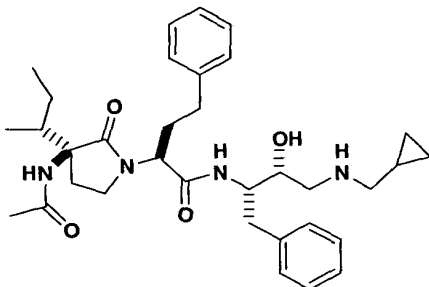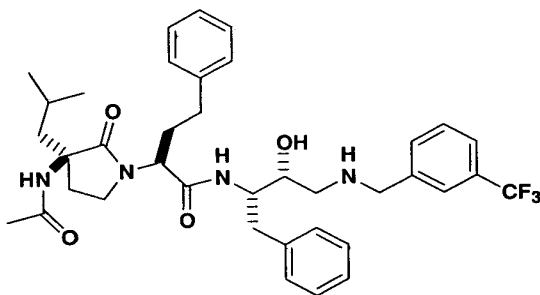

20

Following the general procedure for Example (3),
but using the intermediate of Preparation (N) the
title compound as a TFA salt was obtained as a white

25 solid (15.3 mg) in 49 % yield. LC-MS (column = XTERRA
C18 S7, 3 x 50 mm, start %B = 0, final %B = 100,
gradient time = 2 min, flow rate = 5 ml/min) m/e
681.54 (M + H)$^+$, t_R 1.69.min. ^{1}H NMR (400 MHz,

CHLOROFORM-D) δ ppm 0.96 (m, 7 H) 1.48 (dd, *J*=14.43, 6.85 Hz, 1 H) 1.66 (m, 1 H) 1.81 (m, 4 H) 2.05 (m, 2 H) 2.26 (m, 4 H) 2.63 (dd, *J*=13.82, 10.39 Hz, 1 H) 3.11 (m, 6 H) 3.43 (m, 1 H) 3.84 (m, 1 H) 4.16 (m, 4 H) 6.00 (m, 1 H) 7.06 (m, 3 H) 7.21 (m, 6 H) 7.47 (t, *J*=7.70 Hz, 1 H) 7.67 (m, 4 H).

EXAMPLE 35

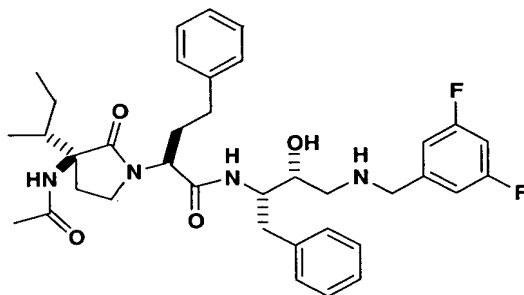2-(3(S)-Acetylamino-3(S)-isobutyl-2-oxo-pyrrolidin-1-yl)-N-[(1S, 2R)-1-benzyl-2-hydroxy-3-benzylamino-propyl]-4-phenyl-butyramide.

Following the general procedure for Example (3), but using the intermediate of Preparation (O), the title compound as a TFA salt was obtained as a beige solid (29.2 mg) in 43 % yield. LC-MS (column = XTERRA C18 S7, 3 x 50 mm, start %B = 0, final %B = 100, gradient time = 2 min, flow rate = 5 ml/min) m/e 613.40 (M + H)$^+$, t_R 1.55.min. ^{1}H NMR (400 MHz, CHLOROFORM-D) δ ppm 0.92 (m, 7 H) 1.49 (dd, *J*=14.31, 6.72 Hz, 1 H) 1.65 (m, 1 H) 1.81 (m, 1 H) 1.90 (s, 3 H) 2.03 (m, 2 H) 2.31 (m, 4 H) 2.63 (dd, *J*=13.57, 10.64 Hz, 1 H) 3.07 (m, 6 H) 3.46 (m, 1 H) 3.86 (m, 1 H) 4.10 (s, 4 H) 4.85 (s, 4 H) 6.10 (s, 1 H) 7.18 (m, 10 H) 7.69 (d, *J*=8.80 Hz, 1 H).

EXAMPLE 36

(2S)-2-(3(S)-acetylamino-3-((S)-*sec*-butyl)-2-oxo-
pyrrolidin-1-yl)-*N*-[(1S, 2R)-1-benzyl-2-hydroxy-3-(3-
fluoro,5-(trifluoromethyl)benzylamino)-propyl]-4-
5   phenyl-butyramide
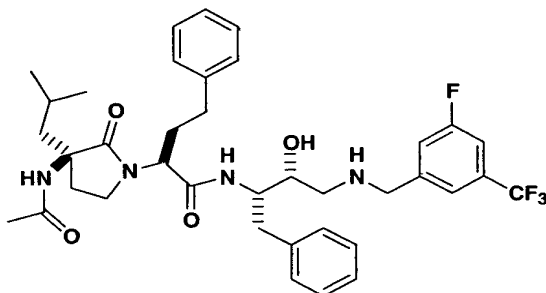
     Following the general procedure for Example (3),
10   but using the intermediate of Preparation (P) the
title compound as a TFA salt was obtained as a white
solid (6.0 mg) in 23 % yield. LC-MS (column = XTERRA
C18 S7, 3 x 50 mm, start %B = 0, final %B = 100,
gradient time = 2 min, flow rate = 5 ml/min) m/e
15   699.48 (M + H)$^+$, t_R 1.69 min.

EXAMPLE 37

2-(3(S)-Acetylamino-3(S)-isobutyl-2-oxo-pyrrolidin-1-
yl)-N-[(1S, 2R)-1-benzyl-3-(2-cyano-ethylamino)-2-
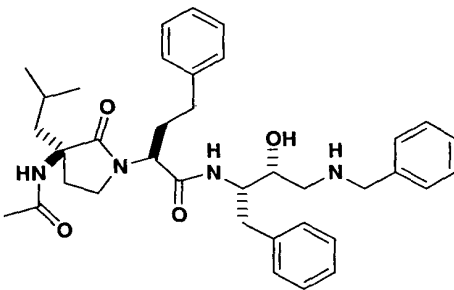20   hydroxy-propyl]-4-phenyl-butyramide
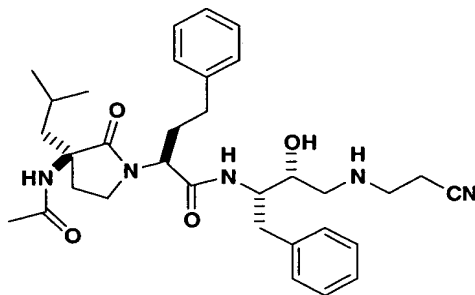
Following the general procedure for Example (3),
but using the intermediate from Preparation (Q) the
title compound as a TFA salt was obtained as a
colorless residue (15.0 mg) in 65 % yield. LC-MS

5   (column = XTERRA C18 S7, 3 x 50 mm, start %B = 0,
final %B = 100, gradient time = 2 min, flow rate = 5
ml/min) m/e 576.59 (M + H)$^+$, t_R 1.55.min. ^{1}H NMR (400
MHz, CHLOROFORM-D) δ ppm 1.02 (dd, J=6.60, 1.71 Hz, 6
H) 1.50 (dd, J=14.43, 6.85 Hz, 1 H) 1.69 (m, 1 H) 1.83

10   (m, J=12.84, 6.48 Hz, 1 H) 2.02 (s, 3 H) 2.14 (m, 1 H)
2.36 (m, 4 H) 3.14 (m, 10 H) 3.95 (m, 1 H) 4.13 (m, 1
H) 5.67 (m, 4 H) 6.13 (s, 1 H) 7.17 (m, 10 H) 7.98 (d,
J=8.80 Hz, 1 H).

15   **EXAMPLE 38**

(2S)-2-(3(S)-acetylamino-3-(cyclopropylmethyl)-2-oxo-
pyrrolidin-1-yl)-N-[(1S, 2R)-1-(3,5-difluorobenzyl)-2-
hydroxy-3-(3-methoxybenzylamino)-propyl]-4-(2-
methoxyphenyl)-butyramide

20
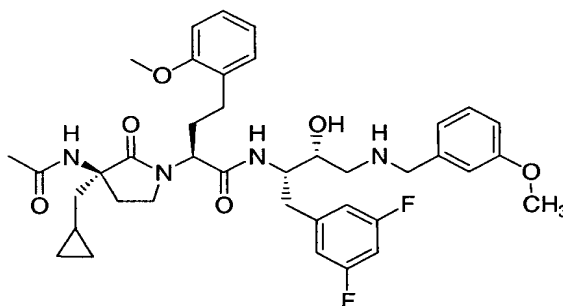
Step (38a): 2-tert-Butoxycarbonylamino-succinic
acid 1-benzyl ester 4-(2,5-dioxo-pyrrolidin-1-yl)

25   ester. A solution of BOC-Asp-OBzl (1.49 g, 4.61 mmol)
and N-Hydroxy succinimide (584 mg, 5.07 mmol) in EtOAc
(20 ml) was cooled to 0 °C. 1,3-

Dicyclohexylcarbodiimide (1.05 g, 5.07 mmol) was added portionwise. White precipitate began forming. The resulting mixture was stirred at 0 °C for 10 min. and then allowed to warm to rt. After stirring for 6 h.

5   at rt the mixture was filtered and concentrated *in vacuo* to give 2-tert-butoxycarbonylamino-succinic acid 1-benzyl ester 4-(2,5-dioxo-pyrrolidin-1-yl) ester as a clear, colorless oil which was used without further purification.

10

    Step (38b): 2-tert-Butoxycarbonylamino-4-hydroxy-butyric acid benzyl ester. A solution of NaBH$_4$ (262 mg, 6.92 mmol) in 4:1 THF:H$_2$O (20 ml) was cooled to 0 °C. To this mixture was slowly added a solution of the

15   ester from step (38a) (4.61 mmol) in THF (12 ml). Gas was evolved. When gas evolution ceased, the mixture was quenched by the careful addition of saturated aqueous ammonium chloride. The resulting solution was stirred at 0 °C for an additional 15 min. EtOAc (200

20   ml) was added and the resulting mixture was washed with saturated aqueous NaCl. The organic layer was dried over MgSO$_4$ and concentrated *in vacuo*. Purification by flash chromatography (silica, 0-10% MeOH/CHCl$_3$) gave 2-tert-butoxycarbonylamino-4-hydroxy-

25   butyric acid benzyl ester (350 mg, 25% for 2 steps) as a clear, colorless oil: ^{1}H NMR (300 MHz, Methanol-D) δ 7.23-7.43 (m, 5 H), 5.06-5.25 (m, 2 H), 4.00-4.57 (m, 2 H), 3.54-3.69 (m, 1 H), 1.67-2.12 (m, 2 H), 1.29-1.50 (m, 9 H). LC-MS (Method A, retention time: 1.25

30   min).

Step (38c): 2-tert-Butoxycarbonylamino-4-iodo-
butyric acid benzyl ester. To a solution of
triphenylphosphine (734 mg, 2.80 mmol) and imidazole
(191 mg, 2.80 mmol) in CH_2Cl_2 (10 ml) at rt was added
iodine (711 mg, 2.80 mmol) portionwise over 5 min.
The mixture first turned yellow, then brown and
developed precipitate. The mixture was stirred at rt
until no pieces of iodine were visible (approx. 5
min.). A solution of alcohol from step (38b) (721 mg,
2.33 mmol) in CH_2Cl_2 (5 ml) was added and the resulting
mixture was stirred at rt for 1h. The reaction was
filtered and concentrated *in vacuo*. Purification by
flash chromatography (silica, 0-25% EtOAc/Hexane) gave
2-tert-butoxycarbonylamino-4-iodo-butyric acid benzyl
ester (430 mg, 44%) as a slightly yellow oil: ^{1}H NMR
(500 MHz, Methanol-D) δ 7.24-7.42 (m, 5 H), 5.07-5.25
(m, 2 H), 4.16-4.31 (m, 1 H), 3.12-3.29 (m, 2 H),
2.01-2.35 (m, 2 H), 1.42 (s, 9 H). LC-MS (Method A,
retention time: 1.71 min), MS *m/z* 420 (M[+]+1).

Step (38d): General Procedure. 2-tert-
Butoxycarbonylamino-4-phenyl-butyric acid benzyl
esters. A suspension of 325 mesh Zinc dust (1.387 g,
28.62 mmol) in DMF (2.5 ml) was treated with 1,2-
dibromoethane (123 μl, 1.43 mmol). The mixture was
heated to 60 °C and stirred for 30 min. After cooling
to rt, chlorotrimethylsilane (37 μl, 0.29 mmol) was
added and the resulting mixture was stirred at rt for
30 min. A solution of iodide (38c) (2.0 g, 4.77 mmol)
in DMF (2.5 ml) was then added and the mixture was
heated to 35 °C. After stirring for 1 h. at 35 °C, at
which time TLC indicated there was no starting iodide

remaining, the mixture was allowed to cool to rt
Pd$_2$(dba)$_3$ (87 mg, 0.095 mmol), tri-o-tolyl phosphine
(116 mg, 0.38 mmol), and 2-methoxyiodobenzene (step
(38d), or other aryl iodides) (4.77 mmol) were added

5 and the mixture was stirred at rt for 16 h. The
reaction was diluted with EtOAc and washed with
saturated aqueous NaCl. The organic layer was dried
over MgSO$_4$ and concentrated *in vacuo*. Purification by
flash chromatography (silica, 0-25% EtOAc/Hexane) gave

10 the desired 2-tert-Butoxycarbonylamino-4-(2-
methoxyphenyl)-butyric acid benzyl ester (30-60%) or
other desired similar esters. 2-tert-
Butoxycarbonylamino-4-(2-methoxyphenyl)-butyric acid
benzyl ester :^{1}H NMR (500 MHz, Methanol-D) δ 7.25-7.41

15 (m, 5 H), 7.15 (t, *J*=7.02 Hz, 1 H), 7.04 (d, *J*=7.32
Hz, 1 H), 6.89 (d, *J*=7.93 Hz, 1 H), 6.82 (t, *J*=7.32
Hz, 1 H), 5.02-5.20 (m, 2 H), 3.91-4.12 (m, 1 H), 3.78
(s, 3 H), 2.56-2.72 (m, 2 H), 1.79-2.14 (m, 2 H),
1.29-1.49 (m, 9 H). LC-MS (Method A, retention time:

20 1.82 min), MS *m/z* 400 (M$^+$+1).

Step (38e): 2-Amino-4-(2-methoxyphenyl)-butyric
acid benzyl ester. A solution of ester from step (38d)
(2.86 mmol) in CH$_2$Cl$_2$ (5 ml) was treated with

25 trifluoroacetic acid (5 ml). The mixture was stirred
at rt for 1 h. The reaction was then concentrated *in
vacuo*. The residue was taken up in EtOAc. The
resulting solution was washed with saturated aqueous
NaHCO$_3$, saturated aqueous NaCl, dried over MgSO$_4$ and

30 concentrated *in vacuo*. Purification by flash
chromatography (silica, 0-20% MeOH/CHCl$_3$) gave the
desired amine (quantitative). ^{1}H NMR (500 MHz,
Methanol-D) δ 7.29-7.43 (m, 5 H), 7.14-7.21 (m, 1 H),

7.04 (dd, *J*=7.32, 1.53 Hz, 1 H), 6.91 (d, *J*=8.24 Hz, 1 H), 6.78-6.86 (m, 1 H), 5.20 (q, *J*=12.21 Hz, 2 H), 3.82 (t, *J*=6.26 Hz, 1 H), 2.56-2.79 (m, 2 H), 1.96-2.16 (m, 2 H). LC-MS (Method A, retention time: 1.32

5   min), MS *m/z* 300 (M$^+$+1).

Step (38f): The acid from step (6e) (250 mg, 825 μmol, 1 eq) and the amine from step (38e) (296 mg, 990 μmol, 1.2 eq) were dissolved in CH_2Cl_2 (6.5 mL) and DMF

10   (3.2 mL). PyBOP (515 mg, 990 μmol, 1.2 eq) was then added, followed by DIEA (575 μL, 3.30 mmol, 4 eq). The reaction was allowed to stir at room temperature overnight. Water was added, and the mixture extracted three times into CH_2Cl_2. The combined organic layers

15   were dried over $MgSO_4$, filtered, and concentrated *in vacuo*. Chromatography (5% to 40% EtOAc/Hexane) yielded 336.6 mg of product (70% yield).
1H NMR (500 MHz, CHLOROFORM-D) δ ppm 7.32 (m, 10 H), 7.17 (m, 1 H), 7.03 (dd, *J*=7.32, 1.53 Hz, 1 H), 6.83

20   (m, 2 H), 6.66 (d, *J*=7.63 Hz, 1 H), 5.92 (m, 1 H), 5.68 (m, 1 H), 5.07 (m, 6 H), 4.70 (m, 1 H), 3.76 (m, 3 H), 2.96 (m, 1 H), 2.65 (m, 1 H), 2.57 (m, 1 H), 2.48 (dd, *J*=14.50, 6.56 Hz, 1 H), 2.15 (m, 1 H), 2.04 (m, 2 H), 1.81 (dd, *J*=14.50, 5.95 Hz, 1 H), 0.59 (m, 1

25   H), 0.37 (m, 2 H), 0.03 (m, 2 H). LC-MS (Method B, retention time: 2.820 min), MS *m/z* 585 (M$^+$+1).

Step (38g): The amide from step (38f) (333 mg, 570 μmol, 1 eq) was dissolved in ether (1.8 mL). A

30   solution of sodium periodate (265 mg) in water (1.8 mL) was added, followed by a 2.5 wt % solution of osmium tetroxide in *t*-butanol (390 μL). The combined

reagents were stirred rapidly at room temperature overnight. Ether was added, mixed with the reaction mixture and decanted (2 times). The combined organic layers were dried over MgSO$_4$, filtered, and

5   concentrated *in vacuo*. The resulting residue was dissolved in CH$_2$Cl$_2$ and cooled to 0 °C. Simultaneously, triethyl silane (2.6 mL) and TFA (2.6 mL) was added over approx. 90 seconds. The reaction was kept at 0 °C for 3 h. Solvents were removed *in*

10   *vacuo*. Chromatography (5% to 40% EtOAc/Hexane) yielded 163.8 mg product (50% yield). ^{1}H NMR (500 MHz, CHLOROFORM-D) δ ppm 7.31 (m, 10 H), 7.15 (m, 2 H), 6.83 (m, 2 H), 5.08 (m, 4 H), 4.87 (dd, *J*=11.14, 4.43 Hz, 1 H), 4.11 (m, 1 H), 3.76 (m, 3 H), 3.40 (m, 2 H),

15   2.71 (m, 1 H), 2.56 (m, 1 H), 2.44 (m, 2 H), 2.24 (m, 1 H), 1.92 (m, 1 H), 1.65 (m, 1 H), 1.54 (m, 1 H), 0.70 (m, 1 H), 0.38 (m, 2 H), -0.01 (m, 2 H). LC-MS (Method A, retention time: 1.973 min), MS *m/z* 571 (M$^+$+1).

20

    Step (38h): The lactam from step (38g) (160 mg, 281 μmol) was dissolved in ethanol (25 mL). Ten percent Pd on carbon was added, and the reaction stirred under balloon pressure hydrogen for 4 h. The

25   reaction was filtered over celite, and concentrated *in vacuo*. The residue was dissolved in pyridine, and acetic anhydride was added. After stirring at room temperature overnight, the solvents were removed *in vacuo*. Chromatography yielded pure lactam acid. ^{1}H

30   NMR (500 MHz, CD$_3$OD) δ ppm 7.17 (m, 2 H), 6.91 (d, *J*=7.93 Hz, 1 H), 6.83 (t, *J*=6.87 Hz, 1 H), 4.70 (dd, *J*=11.29, 4.27 Hz, 1 H), 3.83 (s, 3 H), 3.60 (m, 2 H),

2.88 (m, 1 H), 2.54 (m, 2 H), 2.39 (m, 1 H), 2.18 (m, 1 H), 2.05 (m, 1 H), 1.94 (s, 3 H), 1.75 (m, 1 H), 1.59 (dd, *J*=13.89, 7.17 Hz, 1 H), 0.85 (m, 1 H), 0.49 (m, 2 H), 0.18 (m, 1 H), 0.09 (m, 1 H). LC-MS (Method A, retention time: 1.823 min), MS *m/z* 389 (M$^+$+1).

Step (38i): The acid from step (38h) (22.1 mg, 57.0 µmol) and the amine of Preparation (B) (31.0 mg, 71.2 µmol, 1.25 eq) were dissolved in DMF (670 µL). PyBOP (36 mg) and DIEA (40 µL) were added, and the reaction stirred at room temperature overnight. The reaction was purified by reverse-phase chromatography, and concentrated *in vacuo*. The residue was stirred in 1:1 TFA/CH$_2$Cl$_2$ for 2 h, then concentrated *in vacuo*. The residue was trapped on SCX resin, then eluted with 2 M NH$_3$/MeOH to provide the title compound of Example (38) after concentration *in vacuo*. ^{1}H NMR (500 MHz, CHLOROFORM-D) δ ppm 7.40 (d, *J*=9.16 Hz, 1 H), 7.16 (m, 2 H), 7.05 (m, 1 H), 6.86 (m, 3 H), 6.78 (m, 4 H), 6.52 (m, 1 H), 6.30 (s, 1 H), 4.10 (m, 1 H), 3.78 (m, 9 H), 3.62 (m, 1 H), 3.36 (m, 1 H), 3.21 (q, *J*=8.44 Hz, 1 H), 2.94 (m, 1 H), 2.71 (m, 3 H), 2.52 (m, 3 H), 2.36 (m, 1 H), 2.25 (m, 1 H), 2.01 (m, 5 H), 1.74 (dd, *J*=14.04, 5.49 Hz, 1 H), 1.48 (dd, *J*=14.19, 7.78 Hz, 1 H), 1.27 (m, 1 H), 0.78 (dd, *J*=7.63, 5.19 Hz, 1 H), 0.58 (m, 2 H), 0.17 (m, 2 H). LC-MS (Method B, retention time: 2.263 min), MS *m/z* 707 (M$^+$+1).

EXAMPLE 39

(2S)-2-(3(S)-acetylamino-3-cyclopropylmethyl)-2-oxo-
pyrrolidin-1-yl)-N-[(1S, 2R)-1-(3,5-difluorobenzyl)-2-
hydroxy-3-(3-methoxybenzylamino)-propyl]-4-(3,4-
5   methylenedioxyphenyl)-butyramide
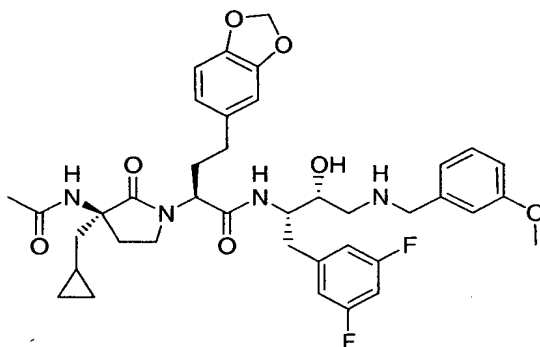
Step (39a):   The iodide of step (38c) was
10 reacted in the manner of step (38d) with 1-iodo-3,4-
methylenedioxybenzene to provide the compound of step
(39a). ^{1}H NMR (500 MHz, Methanol-D) δ 7.25-7.40 (m, 5
H), 6.53-6.74 (m, 3 H), 5.87 (s, 2 H), 5.04-5.24 (m, 2
H), 4.09 (q, J=7.02 Hz, 1 H), 2.46-2.65 (m, 2 H),
15 1.79-2.09 (m, 2 H), 1.26-1.50 (m, 9 H). LC-MS (Method
A, retention time: 1.83 min), MS m/z 414 (M$^+$+1).

Step (39b):   Boc-amino acid (39a) was reacted in
the manner of step (38e) to provide the amine of step
20 (39b). ^{1}H NMR (500 MHz, Methanol-D) δ 7.33-7.47 (m, 5
H), 6.51-6.73 (m, 3 H), 5.84-5.91 (m, 2 H), 5.15-5.40
(m, 2 H), 4.05 (t, J=6.26 Hz, 1 H), 2.44-2.70 (m, 2
H), 1.99-2.23 (m, 2 H). LC-MS (Method A, retention
time: 1.26 min), MS m/z 314 (M$^+$+1).

25

Step (39c): The acid from step (6e) (200 mg, 660 μmol, 1 eq) and the amine of step (39b) (248 mg, 792 μmol, 1.2 eq) were reacted in the manner of step (38f). ^{1}H NMR (500 MHz, CHLOROFORM-D) δ ppm 7.32 (m, 10 H), 6.66 (m, 2 H), 6.56 (d, J=1.53 Hz, 1 H), 6.51 (d, J=7.93 Hz, 1 H), 5.89 (s, 2 H), 5.79 (m, 1 H), 5.67 (m, 1 H), 5.21 (m, 1 H), 5.09 (m, 5 H), 4.66 (m, 1 H), 2.91 (dd, J=14.34, 7.63 Hz, 1 H), 2.49 (m, 3 H), 2.10 (m, 1 H), 1.96 (m, 2 H), 1.82 (dd, J=14.34, 6.10 Hz, 1 H), 0.57 (m, 1 H), 0.38 (m, 2 H), 0.02 (m, 2 H). LC-MS (Method B, retention time: 2.777 min), MS m/z 599 (M^++1).

Step (39d): The product from step (39c) was reacted in the manner of step (38g). ^{1}H NMR (500 MHz, CHLOROFORM-D) δ ppm 7.31 (m, 10 H), 6.67 (m, 3 H), 5.89 (s, 2 H), 5.48 (s, 1 H), 5.09 (m, 4 H), 4.85 (dd, J=10.83, 4.43 Hz, 1 H), 3.35 (m, 2 H), 2.58 (m, 2 H), 2.40 (m, 2 H), 2.24 (m, 1 H), 2.02 (m, 1 H), 1.63 (m, 1 H), 1.52 (m, 1 H), 0.69 (m, 1 H), 0.38 (m, 2 H), - 0.01 (m, 2 H). LC-MS (Method A, retention time: 1.933 min), MS m/z 585 (M^++1).

Step (39e): The product from step (39d) was reacted in the manner of step (38h) and taken on without further purification.

Step (39f): The product from step (39e) was reacted in the manner of step (38i) to produce the title compound of Example (39). ^{1}H NMR (500 MHz, CHLOROFORM-D) δ ppm 7.46 (m, 1 H), 7.23 (m, 2 H), 6.66 (m, 10 H), 6.28 (s, 1 H), 5.90 (m, 2 H), 4.11 (m, 1

H), 3.78 (m, 5 H), 3.55 (m, 1 H), 3.31 (m, 1 H), 3.17
(m, 1 H), 2.98 (dd, J=14.19, 3.51 Hz, 1 H), 2.71 (m, 3
H), 2.35 (m, 5 H), 2.01 (m, 5 H), 1.70 (dd, J=14.19,
5.34 Hz, 1 H), 1.47 (dd, J=14.04, 7.93 Hz, 1 H), 0.76

5   (m, 1 H), 0.60 (m, 2 H), 0.19 (m, 2 H). LC-MS (Method
B, retention time: 2.107 min), MS m/z 721 (M^++1).

EXAMPLE 40

(2S)-2-(3(S)-acetylamino-3-(cyclopropylmethyl)-2-oxo-
10   pyrrolidin-1-yl)-N-[(1S, 2R)-1-(3,5-difluorobenzyl)-2-
hydroxy-3-(3-methoxybenzylamino)-propyl]-4-(3-
fluorophenyl)-butyramide
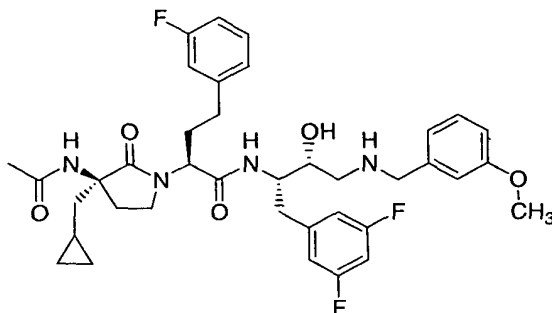
15

Step (40a): The iodide of step (38c) was reacted
in the manner of step (38d) with 1-iodo-3-
fluorobenzene to provide the compound of step (40a).
^{1}H NMR (500 MHz, Methanol-D) δ 7.27-7.41 (m, 5 H),
20   6.83-7.28 (m, 4 H), 5.06-5.23 (m, 2 H), 4.11 (d,
J=4.58 Hz, 1 H), 2.57-2.73 (m, 2 H), 1.84-2.13 (m, 2
H), 1.27-1.49 (m, 9 H).

Step (40b): Boc-amino acid from step (40a) was
25   reacted in the manner of step (38e) to provide the
compound of step (40b). ^{1}H NMR (500 MHz, Methanol-D) δ
7.34-7.48 (m, 5 H), 7.23-7.31 (m, 1 H), 6.89-6.98 (m,

2 H), 6.86 (d, J=10.07 Hz, 1 H), 5.21-5.38 (m, 2 H), 4.08 (t, J=6.10 Hz, 1 H), 2.54-2.79 (m, 2 H), 2.00-2.30 (m, 2 H). LC-MS (Method A, retention time: 1.30 min), MS m/z 288 (M^++1).

Step (40c): The acid from step (6e) (250 mg, 825 μmol, 1 eq) and the amine from step (40b) (284 mg, 990 μmol, 1.2 eq) were reacted in the manner of step (38f). ^{1}H NMR (500 MHz, CHLOROFORM-D) δ ppm 7.32 (m, 10 H), 7.19 (m, 1 H), 6.86 (m, 2 H), 6.74 (m, 2 H), 5.68 (m, 2 H), 5.20 (m, 1 H), 5.10 (m, 5 H), 4.67 (dd, J=12.51, 7.32 Hz, 1 H), 2.90 (dd, J=14.19, 7.78 Hz, 1 H), 2.57 (m, 2 H), 2.49 (m, 1 H), 2.15 (m, 1 H), 1.97 (m, 2 H), 1.85 (dd, J=14.34, 6.10 Hz, 1 H), 0.58 (m, 1 H), 0.39 (m, 2 H), 0.03 (m, 2 H). LC-MS (Method A, retention time: 1.967 min), MS m/z 573 (M^++1).

Step (40d): The product from step (40c) was reacted in the manner of step (38g). ^{1}H NMR (500 MHz, CHLOROFORM-D) δ ppm 7.32 (m, 10 H), 7.20 (m, 1 H), 6.96 (m, 2 H), 6.86 (m, 1 H), 5.46 (s, 1 H), 5.09 (m, 4 H), 4.87 (dd, J=10.99, 4.58 Hz, 1 H), 3.36 (m, 2 H), 2.67 (m, 2 H), 2.38 (m, 2 H), 2.31 (m, 1 H), 2.03 (m, 1 H), 1.63 (m, 1 H), 1.51 (dd, J=14.34, 7.02 Hz, 1 H), 0.69 (m, 1 H), 0.39 (m, 2 H), 0.00 (m, 2 H). LC-MS (Method A, retention time: 1.953 min), MS m/z 559 (M^++1).

Step (40e): The product from step (40d) was reacted in the manner of step (38h). LC-MS (Method A, retention time: 1.283 min), MS m/z 377 (M^++1).

Step (40f): The product from step (40e) was reacted in the manner of step (38i) to provide the title compound of Example (40). ^{1}H NMR (500 MHz, CHLOROFORM-D) δ ppm 7.41 (d, J=9.16 Hz, 1 H), 7.20 (m, 2 H), 6.82 (m, 8 H), 6.53 (m, 1 H), 6.29 (s, 1 H), 4.13 (m, 1 H), 3.79 (m, 6 H), 3.56 (m, 1 H), 3.33 (m, 1 H), 3.16 (m, 1 H), 2.96 (dd, J=14.34, 3.66 Hz, 1 H), 2.69 (m, 3 H), 2.46 (m, 4 H), 2.23 (m, 1 H), 2.12 (m, 1 H), 2.01 (m, 3 H), 1.70 (dd, J=14.04, 5.49 Hz, 1 H), 1.47 (dd, J=14.04, 7.93 Hz, 1 H), 0.76 (m, 1 H), 0.60 (m, 2 H), 0.19 (m, 2 H). LC-MS (Method B, retention time: 2.160 min), MS m/z 695 (M$^+$+1).

EXAMPLE 41

(2S)-2-(3(S)-acetylamino-3-(cyclopropylmethyl)-2-oxo-pyrrolidin-1-yl)-N-[(1S, 2R)-1-(3,5-difluorobenzyl)-2-hydroxy-3-(3-methoxybenzylamino)-propyl]-4-(4-fluorophenyl)-butyramide
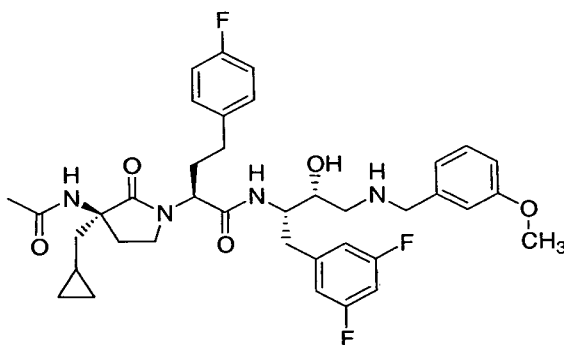
Step (41a): The iodide of step (38c) was reacted in the manner of step (38d) with 1-iodo-4-fluorobenzene to provide the compound of step (41a). ^{1}H NMR (500 MHz, Methanol-D) δ 7.26-7.44 (m, 5 H), 6.68-7.20 (m, 4 H), 5.03-5.24 (m, 2 H), 3.92-4.15 (m,

1 H), 2.50-2.75 (m, 2 H), 1.82-2.11 (m, 2 H), 1.30-1.52 (m, 9 H).

Step (41b): Boc-amino acid from step (41a) was reacted in the manner of step (38e) to provide the compound of step (41b). ^{1}H NMR (500 MHz, Methanol-D) δ 7.33-7.47 (m, 5 H), 7.08-7.16 (m, 2 H), 6.93-7.04 (m, 2 H), 5.20-5.38 (m, 2 H), 4.07 (t, J=6.26 Hz, 1 H), 2.50-2.79 (m, 2 H), 2.05-2.23 (m, 2 H). LC-MS (Method A, retention time: 1.32 min), MS m/z 288 (M$^+$+1).

Step (41c): The acid from step (6e) (250 mg, 825 μmol, 1 eq) and the amine from step (41b) (284 mg, 990 μmol, 1.2 eq) were reacted in the manner of step (38f). ^{1}H NMR (500 MHz, CHLOROFORM-D) δ ppm 7.33 (m, 12 H), 7.01 (m, 2 H), 6.92 (m, 2 H), 6.69 (d, J=7.63 Hz, 1 H), 5.69 (m, 2 H), 5.18 (m, 6 H), 4.66 (m, 1 H), 2.90 (m, 1 H), 2.52 (m, 3 H), 2.12 (m, 1 H), 1.96 (m, 2 H), 1.85 (m, 1 H), 0.57 (m, 1 H), 0.39 (m, 2 H), 0.03 (m, 2 H). LC-MS (Method B, retention time: 2.840 min), MS m/z 573 (M$^+$+1).

Step (41d): The product from step (41c) was reacted in the manner of step (38g). ^{1}H NMR (500 MHz, CHLOROFORM-D) δ ppm 7.30 (m, 10 H), 7.15 (m, 2 H), 6.93 (m, 2 H), 5.46 (s, 1 H), 5.09 (m, 4 H), 4.85 (dd, J=10.99, 4.27 Hz, 1 H), 3.36 (s, 2 H), 2.63 (m, 2 H), 2.39 (m, 2 H), 2.27 (m, 1 H), 2.08 (m, 1 H), 1.62 (dd, J=14.04, 6.41 Hz, 1 H), 1.50 (m, 1 H), 0.70 (m, 1 H), 0.39 (m, 2 H), 0.02 (m, 2 H). LC-MS (Method A, retention time: 1.947 min), MS m/z 559 (M$^+$+1).

Step (41e): The product from step (41d) was reacted in the manner of step (38h). LC-MS (Method A, retention time: 1.280 min), MS *m/z* 377 (M⁺+1).

5   Step (41f): The product from step (41e) was reacted in the manner of step (38i) to provide the title compound of Example (41). ¹H NMR (500 MHz, CHLOROFORM-D) δ ppm 7.41 (d, *J*=9.16 Hz, 1 H), 7.21 (m, 2 H), 7.06 (m, 2 H), 6.92 (m, 4 H), 6.77 (m, 3 H), 10   6.52 (m, 1 H), 4.12 (m, 1 H), 3.79 (m, 5 H), 3.56 (m, 1 H), 3.33 (m, 1 H), 3.16 (m, 1 H), 2.96 (dd, *J*=14.19, 3.81 Hz, 1 H), 2.70 (m, 3 H), 2.44 (m, 4 H), 2.22 (m, 1 H), 2.09 (m, 1 H), 2.00 (m, 3 H), 1.69 (dd, *J*=14.04, 5.49 Hz, 1 H), 1.47 (dd, *J*=14.04, 7.93 Hz, 1 H), 0.76 15   (m, 1 H), 0.59 (m, 2 H), 0.19 (m, 2 H). LC-MS (Method B, retention time: 2.153 min), MS *m/z* 695 (M⁺+1).

EXAMPLE 42

20   (2S)-2-(3(S)-acetylamino-3-(cyclopropylmethyl)-2-oxo-pyrrolidin-1-yl)-*N*-[(1S, 2R)-1-(3,5-difluorobenzyl)-2-hydroxy-3-(3-methoxybenzylamino)-propyl]-4-(3-methoxyphenyl)-butyramide
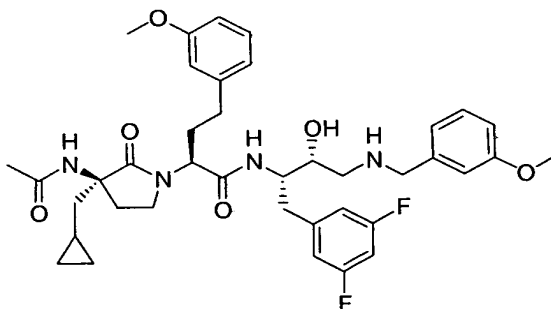
25

Step (42a): The iodide of step (38c) was reacted in the manner of step (38d) with 1-iodo-3-

methoxybenzene to provide the compound of step (42a).
^{1}H NMR (300 MHz, Methanol-D) δ 7.24-7.42 (m, 5 H),
6.98-7.12 (m, 2 H), 6.80 (d, J=8.42 Hz, 2 H), 5.01-
5.25 (m, 2 H), 3.99-4.21 (m, 1 H), 3.74 (s, 3 H),
5   2.45-2.69 (m, 2 H), 1.76-2.13 (m, 2 H), 1.27-1.52 (m,
9 H). LC-MS (Method A, retention time: 1.83 min), MS
m/z 400 (M$^+$+1).

      Step (42b): The boc-amino acid from step (42a)
10 was reacted in the manner of step (38e) to provide the
compound of step (42b). ^{1}H NMR (300 MHz, Methanol-D) δ
7.24-7.43 (m, 5 H), 7.02 (d, J=8.42 Hz, 2 H), 6.72-
6.84 (m, 2 H), 5.06-5.24 (m, 2 H), 3.73 (s, 3 H), 3.46
(t, J=6.40 Hz, 1 H), 2.40-2.70 (m, 2 H), 1.72-2.08 (m,
15 2 H). LC-MS (Method A, retention time: 1.28 min), MS
m/z 300 (M$^+$+1).

      Step (42c): The acid from step (6e) (269.5 mg,
889 µmol, 1 eq) and the amine from step (42b)(319.1
20 mg, 1.067 mmol, 1.2 eq) were reacted in the manner of
step (38f).
^{1}H NMR (500 MHz, CHLOROFORM-D) δ ppm 7.32 (m, 12 H),
7.16 (t, J=7.93 Hz, 1 H), 6.68 (m, 4 H), 5.80 (s, 1
H), 5.66 (m, 1 H), 5.19 (m, 1 H), 5.08 (m, 5 H), 4.69
25 (m, 1 H), 3.75 (s, 3 H), 2.90 (m, 2 H), 2.52 (m, 2 H),
2.15 (m, 1 H), 2.00 (m, 1 H), 1.79 (dd, J=14.50, 5.95
Hz, 1 H), 0.56 (m, 1 H), 0.38 (m, 2 H), 0.02 (m, 2 H).
LC-MS (Method B, retention time: 2.790 min), MS *m/z*
585 (M$^+$+1).

30

Step (42d): The product from step (42c) was reacted in the manner of step (38g) and taken on without further purification.

5   Step (42e): The product from step (42d) was reacted in the manner of step (38h) and taken on without further purification.

Step (42f): The product from step (42e) was
10   reacted in the manner of step (38i). ^{1}H NMR (500 MHz, CHLOROFORM-D) δ ppm 7.42 (d, J=9.16 Hz, 1 H), 7.17 (m, 3 H), 6.69 (m, 11 H), 6.29 (s, 1 H), 3.79 (m, 8 H), 3.57 (dd, J=9.61, 5.34 Hz, 1 H), 3.31 (m, 1 H), 3.16 (q, J=8.55 Hz, 1 H), 2.96 (dd, J=14.19, 3.51 Hz, 1 H),
15   2.70 (m, 2 H), 2.47 (m, 4 H), 2.18 (m, 2 H), 2.03 (s, 3 H), 1.70 (dd, J=14.04, 5.49 Hz, 2 H), 1.47 (dd, J=14.04, 7.93 Hz, 2 H), 0.77 (m, 1 H), 0.60 (m, 2 H), 0.19 (m, 2 H). LC-MS (Method B, retention time: 2.167 min), MS m/z 707 (M^++1).

20

BIOLOGICAL METHODS

There are a number of methods by which inhibitors of the BACE enzyme can be identified experimentally. The enzyme can be obtained from membrane samples from
25   natural tissues or cultured cells or can be expressed recombinantly in a host cell by well known methods of molecular biology. The whole enzyme or a portion thereof can be expressed, for example, in bacterial, insect or mammalian cells to obtain a catalytically
30   active enzyme species. The enzymatic activity and/or ligand binding capability of the enzyme can be assessed within these membrane samples, or the enzyme can be purified to varying extents. As an

illustrative example, the nucleic acid sequence
encoding the pro and catalytic domains of human BACE
can be appended on the 5' end with an untranslated and
signal sequence from the gene for

5   acetylcholinesterase, and on the 3' end with a
sequence encoding a poly-histidine tag. This cDNA can
then be expressed in Drosophila melanogaster S2 cells
in which the signal and pro sequences of the
transcribed/translated protein are removed by cellular

10 proteases and the catalytic domain, appended by a C-
terminal poly-histidine tag, is secreted out into the
cellular medium. The enzyme can then be purified from
the culture medium by nickel affinity chromatography
by methods well known to those trained in the art

15 [Mallender, W. *et al.*, (2001) "Characterization of
recombinant, soluble beta-secretase from an insect
cell expression system." *Mol. Pharmacol.* **59**: 619-626].
Similar strategies for expressing and purifying
various forms of BACE in bacterial, mammalian and

20 other cell types would be known to one skilled in the
art.

The enzyme thus obtained can be put in contact
with a test compound and with an appropriate substrate
on which enzyme-mediated peptide bond hydrolysis is

25 known to occur. The ability of the test compound to
diminish the rate of substrate hydrolysis can then be
quantified as a measure of inhibition potency.
Appropriate substrates can be prepared as peptides,
proteins, or chemically modified versions of peptides

30 or proteins, that contain an amino acid sequence that
is recognized as a substrate by the enzyme. For
example, the amino acid sequence immediately proximal
to the beta-cleavage site within Swedish mutant APP is

known to be recognized as a substrate for BACE. It
has been demonstrated that the amino acid sequence X-
EVNLDAEFK(Y), (SEQ. ID. NO.:1), in which X is a
chemical group appended to the N-terminus of the

5   peptide and Y is a chemical group appended to the
epsilon amino group of the C-terminal lysine side
chain, is efficiently cleaved by BACE at the peptide
bond between the L and D residues. Longer peptide and
protein substrates can also be designed by extending

10   the amino acid composition on the N-terminal, the C-
terminal, or both ends.

In one known application of (SEQ. ID. NO.:1), X
is a 7-methoxycoumarin-4-acetyl (MCA) group and Y is a
dinitrophenyl (DNP) group [Marcinkeviciene, J. *et al.*,

15   (2001) "Mechanism of inhibition of beta-site amyloid
precursor protein-cleaving enzyme (BACE) by a statine-
based peptide." *J. Biol. Chem.* **276**: 23790-23794].
When this peptide is intact, the natural fluorescence
of the MCA group is quenched by its proximity to the

20   DNP group. Upon enzyme-mediated hydrolysis, the MCA
and DNP groups are separated onto different peptide
fragments and the fluorescence of the MCA group is
thus revealed. The increase in fluorescence intensity
that accompanies enzyme-mediated peptide hydrolysis

25   can be measured as a function of time to quantify the
velocity of enzyme catalysis. In a typical assay
system the BACE enzyme is diluted into a buffer system
composed of 50 mM acetate, pH 4.5 containing 0.25
mg/ml bovine serum albumin. To this is added neat

30   dimethyl sulfoxide (DMSO) or a stock solution of test
compound dissolved in DMSO so that the final amount of
DMSO in all assays is held constant at 2.5% (v:v).
The enzymatic reaction is then initiated by addition

of a stock solution of the substrate peptide to a
known concentration. The fluorescence increase with
time after substrate addition is monitored with an
appropriate fluorescence detection instrument, such as
5 a microplate reader or spectrofluorometer. The slope
of the signal vs. time plot (progress curve) for
samples to which only DMSO was added is taken as a
measure of the uninhibited velocity and represents
100% enzymatic activity. The diminution of velocity
10 that is observed at a known concentration of test
compound is used to define the % inhibition of
activity as follows: % inhibition = $100*(1-(vi/v0))$
where vi is the velocity in the presence of test
compound at a known concentration and v0 is the
15 velocity of the uninhibited enzyme. A compound is
considered active in this assay if its IC_{50} is less
than 50 μM. Activity of example compounds of the
invention is provided in Table 1, wherein +++ denotes
activity of 0.1 μM or greater potency, ++ denotes
20 potency in the range of 0.1 to 1.0 μM and + denotes
potency in the range between 1 μM and 50 μM.

Table 1

Compounds of Example	Activity Rating[a]
1	+++
2	+++
3	+++
4	++
5	+
6	+
7	+++

Compounds of Example	Activity Rating[a]
8	+++
9	+++
10	+++
11	++
12	++
13	+
14	+++
15	+
16	+++
17	++
18	+++
19	++
20	++
21	++
22	++
23	+
24	+++
25	+++
26	+
27	+
28	+++
29	+++
30	+++
31	++
32	++
33	+++
34	+++
35	+++
36	+++
37	+++
38	+++

Compounds of Example	Activity Rating[a]
39	+++
40	+++
41	+++
42	+++

[a]Activity based on IC_{50} values:

+++ = <0.1 μM

++ = 0.1 - 1.0 μM

+ = 1.0 - 50 μM

In an alternative application using the amino
acid sequence X-EVNLDAEFK(Y), X is an acetyl group
and Y is DNP. This peptide is put in contact with
BACE and the test compound in a fashion similar to
that described above. After a fixed period of time
the enzymatic reaction is stopped by denaturing the
enzyme by addition of a known amount of
trifluoroacetic acid (TFA) or by heating the sample in
a boiling water bath for 5 min. The sample is then
loaded onto a C18 or other appropriate reverse phase
HPLC column and the substrate peptide is separated
from the product peptide fragments by isocratic or
gradient elution methods well known to those trained
in the art. The substrate and the C-terminal product
peptide fragment can be identified by the absorbance
at ca. 350 nm imparted by the DNP group. The area
under the C-terminal product peak can be quantified as
a measure of enzyme activity and the diminution of
this activity can be used to define the inhibition
potency of test compounds as described above. A

compound is considered active in this assay if its IC_{50} is less than 50μM.

<u>In vitro assay to identify β-secretase inhibitor based</u>
<u>on the inhibition of Aβ formation from membrane</u>
<u>preparations.</u>

     An isolated membrane fraction which contains functionally active β-secretase and β-APP substrates can generate β-secretase cleavage products including Aβ (Roberts, S.B.; Hendrick, J. P.; Vinitsky, A.; Lewis, M.; Smith, D.W.; Pak, R. PCT Publication WO 01/0175435; Fechteler, K.; Kostka, M.; Fuchs, M. Patent Application No. DE 99-19941039; Shearman, M.; Beher, D. <u>et al.</u>,, <i>Biochemistry,</i> <u>2000</u>, 39, 8698-8704; Zhang, L. Song, L. <u>et al.</u>, <i>Biochemistry</i> <u>2001</u>, 40, 5049-5055). An isolated membrane fraction can be prepared from human derived cell lines such as HeLa and H4 which have been transfected with wild type or mutant forms of β-APP or a human alkaline phosphatase β-APP fusion construct, and stably express high levels of β-secretase substrates. The endogenous β-secretase present in the isolated membranes prepared at 0-4 °C cleaves the β-APP substrates when the membranes are shifted from 0-4 to 37 °C. Detection of the cleavage products including Aβ can be monitored by standard techniques such as immunoprecipitation (Citron, M.; Diehl, T.S. <u>et al.</u>, <i>Proc. Natl. Acad. Sci. USA,</i> <u>1996</u>, 93,13170-13175), western blot (Klafki, H.-W.; Ambramowski, D. <u>et al.</u>, <i>J. Biol. Chem..</i> <u>1996</u>, 271, 28655-28659), enzyme linked immunosorbent assay (ELISA) as demonstrated by Seubert, P.; Vigo-Pelfrey,

C. _et al._, _Nature,_ 1992, **359**, 325-327, or by a
preferred method using time-resolved fluorescence of
the homogeneous sample containing membranes and Aβ
(Roberts, S.B.; Hendrick, J. P.; Vinitsky, A.; Lewis,
5   M.; Smith, D.W.; Pak, R. PCT Publication WO
01/0175435; Shearman, M.; Beher, D. _et al._,
Biochemistry, 2000, **39**, 8698-8704). The Aβ present in
a homogeneous sample containing membranes can be
detected by time-resolved fluorescence with two
10   antibodies that recognize different epitopes of Aβ.
One of the antibodies recognizes an epitope that is
present in Aβ but not present in the precursor
fragments; preferably the antibody binds the carboxyl
terminus of Aβ generated by the β-secretase cleavage.
15   The second antibody binds to any other epitope present
on Aβ. For example, antibodies that bind the N-
terminal region (e.g., 26D6-B2-B3 [®] SIBIA
Neurosciences, La Jolla, CA) or bind the C-terminal
end (e.g., 9S3.2 [®] antibody, Biosolutions, Newark, DE)
20   of the Aβ peptide are known. The antibodies are
labeled with a pair of fluorescent adducts that
transfer fluorescent energy when the adducts are
brought in close proximity as a result of binding to
the N- and C-terminal ends or regions of Aβ. A lack
25   of fluorescence is indicative of the absence of
cleavage products, resulting from inhibition of β-
secretase. The isolated membrane assay can be used to
identify candidate agents that inhibit the activity of
β-secretase cleavage and Aβ production.

30   A typical membrane-based assay requires 45 μg
membrane protein per well in a 96- or 384-well format.

Membranes in a neutral buffer are combined with the
test compound and shifted from 0-4 to 37 °C. Test
agents may typically consist of synthetic compounds,
secondary metabolites from bacterial or fungal
5   fermentation extracts, or extracts from plant or
marine samples. All synthetic agents are initially
screened at doses ranging from 10-100 μM or in the
case of extracts at sufficient dilution to minimize
cytotoxicity. Incubation of the membranes with the
10   test agent will continue for approximately 90 minutes
at which time fluorescence labeled antibodies are
added to each well for Aβ quantitation. The time-
resolved fluorescence detection and quantitation of Aβ
is described elsewhere (Roberts, S.B.; Hendrick, J.
15   P.; Vinitsky, A.; Lewis, M.; Smith, D.W.; Pak, R. PCT
Publication WO 01/0175435; Shearman, M.; Beher, D. _et_
_al._, _Biochemistry_, 2000. 39, 8698-8704). Results are
obtained by analysis of the plate in a fluorescence
plate reader and comparison to the mock treated
20   membranes and samples in which known amounts of Aβ
were added to construct a standard concentration
curve. A positive acting compound is one that
inhibits the Aβ relative to the control sample by at
least 50% at the initial tested concentration.
25   Compounds of the present invention are considered
active when tested in the above assay if the IC_{50} value
for the test compound is less than 50 μM. A preferred
IC_{50} value is less than 1 μM. A more preferred IC_{50}
value is less than 0.1 μM. If a compound is found to
30   be active then a dose response experiment is performed
to determine the lowest dose of compound necessary to
elicit the inhibition of the production of Aβ.

In Vivo Assays for the determination of Aβ reduction
by a β-secretase inhibitor.

    In vivo assays are available to demonstrate the
inhibition of β-secretase activity. In these assays,

5   animals, such as mice, that express normal levels of
APP, β- and γ-secretase or are engineered to express
higher levels of APP and hence Aβ can be used to
demonstrate the utility of β-secretase inhibitors, as
demonstrated with γ-secretase inhibitors [Dovey, H. et

10 al., (2001), J. Neurochem. 76: 173-181]. In these
assays, β-secretase inhibitors are administered to
animals and Aβ levels in multiple compartments, such
as plasma, cerebral spinal fluid, and brain extracts,
are monitored for Aβ levels using methods previously

15 outlined. For instance, Tg2576 mice, which
overexpress human APP, are administered β-secretase
inhibitors by oral gavage at doses that will cause
measurable Aβ lowering, typically less than 100 mg/kg.
Three hours after dosing plasma, brain, and CSF are

20 collected, frozen in liquid nitrogen, and stored at –
80° C until analysis. For Aβ detection, plasma is
diluted 15-fold in PBS with 0.1% Chaps while CSF is
diluted 15-fold in 1% Chaps with protease inhibitors
(5 µg/ml leupeptin, 30 µg/ml aprotinin, 1 mM

25 phenylmethylsulfonylfluoride, 1 µM pepstatin). Brains
are homogenized in 1% Chaps with protease inhibitors
using 24 ml solution/g brain tissue. Homogenates were
then centrifuged at 100,000 x g for 1 hr at 4° C. The
resulting supernatants were then diluted 10-fold in 1%

30 Chaps with protease inhibitors. Aβ levels in the
plasma, CSF, and brain lysate can then be measured

using time-resolved fluorescence of the homogenous
sample or one of the other methods previously
described.

A β-secretase inhibitor is considered active in
one of the above *in vivo* assays if it reduces Aβ by at
least 50% at a dosage of 100mg/kg.

All references cited herein are hereby
incorporated in their entirety herein by reference.

DOSAGE AND FORMULATION

The compounds of the present invention can be
administered orally using any pharmaceutically
acceptable dosage form known in the art for such
administration. The active ingredient can be supplied
in solid dosage forms such as dry powders, granules,
tablets or capsules, or in liquid dosage forms, such
as syrups or aqueous suspensions. The active
ingredient can be administered alone, but is generally
administered with a pharmaceutical carrier. A
valuable treatise with respect to pharmaceutical
dosage forms is Remington's Pharmaceutical Sciences,
Mack Publishing.

The compounds of the present invention can be
administered in such oral dosage forms as tablets,
capsules (each of which includes sustained release or
timed release formulations), pills, powders, granules,
elixirs, tinctures, suspensions, syrups, and
emulsions. Likewise, they may also be administered in
intravenous (bolus or infusion), intraperitoneal,
subcutaneous, or intramuscular form, all using dosage
forms well known to those of ordinary skill in the
pharmaceutical arts. An effective but non-toxic

amount of the compound desired can be employed to
prevent or treat neurological disorders related to β-
amyloid production or accumulation, such as
Alzheimer's disease and Down's Syndrome.

5   The compounds of this invention can be
administered by any means that produces contact of the
active agent with the agent's site of action in the
body of a host, such as a human or a mammal., They
can be administered by any conventional means

10   available for use in conjunction with pharmaceuticals,
either as individual therapeutic agents or in a
combination of therapeutic agents. They can be
administered alone, but generally administered with a
pharmaceutical carrier selected on the basis of the

15   chosen route of administration and standard
pharmaceutical practice.

The dosage regimen for the compounds of the
present invention will, of course, vary depending upon
known factors, such as the pharmacodynamic

20   characteristics of the particular agent and its mode
and route of administration; the species, age, sex,
health, medical condition, and weight of the
recipient; the nature and extent of the symptoms; the
kind of concurrent treatment; the frequency of

25   treatment; the route of administration, the renal and
hepatic function of the patient, and the effect
desired. An ordinarily skilled physician or
veterinarian can readily determine and prescribe the
effective amount of the drug required to prevent,

30   counter, or arrest the progress of the condition.

Advantageously, compounds of the present
invention may be administered in a single daily dose,

or the total daily dosage may be administered in divided doses of two, three, or four times daily.

The compounds for the present invention can be administered in intranasal form via topical use of

5   suitable intranasal vehicles, or via transdermal routes, using those forms of transdermal skin patches well known to those of ordinary skill in that art. To be administered in the form of a transdermal delivery system, the dosage administration will, of course, be

10 continuous rather than intermittant throughout the dosage regimen.

In the methods of the present invention, the compounds herein described in detail can form the active ingredient, and are typically administered in

15 admixture with suitable pharmaceutical diluents, excipients, or carriers (collectively referred to herein as carrier materials) suitably selected with respect to the intended form of administration, that is, oral tablets, capsules, elixirs, syrups and the

20 like, and consistent with conventional pharmaceutical practices.

For instance, for oral administration in the form of a tablet or capsule, the active drug component can be combined with an oral, non-toxic, pharmaceutically

25 acceptable, inert carrier such as lactose, starch, sucrose, glucose, methyl callulose, magnesium stearate, dicalcium phosphate, calcium sulfate, mannitol, sorbitol and the like; for oral administration in liquid form, the oral drug

30 components can be combined with any oral, non-toxic, pharmaceutically acceptable inert carrier such as ethanol, glycerol, water, and the like. Moreover, when desired or necessary, suitable binders,

lubricants, disintegrating agents, and coloring agents
can also be incorporated into the mixture. Suitable
binders include starch, gelatin, natural sugars such
as glucose or β-lactose, corn sweeteners, natural and
5   synthetic gums such as acacia, tragacanth, or sodium
alginate, carboxymethylcellulose, polyethylene glycol,
waxes, and the like. Lubricants used in these dosage
forms include sodium oleate, sodium stearate,
magnesium stearate, sodium benzoate, sodium acetate,
10   sodium chloride, and the like. Disintegrators
include, without limitation, starch, methyl cellulose,
agar, bentonite, xanthan gum, and the like.

The compounds of the present invention can also
be administered in the form of liposome delivery
15   systems, such as small unilamellar vesicles, large
unilamallar vesicles, and multilamellar vesicles.
Liposomes can be formed from a variety of
phospholipids, such as cholesterol, stearylamine, or
phosphatidylcholines.
20   Compounds of the present invention may also be
coupled with soluble polymers as targetable drug
carriers. Such polymers can include
polyvinylpyrrolidone, pyran copolymer,
polyhydroxypropylmethacrylamide-phenol,
25   polyhydroxyethylaspartamidephenol, or
polyethyleneoxide-polylysine substituted with
palmitoyl residues. Furthermore, the compounds of the
present invention may be coupled to a class of
biodegradable polymers useful in achieving controlled
30   release of a drug, for example, polylactic acid,
polyglycolic acid, copolymers of polylactic and
polyglycolic acid, polyepsilon caprolactone,
polyhydroxy butyric acid, polyorthoesters,

polyacetals, polydihydropyrans, polycyanoacylates, and crosslinked or amphipathic block copolymers of hydrogels.

Gelatin capsules may contain the active

5 ingredient and powdered carriers, such as lactose, starch, cellulose derivatives, magnesium stearate, stearic acid, and the like. Similar diluents can be used to make compressed tablets. Both tablets and capsules can be manufactured as sustained release

10 products to provide for continuous release of medication over a period of hours. Compressed tablets can be sugar coated or film coated to mask any unpleasant taste and protect the tablet from the atmosphere, or enteric coated for selective

15 disintegration in the gastrointestinal tract. Liquid dosage forms for oral administration can contain coloring and flavoring to increase patient acceptance.

In general, water, a suitable oil, saline,

20 aqueous dextrose (glucose), and related sugar solutions and glycols such as propylene glycol or polyethylene glycols are suitable carriers for parenteral solutions. Solutions for parenteral administration preferably contain a water soluble salt

25 of the active ingredient, suitable stabilizing agents, and if necessary, buffer substances. Antioxidizing agents such as sodium bisulfite, sodium sulfite, or ascorbic acid, either alone or combined, are suitable stabilizing agents. Also used are citric acid and its

30 salts and sodium EDTA. In addition, parenteral solutions can contain preservatives, such as benzalkonium chloride, methyl- or propyl-paraben, and chlorobutanol.

Suitable pharmaceutical carriers are described in Remington's Pharmaceutical Sciences, Mack Publishing Company, a standard reference text in this field.